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An Annotated Checklist of the Ground Beetles (Coleoptera: Carabidae) of Mexico, Including Some Nomenclatural Changes

Lista Taxonómica Anotada de Escarabajos Carábidos (Coleoptera: Carabidae) de México, Incluyendo Algunos Cambios Nomenclaturales

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Dedication

*Dedicated to the memory of George E. Ball and Terry L. Erwin,
both of whom were exceptional carabidologists*

Dedicatoria

*Dedicado a la memoria de George E. Ball y Terry L. Erwin,
ambos quienes en vida fueron especialistas excepcionales en carábidos*

ABSTRACT

The first comprehensive checklist of the ground beetles in the family Carabidae is presented for Mexico with specimen records to the state level with sources. Data from over 69,300 dataset records as well as published records is presented in this checklist. A total of 1,634 described carabid species in 45 tribes and 179 genera are recorded from Mexico; 1,030 of those species were described from type material collected only in Mexico. Morphospecies (undescribed species) are not included in this checklist. The number of carabid species recorded from Mexico could potentially be as high as 2,074 if morphospecies were included. The sources of the state records are included for each of the species. This checklist also includes type locality information for over 200 species known only from type material. The name *Lebia interrupta* Chaudoir, 1870: 210 is given **new status** as a valid species. Three **new synonymies** are proposed, with the senior synonym of each combination listed first: *Chlaenius forreri* Bates, 1884 = *Chlaenius pimalicus* Casey, 1914; *Lebia cymindoides* Bates, 1883 = *Lebia scapula* G. Horn, 1885; and *Lebia interrupta* Chaudoir, 1870 = *Lebia tschitscherini* Lutshnik, 1922.

Key words: checklist, Carabidae, new status, synonymies, Mexico

RESUMEN

La presente publicación científica documenta la primera lista taxonómica exhaustiva de escarabajos carábidos (familia Carabidae) de México, con registros de especímenes a nivel estatal y sus respectivas fuentes. La lista incluye datos provenientes de más de 69,300 registros de bases de datos, así como registros publicados. Se registran en México un total de 1,634 especies de escarabajos carábidos, pertenecientes a 45 tribus y 179 géneros; 1,030 especies fueron descritas de material tipo recolectado únicamente en México. No se incluyen morfoespecies (especies no descritas) en la lista. El número de especies de carábidos registradas en México podría llegar a ser de hasta 2,074 si se incluyeran las morfoespecies. Para cada especie se indica la fuente del registro estatal. Esta lista también proporciona información sobre las localidades tipo para más de 200 especies conocidas únicamente a partir de este tipo de material. El nombre *Lebia interrupta* Chaudoir, 1870: 210 es elevado a **nuevo estatus** válido como especie. Se proponen tres nuevas sinonimias, listando primero el sinónimo sénior en cada caso: *Chlaenius forreri* Bates, 1884 = *Chlaenius pimalicus* Casey, 1914; *Lebia cymindoides* Bates, 1883 = *Lebia scapula* G. Horn, 1885; y *Lebia interrupta* Chaudoir, 1870 = *Lebia tschitscherini* Lutshnik, 1922.

Palabra Clave: lista taxonómica, Carabidae, nuevo estatus, sinonimias, México.

The carabid (ground beetle) fauna of Mexico has been included in part by many individual publications over the years. Csiki (1927-1933) included the Mexican carabid species with the world fauna in his catalogues. Blackwelder (1944) and Erwin et al. (1977) included the Mexican species in their respective western hemisphere catalogues. Blackwelder (1944) had a total of 1,009 species from Mexico, and Erwin et al. (1977) had 977 species from Mexico. The Carabidae de México webpage (Barriaga-Tunon, J. E.

2008) appears to be somewhat incomplete, as it only lists 731 described species. Ball and Shpeley (2000: 381) stated that the carabid fauna of Mexico numbered 1,957 species in 171 genera. Printout of an Excel spreadsheet dated June 19, 1997 is marked as Terry L. Erwin's preliminary *Checklist of the Carabidae of the Western Hemisphere* (Ball and Shpeley 2000: 387). This *Checklist of the Carabidae of the Western Hemisphere* (Erwin, 1995) was the source of the rather high number of Mexican carabid species compared

to that of previous authors. However, careful examination of this list confirmed there were only 166 genera and the 1,957 species were actually comprised of 1,267 described species, 473 morphospecies (undescribed species) and 217 species which were now excluded (typos, duplicates, synonymies, no records from Mexico, etc.). Unfortunately, Erwin's preliminary Checklist was taken at face value, and I accept responsibility for not vetting the checklist prior to the Ball and Shpeley publication in the year 2000. To sum up, the number of described Mexican species at the time of publication in the year 2000 would be 1,267. If the 473 morphospecies were included in the count, that would bring the potential number of Mexican carabid species to 1,740, not 1,957, the number on page 381 in the 2000 publication. Recently, Cerón-Gómez et al. (2022: 2; 2025b: 45) stated that the potential species richness of carabids in Mexico was 2,012 species. Just as Blackwelder (1944) and Erwin et al. (1977) included species which they believed might one day be found in Mexico, this checklist also includes 69 species for which there are currently no specimens recorded from Mexico. Therefore, these 69 species are not included in the species count for Mexico carabids.

MATERIALS

This manuscript is partly based on a preliminary *Checklist of the Carabidae of the Western Hemisphere* (Erwin, 1995) which Terry L. Erwin shared with George E. Ball and I around 1997. The Mexican portion of Erwin's checklist was based on Mexican carabid species housed in the E.H. Strickland Museum (UASM). Since then, the list has been updated with specimen records from several institutional collections as well as published records and web site records. The E.H. Strickland Entomological Museum has over 70,800 carabids which were collected in Mexico, nearly all of which are digitized (University of Alberta Museums, 2025); over 50,800 specimens are determined to the species or subspecies level, and the remaining 20,000+ specimens are determined to genus and morphospecies, with some of those representing what are believed to be new species, or determined simply as "sp.". There are 440 Mexican morphospecies in the Strickland Museum which are considered to be new to science. Table 1 is a summary of the numbers of morphospecies in their 37 respective genera. Morphospecies are not included in the species listing of this checklist, but are included with the described species to give an indication of potential species richness for Mexico. The Global Biodiversity Information Facility web site (GBIF, 2022) had 10,330 records of described species of carabid specimens with geographic coordinates which were downloaded (GBIF, 2023). The GBIF download included 1,381 Strickland Museum specimen records and 623 IBUNAM specimen records. The GBIF specimen records added many state records as well as additional species to this checklist. More than 5,700 digital specimen records of described species were accessed on the Universidad Nacional Autónoma de México website (IBUNAM, 2025). Over 2,200 digital specimen records of described species were received from the California Academy of Sciences Collection (CASC) and over 1,000 records from the Uni-

versity of Arizona Insect Collection (UAIC). There were 696 institutional records of described species hosted on the Strickland Museum site. Another 667 records of described species were received from the Snow Entomological Collection (SEMC). Some of the records also came from publications, dating from 1833 to 2025. See Table 2 for a listing of all the sources. All species determinations were taken as being correct and no attempt was made to check all of the identifications of over 69,300 specimens. However, a few determinations which appear to be incorrect are noted in the checklist. Many of the locality place names in Bates' publication were verified to state using the gazetteer published by Selander and Vaurie (1962). The Mexican travel guide *Caminos de Mexico* (Guia Goodrich-Euzkadi, 1964) also provided the correct state for a number of place names.

TERMS

The term "species" is applied to both species and subspecies similar to previous authors: Cerón-Gómez et al. (2022: 1); Zaragoza-Caballero et al. (2019: 16, 18-25); and Burgos-Solorio and Trejo-Loyo (2001: 72, 81-82). For example, a species may have three subspecies – the nominate subspecies (*s. str.*), and two subspecies (*s. lat.*). In this checklist, the count for the example in the preceding sentence would be three species. The term "morphospecies" is applied to specimens which are morphologically different from other specimens, whether described or undescribed, in a given genus, and may be assigned a number or letter designation for ease of sorting. Morphospecies can be new to science awaiting description, or simply undetermined due to lack of literature and access to type material for comparison. "Potential species richness" is expressed by the sum of the number of described species plus the number of undescribed species (morphospecies).

CHECKLIST OF CARABIDAE OF MEXICO

This checklist follows the names and arrangement of tribes as published by Bousquet (2012). Genera, species and subspecies (nominate listed first) are arranged alphabetically for ease of location in the checklist. In this checklist, the tiger beetles are included as members of the family Carabidae (Bousquet 2012). The scientific names of tiger beetles have been updated to the list published by Wiesner (2020). Scientific species names were determined to be either valid or invalid (synonyms) by using the online resource of Lorenz whose website was incorporated into GBIF (2022 - 2025) and the Catalogue of Life (2025). Additionally, Lorenz 2005 and Wiesner 2020 were used to confirm validity of scientific names. See Table 3 for standard abbreviations used in this checklist for each of the 32 states in Mexico. Some older specimens have only Mexico on their collection label. Some of the early species descriptions also had only Mexico for the type locality, no other details as to where the specimen was collected. Both of these broad locality instances appear in the checklist as "Mexico", not to be confused with the abbreviation MEX for the state of Mexico. Each state record is followed by a superscript number or numbers, which represent the source or sources of that record or those records. See Table 3 for

Table 1. Summary of the 436 Strickland Museum morphospecies, with number of morphospecies in their corresponding genera.

Genus	# Morphospecies	Genus	# Morphospecies	Genus	# Morphospecies
<i>Odontocheila</i>	1	<i>Pseudabarys</i>	5	<i>Ega</i>	2
<i>Tetracha</i>	2	<i>Amara</i>	5	<i>Euphorticus</i>	1
<i>Ardistomis</i>	8	<i>Badister</i>	1	<i>Lachnophorus</i>	4
<i>Clivina</i>	16	<i>Aztecarpalus</i>	1	<i>Pentagonica</i>	2
<i>Oxydrepanus</i>	1	<i>Bradycellus</i>	8	<i>Leptotrachelus</i>	3
<i>Dyschirius</i>	3	<i>Harpalus</i>	3	<i>Apenes</i>	17
<i>Bembidion</i>	35	<i>Selenophorus</i>	47	<i>Calleida</i>	8
<i>Elaphropus</i>	2	<i>Stenolophus</i>	2	<i>Cymindis</i>	9
<i>Polyderis</i>	1	<i>Dyscolus/Platynus</i>	149	<i>Lebia</i>	68
<i>Tachys</i>	2	<i>Elliptoleus</i>	2	<i>Plochionus</i>	1
<i>Pachyteles</i>	8	<i>Glyptolenus</i>	3	<i>Helluomorphoides</i>	5
<i>Oxycrepis</i>	1	<i>Anchonoderus</i>	8		
<i>Abaris</i>	3	<i>Eucaerus</i>	3		

Table 2. Superscript numbers and source of data that each represents.

¹ Erwin, T.L.: original records

² UASM website records^A

³ GBIF website records^B

⁴ IBUNAM website records^C

⁵ CASC records^D

⁶ UAIC website records^E

⁷ SEMC records^F

⁸ FMNH records^G

⁹ Allen, R.T., 1972

¹⁰ Allen, R.T. & G.E. Ball, 1980

¹¹ Anderson, R.S., per. comm. 7.09.2023

¹² Ball, G.E. & D.R. Maddison, 1987

¹³ Barr, T.C., Jr., 1982a

¹⁴ Barr, T.C., Jr., 1982b

¹⁵ Bates, H.W., 1881

¹⁶ Bates, H.W., 1882

¹⁷ Bates, H.W., 1883

¹⁸ Bates, H.W., 1884

¹⁹ Bates, H.W., 1891

²⁰ Bousquet, Y., 1996

²¹ Bousquet, Y., 2012

²² Casale, A., 2008

²³ Cazier, M.A., 1954

²⁴ Cerón-Gómez, R. et al., 2022

²⁵ Cerón-Gómez, R. et al., 2024

²⁶ Cerón-Gómez, R. & J. Marquez, 2025a

²⁷ Cerón-Gómez, R. et al., 2025b

²⁸ Davidson, R.L., 1980

²⁹ Erwin, T.L., 1970

³⁰ Erwin, T.L., 1974

^A University of Alberta E.H. Strickland Entomological Museum (UASM), University of Alberta, Edmonton, Alberta T6G 2E9, Canada: <https://search.museums.ualberta.ca>.

^B Global Biodiversity Information Facility: <https://www.gbif.org>.

^C Universidad Nacional Autónoma de México (UNAM), Instituto de Biología: <https://datosabiertos.unam.mx/biodiversidad/>.

^D California Academy of Sciences Collection (CASC), Golden Gate Park, San Francisco, California: <https://doi.org/10.15468/3ibpmo>.

^E University of Arizona Insect Collection (UAIC), Tucson, Arizona: <https://ecdysis.org/collections/listtabledisplay.php?db=73&country=Mexico&taxa=Carabidae&usethes=1&taxontype=2>.

^F Snow Entomological Museum (SEMC), University of Kansas, Lawrence, Kansas.

^G Field Museum of Natural History (FMNH), Chicago, Illinois.

³¹ Erwin, T.L., 2000

³² Frania, H.E. & G.E. Ball, 2007

³³ Gidaspow, T. 1959

³⁴ Gidaspow, T. 1968

³⁵ Goulet, H., 1974

³⁶ Hilchie, G.R. & G.E. Ball, 2024

³⁷ Hunting, W.M., 2013

³⁸ Lassalle, B., 2009

³⁹ Lassalle, B. & Van Den Berghe, E., 2010

⁴⁰ Lassalle, B. & Van Den Berghe, E., 2011a

⁴¹ Lassalle, B. & Van Den Berghe, E., 2011b

⁴² Lassalle, B. & Van Den Berghe, E., 2013

⁴³ Liebherr, J.K., 1989

⁴⁴ Liebherr, J.K., 1991

⁴⁵ Liebherr, J.K., 1994

⁴⁶ Mateu, J., 1974a

⁴⁷ Mateu, J., 1974b

⁴⁸ Messer P.W. & B.T. Raber, 2021

⁴⁹ Messer P.W. & B.T. Raber, 2025

⁵⁰ Montes de Oca et al., 2014

⁵¹ Palacios-Vargas, J.G. et al., 2014-2015

⁵² Ramírez-Hernández, A. et al., 2018

⁵³ Reddell, J.R., 1982

⁵⁴ Shpeley, D. & T.R. Van Devender, 2022

⁵⁵ Whitehead, D.R., 1972

⁵⁶ Wiesner, J., 2020

⁵⁷ Will, K.W., pers. comm. 30.08.2023

⁵⁸ Will, K.W., 2002

⁵⁹ type locality state in original description

the superscript numbers which appear in this checklist and corresponding sources of data for each of the superscript numbers. Additional sources are listed with a reference after the state record.

Most of the original Mexican state entries (designated by a superscript 1, i.e. ¹) in the preliminary Checklist from Terry L. Erwin appear to be based on published or prepublication locality records. For example, the tiger beetles are treated in Erwin and Pearsons's Caraboidea of the Western Hemisphere (Erwin and Pearson, 2008), and nearly all of the state records for the tiger beetles of this checklist have a superscript 1. The same is evident in the genera *Coptodera* and *Onypterygia*, where nearly all of the states for each of the species has a superscript 1, which would have been based on the papers by Shpeley and Ball (1993) and Whitehead and Ball (1997), respectively. Where the publication is known, the reference is given at the beginning of that genus. Other state entries (designated by a superscript 1, i.e. ¹) without recorded specimens, appear to be ones which T.L. Erwin most likely believed the species could potentially occur in that Mexican state.

In this checklist, species which were described from type specimens collected only in Mexico are represented by a bold font checklist species number, i.e., **0001**. Species which type specimens collected in Mexico plus an additional country or multiple countries, as well as species

with non-Mexico collected type specimens, are represented with a regular font checklist number, i.e., 0003. Within this checklist, there are 19 species recorded from Mexico which are suspect, and each is indicated as "most likely mis-identified or mis-labelled". Also, there are 65 species noted with a single asterisk following the species number, i.e., 0003*. These are species which were not described from Mexico, nor are there any confirmed specimen records from Mexico, even though some will have a state abbreviation followed by a superscript 1 indicating the possibility of them being found in Mexico one day. Lastly, there are 204 species noted with a superscript number sign following the species number, i.e., 0025#. These are species which were described from specimens collected in Mexico, but no additional specimens have been recorded for those species since the publication of their scientific names. Species described up to and including 2025 are included in this count. When the type locality is given, the page number in the reference is the page on which the type locality is printed, which may be different from the page number on which the species description begins. It is also possible that there are specimens of some of these species in the hundreds to tens of thousands of unsorted and undetermined carabid specimens housed in collections. The Strickland Museum currently houses over 13,000 specimens collected in Mexico which are only determined to genus.

Table 3. Abbreviations for states in Mexico in which carabids were collected.

Aguascalientes	AGS	Guerrero	GRO	Quintana Roo	QROO
Baja California Norte	BCN	Hidalgo	HGO	San Luis Potosí	SLP
Baja California Sur	BCS	Jalisco	JAL	Sinaloa	SIN
Campeche	CAMP	México (state)	MEX	Sonora	SON
Chiapas	CHIS	Michoacán	MICH	Tabasco	TAB
Chihuahua	CHIH	Morelos	MOR	Tamaulipas	TAMPS
Ciudad de México	CDMX	Nayarit	NAY	Tlaxcala	TLAX
Coahuila	COAH	Nuevo León	NL	Veracruz	VER
Colima	COL	Oaxaca	OAX	Yucatán	YUC
Durango	DGO	Puebla	PUE	Zacatecas	ZAC
Guanajuato	GTO	Querétaro	QRO		

COLEOPTERA
Carabidae

01. Clinidiini

Note: Erwin's superscript ¹ for *Clinidium* species refers to Bell and Bell 1985 and with a superscript ^H refers to Erwin (2011a)

0001. *Clinidium balli* Bell and Bell, 1985
HGO^{1,2,7}, SLP^{1,3,8}
0002. *Clinidium blomi* Bell, 1970
CHIS^{1,2,5}, OAX²
- 0003*. *Clinidium calcaratum* LeConte, 1875
Type locality: "Vancouver Island" [British Columbia] and "Oregon", LeConte 1875: 164; "Mexico"¹; "probably Baja California", Erwin 2011a: 249
- 0004*. *Clinidium extrarium* Bell and Bell, 1978
Type locality: "Am. Bor.", Bell and Bell 1978: 63; "Mexico"¹
0005. *Clinidium guatemalenum* Sharp, 1899
CHIS^H
0006. *Clinidium halfpteri* Bell and Bell, 1985
CHIS^{1,3}, VER^{1,3}
0007. *Clinidium iviei* Bell and Bell, 1985
OAX^{1,2,3,8}
0008. *Clinidium mexicanum* Chevrolat, 1873
CDMX^{5,7}, CHIS², GRO^{2,3}, HGO^{2,3,7}, JAL², MEX^{1,2,7}, MICH², MOR^{1,3,7}, OAX^{2,3,7}, PUE^{1,2,3,5,7}, SLP³, TLAX², VER^{1,2,3,5}
0009. *Clinidium newtoni* Bell and Bell, 1985
CHIS^{1,3}
0010. *Clinidium reyesi* Bell and Bell, 1987
CHIS¹ (Bell and Bell, 1987:193)
0011. *Clinidium triplehorni* Bell and Bell, 1985
HGO^{1,3,7}, SLP³

02. Notiophilini

0012. *Notiophilus chihuahuae* Casey, 1913
CHIH^{1,2}, DGO^{1,2}, SIN^{1,2}, SON^{1,2,54}
0013. *Notiophilus semiopacus* Eschscholtz, 1833
CHIH¹, SON^{1,2,54}
0014. *Notiophilus specularis* Bates, 1881
CHIH¹, COAH^{1,2}, CHIS⁵, GRO^{1,2}, HGO^{1,2}, MEX^{1,2}, MICH^{1,2}, OAX^{1,2}, PUE^{1,2}, QRO^{1,2}, SLP², TLAX^{1,2}, VER^{1,2,3,4,19}

03. Cychrini

0015. *Scaphinotus crenatus* (Motschulsky, 1859)
BCN^{26,34}
0016. *Scaphinotus macrogonus* Bates, 1891
CDMX⁴, CHIH³, DGO^{1,4,19}
0017. *Scaphinotus mexicanus* Bates, 1882; Figure 1A
CHIH², DGO^{1,2,3,4,5,16}, SON^{2,54}
0018. *Scaphinotus punctatus* (LeConte, 1859)
BCN^{1,5,26}

04. Carabini

Note: Erwin's superscript ¹ for Carabini refers to Erwin 2007.

0019. *Callisthenes affinis* (Chaudoir, 1843)
AGS⁴, CDMX^{1,33}, CHIH^{1,3,33}, COAH³, DGO^{1,33}, HGO^{1,24,33}, MEX¹, MOR³, NL^{1,4,33}, OAX^{1,3,5,33}, SIN^{3,7}, SLP¹, TAMPS^{1,33}
0020. *Callisthenes ampliator* (Bates, 1891)
CHIH^{1,19,33}, DGO^{1,19,33}, GTO^{1,19,33}, NL¹⁹, SIN⁵, SON^{1,33}, TAMPS^{1,33}, VER^{1,33}
0021. *Callisthenes morissonii* (G. Horn, 1885)
AGS⁴, DGO^{1,33}
0022. *Callisthenes semilaeve* (LeConte, 1852)
BCN^{1,3,5} (+ Guadalupe Island)
0023. *Callisthenes simplex* (LeConte 1878)
BCN⁷; "California, Arizona, Colorado and Texas" (Gidaspow 1959:272)
0024. *Calopachys blaptoides* (Putzeys, 1846)
GTO^{1,33}, OAX^{1,2,4,33}, PUE^{1,5,33}, VER^{1,33}
0025. *Calopachys omiletemium* (Bates, 1891)
CDMX⁴, GRO^{1,2,3,5,19,33}, MOR⁴, SLP⁴
- 0026#. *Calopachys viridissimus viridissimus* Haury, 1880
"México"^{1,33}, (type locality – "it must come from México", Haury 1880: 164); "Haury described it as a Mexican species" (Gidaspow 1959: 299); "?México" (Blackwelder 1944: 21); "México" (Erwin et al. 1977: 4.6); habitus image (Lassalle and Van Den Berghe 2010: 113)
- 0027#. *Calopachys viridissimus georgeballi* Lassalle and Van Den Berghe, 2010
OAX^{2,39}; (type locality: "env. Oaxaca", Lassalle and Van Den Berghe, 2010: 112); habitus image (Lassalle and Van Den Berghe 2010: 113)
0028. *Calosoma alternans* (Fabricius, 1792)
CAMP³, CHIS^{1,4}, COL⁴, DGO³, GRO⁴, HGO²⁴, JAL⁴, NAY⁴, OAX^{3,4}, PUE⁴, QROO⁴, SIN³, TAMPS², VER^{1,3,4}, YUC³
0029. *Calosoma angulatum* Chevrolat, 1834
BCN^{1,26} (Guadalupe Island), CAMP³, CDMX³, CHIH^{3,4,5}, CHIS^{1,2,6}, COAH^{3,4}, COL^{1,2,3,4,5}, GRO^{1,3,4}, GTO³, JAL^{1,2,3,4,5,7,33}, MEX^{1,33}, MICH^{1,2,3,4,7}, MOR^{1,2,3,4,7,33}, NAY^{1,2,3,4,5,6,7,33}, NL^{1,3,4,33}, OAX^{1,2,3,4}, PUE^{1,3,4,33}, QRO³, SIN^{1,2,3,4,5,7,33}, SLP^{1,2,3,4,5,7,33}, SON^{1,2,3,4,5,7,33,54}, TAMPS^{1,2,3,4,5,6,7,19,33}, VER^{1,2,3,4,5,19,33}
0030. *Calosoma anthracinum anthracinum* Dejean 1831
GRO^{1,33}, "México"³
0031. *Calosoma anthracinum microgonum* Bates, 1891
CHIH⁵, CDMX^{1,19,33}, DGO⁵, JAL^{1,33}, PUE^{1,4,33}, VER^{1,19,33}
0032. *Calosoma atrovirens atrovirens* Chaudoir, 1869
GTO⁴, HGO^{1,3,24,33}, MOR³, QRO³, SLP^{1,3,5,7}, TAMPS^{1,33}
0033. *Calosoma atrovirens explanaticolle* Bates, 1891
CDMX^{1,3}, DGO², GTO^{1,2,19}, JAL^{1,2,3}, MEX², MICH^{1,2,3,4}, MOR^{3,4,5}, OAX¹, PUE^{1,2,4}, QRO^{1,2}, ZAC^{1,5}
- 0034#. *Calosoma atrovirens obscuroides* Lassalle and Van Den Berghe, 2013
OAX⁴² (*obscuroides* is a replacement name for *obscurum* Gehin, 1885 [nec. Letznier, 1850])
0035. *Calosoma aurocinctum* Chaudoir, 1850
CAMP^{1,2,3,4,5,33}, CHIS³, DGO^{1,2,3,33}, GTO², JAL³,

- MOR^{1,33}, NL^{3,4,5,33}, OAX^{1,33}, QRO³, QROO^{5,3}, SLP^{1,3,4,5,7,33}, TAMPS^{1,2,3,4,5,33}, VER^{1,3,5,33}, YUC^{1,2,3,4,5,33}, ZAC³
- 0036[#].** *Calosoma barksdalei* Lassalle and Van Den Berghe, 2013
JAL⁴²
- 0037.** *Calosoma bulleri* Beheim and Breuning, 1943
JAL^{1,33}, “México”³
- 0038.** *Calosoma chihuahua* Gidaspow, 1959
CHIH^{1,2,4,5,33}, DGO⁴
- 0039.** *Calosoma cicatricosum* Chaudoir, 1869
CDMX^{1,33}, MEX^{1,4,5,33}
- 0040.** *Calosoma costipenne costipenne* Chaudoir, 1869
CDMX⁴, GRO^{1,33}, HGO^{1,5,24,33}, MEX^{1,4,33}
- 0041.** *Calosoma costipenne isabellae* Lassalle and Van Den Berghe, 2011
MEX⁴¹
- 0042.** *Calosoma depressicolle depressicolle* Chaudoir, 1869
CAMP³, CDMX^{3,4}, HGO^{2,3,24}, JAL^{1,2,3}, MEX^{1,2,3,4,5,33}, MICH^{1,2,3,5}, MOR^{3,4}, PUE^{2,3,5}
- 0043[#].** *Calosoma derpessicolle altipeta* Jeannel, 1940
MEX^{1,40}
- 0044.** *Calosoma depressicolle asper* Jeannel, 1940
CDMX^{1,40}, GRO⁴⁰, MEX^{1,40}, MICH⁴⁰, ML^{1,40}, VC¹
- 0045[#].** *Calosoma depressicolle sylvestris* Lassalle, 2009
MEX^{38,40}
- 0046.** *Calosoma digueti digueti* (Lapouge, 1924)
DGO⁴, JAL^{1,2,3,33}, TLAX¹, ZAC^{1,2,3,5,33}
- 0047[#].** *Calosoma digueti colimaensis* (Lassalle, 2009)
JAL³⁸ (Häckel et al. 2011:54)
- 0048.** *Calosoma digueti hoegei* Breuning, 1928
JAL^{1,33} (Lassalle and Van Den Berghe 2011:51)
- 0049[#].** *Calosoma digueti mascotaensis* Lassalle and Van Den Berghe 2011
JAL⁴⁰
- 0050[#].** *Calosoma enigmaticum* Lassalle and Van Den Berghe, 2013
JAL⁴²
0051. *Calosoma eremicola* Fall, 1910
BCN^{1,5,26}
- 0052.** *Calosoma flohri flohri* Bates, 1884
COL^{1,33}, JAL^{1,2}, MEX^{1,4,5,33}, MICH^{1,2,3,4,5,18,33}, OAX⁵
- 0053[#].** *Calosoma flohri zirostense* Häckel, Heinz and Sehna, 2011
MICH (Häckel et al. 2011:54)
- 0054.** *Calosoma gebieni* (Breuning, 1931)
MICH^{1,33}, (Häckel et al. 2011:54), OAX³
0055. *Calosoma haydeni haydeni* G. Horn, 1870
CHIH¹, SLP³
- 0056.** *Calosoma haydeni punctulicolle* Bates, 1891
CDMX¹, CHIH^{1,2,19,33}, COAH^{1,33}, DGO^{1,2,3,19,33}, MOR³, NL^{1,19,33}, SLP^{1,5}
- 0057.** *Calosoma laeve* Dejean, 1826
CHIH^{1,2,3,33}, CDMX^{1,3,4,5,33}, GTO^{3,4}, HGO^{1,3,24,33}, JAL^{1,3,5}, MEX^{1,3,4,33}, MICH^{1,3,5,33}, MOR^{1,3,4,33}, NL^{1,33}, PUE^{1,3,4,33}, QRO^{3,4}, SLP⁵, TAMPS¹, VER^{1,33}
- 0058.** *Calosoma laevigatum* Chaudoir, 1869
CDMX^{3,4}, DGO^{1,2,33}, GTO^{1,33}, HGO^{1,5,24,33}, MEX^{1,4}, MICH³, PUE^{1,2,33}, TLAX^{2,3}, VER^{1,2,3,33}
- 0059.** *Calosoma lesnei* Breuning, 1931
MEX³³, MICH (Häckel et al. 2011:54)
0060. *Calosoma macrum* LeConte, 1853
COAH^{1,3}, NL^{3,4,33}, TAMPS^{1,33}
0061. *Calosoma marginale* Casey, 1897
BCN³, CHIH^{1,33}, COAH^{2,3,5}, MOR^{1,33}, NL^{1,3,4,33}, OAX³, SON^{3,6}, TAMPS^{1,2,3,4,33}, VER³
- 0062.** *Calosoma nitidum* Gehin, 1885
VER⁴¹
- 0063.** *Calosoma orizabae* (Jeannel, 1940)
VER^{1,33}
- 0064.** *Calosoma palmeri* G. Horn, 1876
BCN^{1,3,5,26,33} (Guadalupe Island)
0065. *Calosoma parvicolle* Fall, 1910
AGS³, BCN^{1,5,26,33}, CHIH³, JAL³, PUE³, QRO³, SLP³, SON^{1,2,3,4,5,6,33,54}
- 0066.** *Calosoma peregrinator* Guérin-Ménéville, 1844
AGS^{1,3,4,5,33}, BCN^{1,3,5,26,33}, BCS³, CAMP³, CDMX^{1,3,4,33}, CHIH^{1, 2,4,5,6,33}, COAH^{2,6,5}, DGO^{1,2,5,7,33}, GTO^{1,2,3}, HGO^{3,4,24}, JAL^{1,2,3,4,5,7,33}, MEX^{1,3,33}, MICH^{1,33}, MOR^{1,3,4,33}, NAY^{1,2,3,5,33}, NL^{1,2,3,4,33}, OAX⁵, PUE^{1,5}, QRO^{3,4}, SIN^{1,2,4,5}, SLP^{2,3,4}, SON^{1,2,3,5,6,33,54}, TAMPS³, VER^{1,3,5}, ZAC^{1,2,3,33}
- 0067.** *Calosoma politum politum* Chaudoir, 1869
CAMP³, CDMX^{3,4}, HGO³, PUE^{1,2}, MEX^{1,2,3,4,5,33}, MICH^{3,4}, MOR^{3,4}, TAB⁴, TLAX^{1,2}, VER^{2,5}
- 0068.** *Calosoma politum diminutum* Bates, 1891
MEX^{1,33,41}, VER¹
- 0069.** *Calosoma politum morelianum* Bates, 1891
CDMX^{1,33}, JAL^{2,3}, MEX⁴¹, MOR^{1,4,33,41}, VER^{1,33}
- 0070.** *Calosoma porosifrons* Bates, 1891
DGO^{1,2,3,5,19,33}
0071. *Calosoma prominens* LeConte, 1853
BCN^{1,33}, BCS^{3,5,26}, CAMP³, CHIH^{1,2,5}, COAH⁵, DGO^{1,2}, NL⁴, SON^{1,2,3,4,5,6,33,54}
0072. *Calosoma protractum* LeConte, 1862
BCS³, CHIH³, CDMX⁴, DGO^{1,33}, GTO^{1,33}, GRO^{1,2,5,33}, HGO²⁴, JAL^{1,3,5,33}, MEX^{4,5}, MICH^{1,2,5}, MOR^{1,3,4,5,33}, NAY^{1,5}, NL³³, PUE^{1,5,33}, OAX^{1,33}, SLP⁵, SON^{1,2,54}
- 0073.** *Calosoma rufinum* Gehin, 1885
MEX⁴², VC⁴²
0074. *Calosoma sayi* Dejean, 1826
BCS²⁶, CHIS^{1,2,3,4,5}, GRO^{3,4,5}, HGO^{3,5,24}, JAL^{3,5}, MICH³, NAY^{3,5}, NL^{1,3,5,33}, OAX^{1,2,3,4,5,33}, PUE^{1,33}, QROO³, SIN^{1,2,3,4,5,33}, SLP^{1,3,7,33}, SON^{1,3,54}, TAB^{2,3,4}, TAMPS^{1,2,3,4,5,33}, VER^{1,2,3,4,5,33}, YUC^{3,5}
0075. *Calosoma scrutator* (Fabricius, 1775)
AGS³, BCN^{1,26}, BCS^{3,5}, CDMX^{3,4,5}, CHIH^{1,2,3,5,33}, CHIS³, COAH³, COL³, GRO^{3,4}, JAL^{1,3,4,5}, MEX^{3,4}, MICH^{1,3}, MOR³, NAY^{1,2,3,5}, OAX^{1,2,3,4,5,33}, PUE^{3,5}, SIN^{3,5}, SLP³, SON^{1,2,3,5,6,54}, TAMPS^{2,3}, VER^{1,3,4,33}
0076. *Calosoma sponsa* Casey, 1897
BCN¹ (San Benito Island), CHIH³, SON^{5,54}
- 0077.** *Calosoma striatulum striatulum* Chevrolat, 1835
GRO^{1,5,33}, HGO¹, MEX², PUE^{1,4,5,33}, VER^{1,2,4,5,33}
- 0078[#].** *Calosoma striatulum libesensis* Lassalle, 2009
PUE^{38,41}

0079. *Calosoma striatulum striatipennis* Chaudoir, 1869
CDMX⁴, HGO⁴, MEX³, PUE^{1,4,5,33,41}, TLAX¹, VER^{1,5,33}
0080. *Calosoma viridisulcatum* Chaudoir, 1863
COL^{1,3,4,33}, GRO^{1,33}, GTO¹, JAL^{1,2,3,4,5,6,33}, NAY^{1,33}, QRO³³, ZAC^{1,33}
0081. *Carabus forreri* Bates, 1882
CDMX⁴, CHIH^{1,2,3,5,6}, DGO^{1,2,3,4,5,16}, GTO³, SIN², SON^{2,3,6,54}
0082. *Carabus hendrichsi* Bolivar and Pieltain, 1967
NL^{1,2,4,5}, TAMPS^{1,2}

05. Ctenostomatini

0083. *Ctenostoma maculicorne* (Chevrolat, 1856)
OAX^{1,3,56}, VER^{1,4,56}, YUC^{1,56}

06. Amblycheilini

0084. *Amblycheila baroni* (Rivers, 1890)
SON^{1,3,6,54,56}
0085. *Amblycheila halfferi* (Mateu, 1975)
SLP^{1,56}
- 0086*. *Amblycheila hoversoni* Gage, 1990
CHIH¹ (**note:** Erwin included but questioned Chihuahua); “Texas”⁵⁶; (type locality: “25.7 km S George West, Live Oak County, Texas”, Gage 1990: 2)
0087. *Amblycheila nyx* Sumlin, 1991
CHIH¹, COAH^{3,5,56}

07. Megacephalini

Note: Erwin’s superscript ¹ for Megacephalini refers to Erwin and Pearson 2008.

0088. *Oxycheila wittmeri* Wiesner, 1981
VER^{1,2,3,4,56}
0089. *Tetracha affinis* (Dejean, 1825)
NAY⁷, TAMPS³, VER⁶, South America⁵⁶
0090. *Tetracha angustata* (Chevrolat, 1841)
CAMP⁴, CHIH^{1,56}, CHIS^{3,4}, COAH^{1,56}, COL^{4,56}, GRO^{1,4,56}, NAY⁴, OAX^{1,56}, PUE⁴, SIN^{1,4,5}, SLP^{1,4,5}, TAB^{1,56}, TAMPS⁴, VER^{1,4,56}
0091. *Tetracha carolina carolina* (Linnaeus, 1767)
BCN⁵, BCS^{1,3,4,56}, CHIH^{1,3,4,5,56}, CHIS^{1,3,4,56}, CDMX⁵, COAH^{1,3,56}, COL^{1,3,4,56}, DGO^{1,3,56}, GRO^{1,3,4,7,56}, GTO³, HGO^{3,4}, JAL^{3,5,6}, MEX⁵, MICH^{3,4}, MOR^{1,3,4,5,7,56}, NL^{1,3,4,7,56}, OAX^{1,3,4,7,56}, PUE^{1,3,7,56}, QRO^{3,4,56}, SIN^{1,3,4,5,7,56}, SLP^{3,4}, SON^{1,3,4,5,6,7,54,56}, TAB^{1,3,56}, TAMPS^{1,3,4,5,56}, VER^{1,56}, YUC⁷
0092. *Tetracha carolina chevrolatii* (Chaudoir, 1860)
CHIS⁵⁶, SLP^{1,56}, VER^{1,56}, YUC^{1,7,56}
0093. *Tetracha cyanides* Bates, 1881
CDMX⁵, GRO^{1,56}, JAL^{1,5,56}, MICH^{1,56}, MOR^{1,5,56}, NAY^{1,56}, OAX^{1,4}, SLP⁵, VER^{1,56}
0094. *Tetracha sobrina sobrina* (Dejean, 1831)
BCS^{1,56}, GRO³, MOR³, OAX^{1,3,56}, TAB⁵⁶, TAMPS^{1,56}, VER^{1,3,56}
- 0095*. *Tetracha sobrina infuscata* (von Mannerheim, 1837)
“México”¹; (no type locality/area by von Mannerheim 1837: 6); “Cuba” (Blackwelder 1944: 13); “Hispaniola, Puerto Rico, West Indies” (Boyd and Associates 1982: 4; Weisner 2020: 20)

0096. *Tetracha virginica* Linnaeus, 1766
TAB⁵⁶

08. Cicindelini

Note: Erwin’s superscript ¹ for the tribe Cicindelini refers to Erwin and Pearson 2008.

0097. *Beckerium leptalis* (Bates, 1881)
OAX^{3,4,15,56}, VER¹⁵
0098. *Brasiella argentata argentata* (Fabricius, 1801)
VER⁴; **note:** most likely mis-identified or mis-labelled; type locality: “America meridionali” Fabricius 1804: 42; “Mexico”¹, (Blackwelder 1944:17); **note:** “South America” (Wiesner 2020: 298)
0099. *Brasiella chiapasi* van Nidek, 1980
CHIS^{1,56}
0100. *Brasiella hemichrysea hemichrysea* (Chevrolat, 1835)
CAMP⁴, CHIS^{1,2,3,4,23}, HGO^{1,3,4,7}, JAL^{1,4,7}, MICH⁴, NAY^{3,4,7,56}, OAX^{1,3,4,7}, PUE^{4,7}, SIN^{5,23,56}, SLP^{1,3,7,23}, SON¹, TAB^{1,2,3}, TAMPS^{1,3,4}, VER^{1,2,3,4,7,23}, YUC^{1,23}
0101. *Brasiella hemichrysea inspersa* (Chevrolat, 1835)
CHIH⁵⁶, CHIS⁵⁶, DGO⁵⁶, GRO⁵⁶, HGO^{5,56}, JAL^{5,56}, NAY⁵, PUE⁵⁶, QRO⁵⁶, QROO⁵, OAX⁵⁶, SP⁵⁶, SON⁵⁶, TAB⁵⁶, TAMPS⁵⁶, VER^{5,56}, YUC⁵⁶
0102. *Brasiella mandli* van Nidek, 1978
CHIS^{1,5,56}
0103. *Brasiella maya* Cassola and Sawada, 1990
YUC^{1,56}; (type locality: “Yokdzonot, Yaxcaba Municipality, Yucatan, México”, Cassola and Sawada 1990: 6)
0104. *Brasiella praecisa* (Bates, 1890)
COAH⁵, COL^{1,23,56}, GRO^{1,3,4,5,7,23,56}, JAL^{1,2,3,4,56}, MICH^{4,56}, MOR^{1,4,5,23,56}, OAX^{1,56}, PUE^{1,4,5,7,56}, TAMPS⁵⁶
0105. *Brasiella speculans* (Bates, 1890)
GRO^{1,5,23,56}
0106. *Brasiella viridisticta viridisticta* (Bates, 1881)
DGO⁴, MICH^{4,5}, MOR^{4,7}, OAX^{1,4,15,7,23,56}, PUE^{7,56}
0107. *Brasiella viridisticta arizonensis* (Bates, 1884)
CHIH^{1,3,4,23,56}, SON^{2,3,4,5,18,23,54,56}
0108. *Brasiella viridisticta interjecta* (W. Horn, 1935)
JAL^{1,5,23,56}, MICH⁵⁶
0109. *Brasiella wickhami* (W. Horn, 1903)
BCN^{1,56}, BCS^{23,26}, CHIH³, HGO³, SIN^{1,2,3,4,5,23,56}, SLP³, SON^{1,2,4,5,23,54,56}
0110. *Cicindela cazieri* Vogt, 1949
TAMPS⁴
0111. *Cicindela carolae* Gage and McKown, 1991
COAH^{1,3,4,56}
0112. *Cicindela hirticollis hirticollis* Say, 1817
BCN³, CHIH⁴, VER⁴
0113. *Cicindela hirticollis corpuscula* Rump, 1962
BCN^{21,26}
0114. *Cicindela hirticollis gravida* LeConte, 1851
BCN^{3,4,5,6,7,23,56}
0115. *Cicindela hirticollis ponderosa* (Thomson, 1859)
BCN²³, CHIH⁴, VER^{1,56}
0116. *Cicindela latesignata latesignata* (LeConte, 1851)
BCN^{1,3,4,5,7,23,56}, BCS^{3,4}, SON^{3,4,7}

- 0117.** *Cicindela latesignata parkeri* (Cazier, 1948)
BCN^{1,4,5,56}, BCS^{4,56}, SON^{3,4,5,23,54,56}
- 0118.** *Cicindela oregona oregona* (LeConte, 1860)
BCN^{1,3,4,5,56}, COL⁷, JAL³, NL³, SON^{23,56}
- 0119.** *Cicindela pimeriana* (LeConte, 1866)
SON^{1,23,56}
- 0120.** *Cicindela senilis* Horn, 1867
BCN⁴
- 0121.** *Cicindelidia aeneicollis* (Bates, 1881)
AGS^{1,56}, CAMP⁵⁶, CHIH^{3,4,56}, CHIS^{1,3,56}, COL^{1,4,5,23,56}, DGO^{1,7,23,56}, GRO^{1,3,4,23,56}, JAL^{1,2,3,4,5,7,23,56}, MICH^{1,4,5,23,56}, MOR^{1,2,3,4,5,7,23,56}, NAY^{1,4,5,23,56}, OAX^{1,4,15,56}, PUE^{1,5,56}, SIN^{1,2,4,5,7,23,56}, SON^{1,2,5,23,54,56}
- 0122.** *Cicindelidia aterrima* (Klug, 1834)
AGS^{1,4,56}, CDMX^{1,3,4,23,56}, CHIH^{1,3,4,23,56}, DGO^{1,3,4,23,56}, GTO^{1,23,56}, HGO^{1,4,23,24,56}, JAL^{1,3,56}, MEX^{4,23}, MICH^{1,4,5,23,56}, PUE^{1,4,7,56}, QRO^{1,56}, SLP^{1,56}, TLAX⁴, ZAC^{1,4,56}
- 0123.** *Cicindelidia aurora aurora* (Thomson, 1859)
GRO^{1,4,23,56}, OAX^{1,2,3,4,7,23,56}
- 0124.** *Cicindelidia aurora candelarensis* (Johnson, 1998)
OAX^{1,56}
- 0125.** *Cicindelidia aurora romani* (Johnson, 1998)
OAX^{1,3,56}
- 0126.** *Cicindelidia beneshi* (Vargas-Arangua, 1930)
SON^{1,3,4,5,7,23,54,56}
- 0127.** *Cicindelidia carthagera carthagera* (Dejean, 1831)
BCS⁴, CAMP⁵⁶, CHIH⁵⁶, CHIS^{1,4,56}, COL^{1,3,56}, GRO^{1,3,4,7,23,56}, GTO⁵⁶, JAL^{1,3,4,23}, MICH^{1,56}, MOR⁵⁶, NAY^{1,3,4,5,23,56}, NL⁵, OAX^{1,2,3,4,5,23,56}, SIN^{1,2,3,4,5,7,23,56}, SON^{3,4,5,23,54,56}
- 0128.** *Cicindelidia carthagera colosseae* (W. Horn, 1926)
BCN^{1,3,5,23,56}, BCS^{3,5,23,56}, JAL³, SON^{1,5,23,54}
- 0129.** *Cicindelidia chrysippe* (Bates, 1884)
DGO^{1,4,18,23}, SIN^{1,3,4,5}
- 0130.** *Cicindelidia clarina clarina* (Bates, 1881)
CDMX^{1,4,23,56}, COAH⁵, GRO³, HGO^{1,3,4,23,56}, MEX^{1,4,5,23,56}, MICH^{1,5,23,56}, MOR³, OAX^{1,56}, PUE^{1,3,4,23,56}, SLP^{3,4}, ZAC^{1,56}
- 0131.** *Cicindelidia clarina flexifasciata* (Mandl, 1970)
QRO⁵⁶, SLP^{1,56}, TAMPS⁵⁶
- 0132.** *Cicindelidia craveri* (Thomson, 1856)
HGO^{1,56}
- 0133.** *Cicindelidia cyaniventris cyaniventris* (Chevrolat, 1834)
CAMP^{3,4,7}, CDMX⁴, CHIH⁵⁶, CHIS^{1,2,3,4,5,56}, HGO^{3,4,56}, JAL^{1,2,56}, PUE^{5,3}, QRO^{3,56}, QROO^{4,7}, OAXC^{7,56}, SLP^{1,2,4,5,7,23,56}, TAB⁵⁶, TAMPS^{1,3,7,23,56}, VER^{1,2,3,4,7,23,56}, YUC^{1,2,4,7,23,56}
- 0134.** *Cicindelidia dysenterica* (Bates, 1881)
COL^{1,4,5,23,56}, GRO^{1,4,5,7,23,56}, GTO^{1,3,56}, HGO^{1,4,23,56}, JAL^{3,4,7}, MEX^{1,4,5,7,23,56}, MICH^{1,7,23,56}, MOR^{1,4,5,7,23,56}, NAY^{1,4,23,56}, OAX^{1,3,4,7,15,23,56}, PUE^{1,3,5,7,15,23,56}, SIN^{1,4,5,7,56}, TAMPS^{3,56}, TLAX⁷
- 0135.** *Cicindelidia euthales* (Bates, 1882)
CDMX⁴, DGO^{1,4,16,23,56}, HGO⁷, MEX^{1,23,56}, TLAX⁷
- 0136.** *Cicindelidia fera* (Chevrolat, 1834)
CHIH⁵⁶, CHIS^{23,56}, COAH^{23,56}, COL⁵⁶, DGO^{23,56}, GRO^{4,5,7,23,56}, JAL⁵⁶, MICH⁴, MOR^{3,5}, NAY^{4,23,56}, QRO^{3,56}, OAX^{23,56}, SIN^{4,5,3,7,23,56}, SON^{14,5,23,54,56}, TAB^{23,56}, VER^{3,4,56}
- 0137.** *Cicindelidia flohri* (Bates, 1878)
CDMX^{1,4,23,56}, CHIH^{1,3,4,23,56}, DGO^{1,3,4,23,56}, GRO⁵, GTO^{1,23,56}, HGO²⁴, JAL^{3,5}, MEX^{1,3,23,56}, MICH^{1,4,23,56}, PUE^{1,56}, TLAX^{1,56}, VER^{1,56}, ZAC^{1,23,56}
- 0138.** *Cicindelidia guerrenensis* (Bates, 1890)
COL^{1,4,56}, GRO^{1,3,4,7,23,56}, MICH^{3,4}, MOR^{1,3,4,5,56}, OAX^{1,56}, PUE^{1,5,56}
- 0139.** *Cicindelidia hemorrhagica hemorrhagica* (LeConte, 1851)
BCN^{1,3,4,5,23,56}, MICH³, SON^{3,5,54}
- 0140.** *Cicindelidia hemorrhagica hentziana* (Leng, 1918)
BCN¹, BCS^{5,23,56}
- 0141.** *Cicindelidia hornii hornii* (Schaupp, 1884)
CHIH^{1,2,3,4,23,56}, DGO^{1,3,7,23,56}
- 0142.** *Cicindelidia hornii scotina* (Bates, 1890)
CHIH^{1,23,56}, DGO^{1,5,23,56}, ZAC⁵⁶
- 0143.** *Cicindelidia hydrophoba hydrophoba* (Chevrolat, 1835)
CAMP^{3,4,7}, CDMX⁴, CHIH^{4,56}, CHIS^{4,7}, COL⁴, GRO^{3,4,7}, HGO⁵⁶, JAL^{3,4,7}, MEX^{4,7}, MICH^{4,7}, MOR^{3,4,7}, NAY^{4,7}, OAX^{3,4,7}, SIN^{3,4,7}, SLP⁵⁶, VER^{1,4,5,7,23,56}, YUC^{4,7}
- 0144.** *Cicindelidia hydrophoba atroreducta* (W. Horn, 1930)
QRO¹, YUC^{1,2,3,23}
- 0145.** *Cicindelidia hydrophoba quinquenotata* (Gistel, 1837)
CHIH⁵, CHIS^{1,5,56}, GRO^{1,3,23,56}, JAL⁵⁶, OAX^{1,2,3,5,23,56}, VER³
- 0146.** *Cicindelidia hydrophoba taretana* (Bates, 1884)
COL^{1,23,56}, GRO^{1,2,3,23,56}, JAL^{3,56}, MEX^{1,23,56}, MICH^{1,3,18,23,56}, MOR^{1,3,23,56}, NAY^{1,56}, PUE^{1,56}, SIN^{1,2,23,56}, SON^{1,23,56}
- 0147.** *Cicindelidia ioessa* (Bates, 1881)
MEX⁴, MOR⁴, PUE^{1,15,23,56}, TLAX^{1,23,56}
- 0148.** *Cicindelidia juvenilis* (W. Horn, 1897)
JAL^{1,4,5,7,56}, NAY^{1,3,4,7,56}, SIN^{1,3,4,5,7,23,56}, SON^{1,5,23,54,56}
- 0149.** *Cicindelidia klugii* (Dejean, 1831)
CDMX^{1,4,5,23,56}, HGO^{1,23,56}, MEX^{1,5,23,56}, MOR⁵, OAX⁵, PUE⁷
- 0150.** *Cicindelidia laetipennis* W. Horn 1913
COAH^{1,23,56}
- 0151.** *Cicindelidia lisaannae* (Gage, 1991)
CHIH⁵⁶, CHIS^{1,3,4,56}
- 0152.** *Cicindelidia luteolineata* (Chevrolat, 1856)
COL^{1,23,56}, CDMX^{1,4,5,23,56}, GTO⁴, JAL^{1,4,5,7,23,56}, MEX^{1,2,3,23,56}, MICH^{1,4,23,56}
- 0153.** *Cicindelidia melissa* Duran and Roman, 2014
CHIH^{2,3,56}, DGO⁵⁶, SLP³, SON^{2,3,54,56}
- 0154.** *Cicindelidia nebuligera* (Bates, 1890); Figure 1B
CDMX⁵, CHIH⁵, DGO^{1,2,3,4,23,56}, ZAC^{1,5,56}
- 0155.** *Cicindelidia nigrilabris* (Bates, 1890)
CHIH^{1,3,4,23,56}, DGO^{1,2,3,5,23,56}, HGO^{1,4,56}, MEX^{7,56}, MICH^{1,3,56}, PUE⁴
- 0156.** *Cicindelidia nigrocoerulea nigrocoerulea* (LeConte, 1848)
AGS^{1,5,56}, CHIH^{1,3,4,7,23,56}, COAH^{1,4,23,56}, DGO^{1,3,4,7,23,56}, MEX⁵⁶, NL⁵⁶, SLP^{1,56}, SON^{1,23,56}, ZAC^{1,56}

0157. *Cicindelidia nigrocoerulea subtropica* (Vogt, 1949)
TAMPS^{1,56}
0158. *Cicindelidia oaxacensis* (Johnson, 1994)
OAX^{1,2,3,4,56}
0159. *Cicindelidia obsoleta latemaculata* (Becker, 1897)
DGO^{1,3,4,23,56}
0160. *Cicindelidia obsoleta santaclarae* (Bates, 1890); Figure 1C
CHIH^{3,4,5,23,56}, DGO^{1,4,5,23,56}, SON^{2,54}
0161. *Cicindelidia obsoleta vulturina* (LeConte, 1853)
COAH^{1,23,56}, COL⁵⁶
0162. *Cicindelidia ocellata ocellata* (Klug, 1834)
AGS^{1,4,23,56}, CAMP^{3,4}, CDMX^{1,23,56}, CHIH^{3,4,7,56}, CHIS^{1,2,4,7,23,56}, COAH^{2,3}, COL^{1,23,56}, DGO^{2,4,7}, GRO^{3,4,7,23}, GTO^{1,3,4,23,56}, HGO⁵⁶, JAL^{1,2,3,4,23,56}, MICH^{1,2,4,7,23,56}, MOR^{1,2,3,4,7,23,56}, NAY^{1,3,4,23,56}, NL³, OAX^{1,2,3,4,7,23,56}, PUE⁴, QRO^{3,4}, SIN^{1,2,3,4,7,23,56}, SLP^{1,3,4,7,23,56}, SON^{1,2,3,4,7,23,54,56}, TAB^{1,2,3,4,56}, TAMPS^{2,3,4}, VER^{1,2,3,4,7,23,56}, YUC^{1,3,4,7,23,56}, ZAC^{1,4,23,56}
0163. *Cicindelidia ocellata rectilatera* (Chaudoir, 1843)
CHIH^{1,3,4,5,23,56}, COAH^{1,3,7,23,56}, COL⁵⁶, DGO^{3,4,5}, NL^{1,3,5,7,23,56}, QRO³, TAB⁵⁶, TAMPS^{1,23,56}, VER²
0164. *Cicindelidia papillosa* (Chaudoir, 1854)
CHIH⁵⁶, CHIS^{1,3,4,7,56}, DGO^{1,23,56}, GRO^{4,23,56}, GTO¹, JAL^{1,56}, MEX^{1,56}, MICH^{1,23,56}, MOR^{1,56}, NAY^{1,2,56}, NL⁴, OAX^{1,2,4,5,23,56}, PUE^{1,4,56}, QRO^{1,3,56}, SIN^{5,56}, SLP^{1,56}, TAMPS^{4,56}, VER^{1,3,4,5,7,23,56}
0165. *Cicindelidia phosphora* (Bates, 1878)
COL^{1,4,5,7,56}, GRO^{1,3,4,23,56}, MICH^{1,56}, MOR^{1,23,56}
0166. *Cicindelidia politula* (LeConte, 1875)
COAH^{1,3,4,23,56}, NL^{1,4,23,56}
0167. *Cicindelidia pseudoaurora* (Johnson, 1998)
GRO^{1,3,4,56}
0168. *Cicindelidia pseudoeuthales* (Sumlin, 1991)
NL⁴
0169. *Cicindelidia pseudoradians* (Johnson, 1998)
CHIS^{1,3,4,56}
0170. *Cicindelidia punctulata punctulata* Olivier, 1790
CHIH^{3,4}, TAMPS⁴
0171. *Cicindelidia punctulata catharinae* (Chevrolat, 1835)
CDMX^{4,5,23,56}, CHIH^{23,56}, CHIS¹, COAH^{3,5}, DGO^{7,23,56}, HGO^{23,56}, MEX^{4,5,23}, NL³, PUE^{2,3,23,56}, QRO⁵⁶, SLP⁵⁶, TAB⁵, TLAX^{3,4,7,56}, ZAC^{4,23,56}
0172. *Cicindelidia punctulata chihuahuae* (Bates, 1890)
CHIH^{4,23,56}, SON^{23,56}
0173. *Cicindelidia radians radians* (Chevrolat, 1841)
CHIH^{4,56}, CHIS^{1,2,3,5,7,23,56}, OAX^{1,2,3,4,5,56}, SLP⁵⁶, TAB⁴, VER^{1,2,3,4,5,7,23,56}
0174. *Cicindelidia roseiventris roseiventris* (Chevrolat, 1834)
CHIS^{1,3,23,56}, OAX^{1,3,7,56}, QRO^{3,7}, SLP⁷, TAB^{1,23,56}, TAMPS^{3,7}, VER^{1,23,56}
0175. *Cicindelidia roseiventris mexicana* (Klug, 1834)
HGO^{1,23,56}, NL^{1,23,56}, SLP^{1,23,56}, TAMPS^{1,5,23,56}, VER^{1,23,56}
0176. *Cicindelidia rufiventris cumatilis* LeConte, 1851
CAMP⁴, CHIS⁷, HGO⁷, MOR⁷, NL⁴, PUE⁷, SLP^{4,7}, TAMPS^{4,7}, VER⁴
0177. *Cicindelidia rufiventris reducens* (W. Horn, 1915)
COL^{1,23,56}, JAL^{1,4,5,56}, VER¹
0178. *Cicindelidia rugatilis* (Bates, 1890)
AGS^{1,4,5,56}, CDMX^{1,23,56}, DGO^{1,4,7,23,56}, GTO³, JAL^{1,3,4,23,56}, MEX^{1,2,3,4,5,23,56}, MICH^{1,5,56}, PUE⁵⁶, TLAX⁵⁶, ZAC^{1,23,56}
0179. *Cicindelidia schauppii* (G. Horn, 1876)
NL^{1,4,19,23,56}, TAMPS³
0180. *Cicindelidia sedecimpunctata sedecimpunctata* (Klug, 1834)
AGS^{3,4}, CDMX⁴, CHIH^{1,3,4,5,23,56}, CHIS^{3,4}, COAH^{1,56}, COL³, DGO^{23,56}, GTO^{1,4,23,56}, HGO^{3,4,7}, JAL^{3,4}, MEX⁴, MOR^{2,3,4}, NAY⁴, NL^{3,4}, OAX^{3,4}, PUE⁷, QRO³, SIN^{1,2,3,5,56}, SON^{1,2,3,4,5,7,23,54,56}, SLP^{2,3,4}, TAMPS^{2,3}, VER^{3,4}, ZAC^{1,3,23,56}
0181. *Cicindelidia sedecimpunctata meridiana* (van Nidek, 1978)
CHIS^{1,56}, OAX^{1,56}
0182. *Cicindelidia sedecimpunctata rubriventris* (Chevrolat, 1835)
CDMX^{1,23,56}
0183. *Cicindelidia sedecimpunctata sallei* (Chevrolat, 1835)
CHIS^{1,56}, GRO^{1,23,56}, JAL⁵, MICH^{1,5,56}, MOR^{1,5,23,56}, OAX^{1,23,56}, PUE^{1,5,56}, QRO¹, SIN^{1,5,56}, SLP^{23,56}, TAMPS^{1,23,56}, VER^{1,5,56}, ZAC^{23,56}
0184. *Cicindelidia semicircularis* (Klug, 1834)
CDMX^{1,4,23,56}, CHIH^{1,4,7,23,56}, CHIS⁵⁶, DGO^{3,4,5,23,56}, GTO^{1,4,23,56}, HGO^{1,4,23,56}, MEX^{1,4,5,23,56}, PUE⁵⁶, SLP^{1,56}, VER⁵⁶, ZAC^{1,23,56}
0185. *Cicindelidia smaragdina* (Chevrolat, 1835)
CDMX^{4,5,56}, HGO^{4,56}, MEX⁵⁶, PUE³, VER^{4,23,56}
0186. *Cicindelidia sommeri* (von Mannerheim, 1837)
BCN^{1,56}, CHIH^{3,4,56}, CHIS^{1,4,56}, COAH⁵, COL^{1,3,23,56}, DGO⁵, GRO^{1,3,4,5,7,23,56}, JAL^{1,2,3,4,23,56}, MEX^{4,5}, MICH^{1,4,7,23,56}, MOR^{1,3,4,5,7,23,56}, NAY^{1,3,4,5,7,56}, OAX^{1,2,3,4,5,7,23,56}, PUE^{1,5,23,56}, QRO⁵⁶, SIN^{1,2,3,5,33,56}, SON^{1,2,3,4,5,23,54,56}, ZAC^{4,56}
0187. *Cicindelidia stephaneae* (McKown and Gage, 1991)
COAH^{1,3,4,56}
0188. *Cicindelidia tenuisignata* (LeConte, 1851)
BCN^{1,4,23,26,56}, CHIH^{1,23,56}, COAH^{1,3,4,7,23,56}, DGO^{1,3,23,56}, NL⁵, SIN^{1,4,5,23,56}, SON^{1,4,5,23,54,56}, TAMPS^{1,23,56}
0189. *Cicindelidia thalestris thalestris* (Bates, 1890)
CHIH^{1,3,4,23,56}, DGO^{1,4,5,23,56}, MEX^{1,56}, SIN⁵, SON⁵⁴
- 0190[#]. *Cicindelidia thalestris cyaneocolorata* Mandl, 1970
DGO⁵⁶
0191. *Cicindelidia trifasciata ascendens* (LeConte, 1851)
BCS^{3,4}, CAMP^{1,4,23}, CDMX^{1,4,5,2,56}, CHIH⁵⁶, CHIS^{1,56}, COAH^{3,4,56}, COL^{1,56}, GRO^{4,7,23,56}, HGO⁵⁶, JAL^{1,3,4,23,56}, MEX^{1,4,23,56}, MICH^{1,4,7,56}, NAY^{1,3,23,56}, NL^{3,4}, OAX^{1,2,3,4,5,56}, QRO^{1,3,4,56}, SIN^{1,2,3,4,5,7,23,56}, SON^{1,5,23,54,56}, TAB¹, TAMPS^{1,3,4,23,56}, TLAX⁵⁶, VER^{1,3,4,23,56}, YUC³
0192. *Cicindelidia trifasciata sigmoidea* (LeConte, 1851)
BCN^{1,3,23,56}, BCS^{3,23}, SIN³
0193. *Cicindelidia vasseletii* (Chevrolat, 1834)
CHIS^{1,3,4,5,56}, OAX^{1,3,4,5,7,56}, VER^{1,3,4,5,23,56}
0194. *Cicindelidia veracruzensis* (Johnson, 1998)
?SLP⁵⁶, VER^{1,4,56}, “México”³
0195. *Cylindera hoegei* (Bates, 1881)
OAX^{1,3,4,15,23,56}

0196. *Cylindera lemniscata lemniscata* (LeConte, 1854)
BCN³, BCS^{3,4}, CHIH^{2,3,7}, COAH^{3,7}, NAY^{1,56}, NL³,
PUE⁵, SIN^{1,3,4,56}, SLP³, SON^{1,2,3,4,7,23,54,56}
0197. *Cylindera lemniscata rebaptisata* (Vaurie, 1951)
BCN^{3,23,26}, BCS^{3,23,26}, CHIH^{1,3,5,23,56}, COAH^{1,3,4,5,23,56},
DGO^{1,3,4,23,56}, NL^{1,3,56}, OAX^{3,4}, SIN^{3,4}, SLP^{3,4,56},
TAMPS^{1,23,56}, ZAC⁵⁶
0198. *Cylindera lemniscata bajacalifornica* (Shook, 1989)
BCN^{1,3,5,56}, BCS^{2,3,4,5,56}
0199. *Cylindera nudata* (W. Horn, 1915)
DGO^{1,23,56}
0200. *Cylindera terricola terricola* (Say, 1824)
BCS^{21,23,26}
0201. *Dromochorus belfragei* Sallé, 1877
TAMPS¹
0202. *Ellipsoptera hamata hamata* (Audouin and Brullé)
CAMP^{7,56}, MOR⁷, QROO^{3,7}, TAB^{3,4,56}, VER^{1,2,3,4,5,7,23,56},
YUC³
0203. *Ellipsoptera hamata monti* (Vaurie, 1951)
QROO⁴, TAMPS^{3,4,56}
0204. *Ellipsoptera hamata pallifera* (Chaudoir, 1852)
QROO^{1,3,4,5,56}, TAB¹, YUC^{1,3,4,5,23,56}
0205. *Ellipsoptera lepida* (Dejean, 1831)
CHIH^{23,56}
0206. *Ellipsoptera marutha* (Dow, 1911)
CHIH^{1,3,7,23,56}
0207. *Ellipsoptera nevadica nevadica* (LeConte, 1875)
COAH³, SON^{1,4,23,54,56}
0208. *Ellipsoptera nevadica citata* (Rumpp, 1977)
SON¹; Arizona and New Mexico, United States⁵⁶
0209. *Ellipsoptera nevadica olmosa* (Vaurie, 1951)
COAH^{1,5,56}, ?TAMPS⁵⁶
0210. *Ellipsoptera nevadica metallica* (Sumlin, 1990)
COAH^{3,4,5,56}
0211. *Ellipsoptera rubicunda* (Harris, 1911)
CHIH⁴
0212. *Ellipsoptera sperata sperata* (LeConte, 1857)
CHIH^{1,4,23,56}, CHIS³, COAH^{1,56}, DGO^{1,3,4,23,56}, SLP⁷,
TAMPS^{1,23,56}
0213. *Ellipsoptera sperta vauriei* (Cazier, 1954)
SON^{1,3,5,7,23,54,56}
0214. *Eunota californica californica* Ménétries, 1843
BCN^{1,4}, BCS^{4,5,23,56}
0215. *Eunota californica brevihamata* (W. Horn, 1908)
SIN^{1,2,3,4,5,23,56}, SON^{1,3,4,5,23,54,56}
0216. *Eunota californica mojavi* (Cazier, 1937)
BCN^{1,5,23,56}, SON^{1,2,5,23,54,56}
0217. *Eunota circumpicta circumpicta* (LaFerté-Sénéctère, 1841)
BCN⁴, TAMPS^{1,473,56}
0218. *Eunota circumpicta johnsonii* (Fitch, 1857)
COAH^{1,4,56}, TAMPS³
0219. *Eunota fulgoris fulgoris* (Casey, 1913)
CHIH^{1,56}; (type locality: “El Paso, Texas”, Casey 1913: 34)
0220. *Eunota gabbii* (G. Horn, 1866)
BCN^{4,5}, BCS^{4,23,56}, SIN^{1,4,5,23,56}, SON^{1,3,4,5,23,54,56}
0221. *Eunota mecocheila* Duran and Roman, 2021
COAH^{2,59}
0222. *Eunota pamphila* (LeConte, 1873)
TAMPS^{1,2,3,4,23,56}
0223. *Eunota rockefelleri* (Cazier, 1954)
SON^{1,5,7,8,23,54,56}
0224. *Eunota severa severa* (LaFerté-Sénéctère, 1841)
CAMP^{3,4}, TAMPS^{1,4,23,56}, VER⁴
0225. *Eunota severa yucatana* (W. Horn, 1896)
YUC^{1,23,56}
0226. *Eunota togata togata* (LaFerté-Sénéctère, 1841)
TAMPS^{1,3,4,23,56}, VER^{3,4}
0227. *Habroscelimorpha auraria* (Klug, 1834)
SIN⁷
0228. *Habroscelimorpha curvata* (Chevrolat, 1834)
CAMP^{3,4}, CHIS⁴, ?DGO⁵⁶, GRO⁴, TAB^{3,4,56},
VER^{1,2,3,4,5,7,23,56}
0229. *Habroscelimorpha dorsalis saulcyi* (Guérin-Ménéville, 1840)
CAMP (Arcas Islands, Gulf of México)^{1,18,23,56}
0230. *Habroscelimorpha dorsalis venusta* (LaFerté-Sénéctère, 1841)
TAMPS^{1,4,23,56}, VER⁴
0231. *Habroscelimorpha eurysepta* (Bates, 1890)
OAX³, SIN^{1,5,23,56}
0232. *Habroscelimorpha wellingi* (Cassola and Sawada, 1990)
YUC^{1,3,4,56}; (type locality: “Uaymitun, Yucatan, México”, Cassola and Sawada 1990: 2)
0233. *Iresia boucardi* Chevrolat, 1856
VER^{1,3,56}
0234. *Microthylax bradti* (Cazier, 1954)
JAL^{4,56}, NAY^{1,5,23,56}, “México”³
0235. *Microthylax digueti* (W. Horn, 1897)
BCN^{1,3,5,23,56}, BCS^{3,5,23,56}, SON^{1,5,23,54,56} (Tiburón Island)
0236. *Microthylax sinaloae sinaloae* (Bates, 1890)
BCS^{3,7}, CHIH³, JAL³, NAY^{3,56}, SIN^{1,5,7,23,56},
SON^{1,5,7,23,54,56}
0237. *Microthylax sinaloae schrammeli* (Cazier, 1954)
BCN^{1,4,5,56}, SON^{1,3,4,5,8,54,56}
0238. *Odontocheila ignita* Chaudoir, 1860
CHIS^{1,56}; **note**: Chaudoir (1860: 320) writes, (English translation): “This pretty species of which I only have one male without abdomen, was noted in the Reiche collection as coming from Mexico, but without more precise information, I must point out that it was not found among the many insects that M. Sallé collected and received from this country.”; recorded from México, Guatemala and El Salvador (Catalogue of Life: <https://www.catalogueoflife.org/data/taxon/48PKK>)
0239. *Odontocheila mexicana* Laporte de Castelnau, 1834
CHIS^{2,3,4,5,7,56}, GRO⁵⁶, JAL⁵⁶, PUE⁵⁶, OAX^{2,4,56}, SLP⁷,
VER^{1,2,3,4,5,56}
0240. *Odontocheila potosiana* Moravec, Broska and Huber, 2017
HGO²⁴, SLP⁵⁶
0241. *Opilidia chlorocephala* (Chevrolat, 1834)
TAB^{1,4,23,56}, VER^{1,2,3,4,5,7,23,56}
0242. *Opilidia leucanoe* (Bates, 1890)
COL^{1,4,5,7,23,56}, GRO^{1,3,4,5,7,23,56}, JAL^{1,23,56}, MICH^{1,4,23,56},
OAX^{1,3,4,7,23,56}

0243. *Opilidia macrocnema macrocnema* (Chaudoir, 1852)
BCN^{1,56}, BCS²³, CHIH⁵⁶, CHIS^{1,3,4,5,56}, GRO^{1,4,23,56},
JAL^{1,4,7,23,56}, NAY^{1,3,4,5,23,56}, OAX^{1,4,23,56}, SIN^{1,2,3,4,5,23,56},
SON^{1,2,3,4,5,7,23,54,56}

0244. *Opilidia macrocnema kino* (Cazier, 1954)
SON^{1,3,5,8,23,54,56}

0245. *Parvindela debilis* (Bates, 1890)
COAH^{1,23,56}, DGO^{1,4,5,23,56}

0246. *Parvindela hualonga* (Schaupp, 1884)
BCN^{1,23,56}

0247. *Parvindela nephelota* (Bates, 1882)
CHIH⁴, DGO^{1,2,3,4,16,23,56}

09. Loricerini

0248. *Loricera aptena* Ball and Erwin, 1969; Figure 1D
CHIH^{1,2}, DGO^{1,2,3,5}, MICH^{1,3}, SIN^{1,2}

0249. *Loricera decempunctata* Eschscholtz, 1833
VER⁴; **note:** most likely mis-identified or mis-labelled;
type locality: “Sitka Sound, Alaska”, [United States],
Eschscholtz 1833: 25.

0250. *Loricera rotundicollis* Chaudoir, 1863
CDMX⁴, GRO^{1,2}, HGO^{1,2,24}, JAL^{1,2}, MEX^{1,2}, MICH^{1,2},
MOR^{1,2}, OAX^{1,2}, PUE^{1,2}, QRO^{1,2}, SIN⁵, SLP^{1,2},
VER^{1,2,3,4,52}

10. Omophronini

0251. *Omophron baenningeri* Dupuis, 1912
DGO², HGO², PUE², VER^{1,2}

0252. *Omophron dentatum* LeConte, 1852
BCN⁵

0253. *Omophron gilae* LeConte, 1852
SON⁵⁴; (type locality: “Ad fluminis Gilae ripas” [= on
the banks of the Gila River], LeConte 1852: 201)

0254. *Omophron gratum* Chaudoir, 1868
CHIH¹, CHIS^{2,57}, COAH⁵, GRO³, GTO², HGO², JAL⁵,
MEX⁵, MICH⁵, MOR⁵, NAY⁵, NL², OAX⁵, PUE^{2,4},
SIN^{1,5}, SLP^{3,5}, SON^{5,54}, TAMPS², VER^{1,4,5}

0255. *Omophron mexicanum* Dupuis, 1912
VER^{3,5}

0256. *Omophron oblitteratum* G. Horn, 1870
CHIH⁵, DGO^{1,2}, SON^{5,54}, ZAC²

0257. *Omophron oblongiusculum* Chevrolat, 1835
CHIS², MEX⁵, OAX^{1,2}, VER⁴

0258. *Omophron sphaericum* Chevrolat, 1835
VER⁴

11. Pasimachini

0259. *Pasimachus ambiguus* Bänninger, 1950
COL^{1,2}, JAL^{3,5}, MICH⁵⁹, NAY^{1,3,5}, QRO⁵, SIN^{1,2}

0260. *Pasimachus atronitens* Casey, 1913
CHIH^{1,5}, SON⁵

0261. *Pasimachus aurocinctus* Chaudoir, 1850
CHIH¹, CHIS⁵, OAX^{1,15}

0262. *Pasimachus californicus* Chaudoir, 1850
AGS³, CHIH^{1,3,6,7}, COAH^{1,2}, DGO^{1,2,3,4,5}, GTO^{3,4,15},
JAL^{1,3,4,5,6}, MEX³, MICH^{3,4,5,6}, NAY⁵, NL^{1,3,4}, PUE³,
QRO³, SLP^{3,7}, SON^{1,2,3,4,5,6,7,54}, ZAC^{1,4}

0263. *Pasimachus cardioderus* Chaudoir, 1880
CHIS^{3,4}

0264. *Pasimachus cuestai* Kohlmann, 1993
OAX^{1,5}, PUE^{1,4}

0265. *Pasimachus duplicatus* LeConte, 1853
COAH^{3,6}

0266. *Pasimachus elongatus* LeConte, 1846
COAH³, NL³, SIN³

0267. *Pasimachus imitator* Bänninger, 1950
NAY⁴

0268. *Pasimachus laevisulcatus* Bates, 1891
DGO^{1,5,19}

0269. *Pasimachus metallicus* (Chaudoir, 1880)
COL^{1,2}, GRO⁴, JAL^{1,3}, MEX³, MICH^{1,3,15}

0270. *Pasimachus mexicanus* Gray, 1832
GTO³, HGO^{1,3,4,5}, MEX³, QRO³, SLP³

0271. *Pasimachus obsoletus* LeConte, 1846
CHIH³, COAH^{3,7}, DGO⁴, SON⁵⁴

0272. *Pasimachus pacificus* Bänninger, 1950
COL⁴, GRO¹, MICH⁴

0273. *Pasimachus perpolitus* Casey, 1913
CHIH^{1,2}, DGO^{1,2}, SIN^{1,2}

0274. *Pasimachus purpuratus purpuratus* (Putzeys, 1845)
CHIS^{1,4}, OAX¹⁵, VER⁴, YUC^{3,8,15}

0275. *Pasimachus purpuratus rodriguezi* (Putzeys, 1866)
CHIS¹, QROO¹, VER¹; “México”³

0276. *Pasimachus quadricollis* Chaudoir, 1880
TAMPS¹; (type locality: “México”, Chaudoir 1880:
lxxxvi); “México” (Blackwelder 1944: 24; Erwin et al.
1977: 4.11)

0277. *Pasimachus quirozi* Flohr, 1887
OAX^{1,2}, VER³

0278. *Pasimachus rotundipennis* Chevrolat, 1834
CAMP^{5,15}, CHIS¹, OAX¹⁵, VER^{1,3,4,5,15}

0279. *Pasimachus sallei* Chaudoir, 1862
HGO²⁴, SLP^{1,5,7}, TAMPS^{1,3,4,5,7}, VER^{1,3,4,5,15}

0280. *Pasimachus sexualis* Bänninger, 1950
CHIS^{1,5,7}

0281. *Pasimachus smithi* Bates, 1891
GRO¹⁹, HGO²⁴, OAX^{1,2}

0282. *Pasimachus subangulatus* (Chaudoir, 1862)
GRO^{1,3,4,5}, HGO⁴, MEX^{3,4,5}, MICH^{3,5}, MOR^{1,3,4,5}

0283. *Pasimachus tolucanus tolucanus* Chaudoir, 1880
CDMX^{4,5}, DGO^{3,4}, HGO⁴, MEX^{4,5,15}, NL⁴, PUE⁵, SLP³

0284. *Pasimachus tolucanus durangoensis* Bänninger,
1950

DGO^{1,2,4,5}, NAY⁵

0285. *Pasimachus velutinus* Van Dyke, 1943
SIN^{1,5}, SON^{1,2,3,5,54}

0286. *Pasimachus viridans* LeConte, 1858; Figure 1E
CHIH^{1,2,3,4,5,6,7}, DGO⁴, SIN^{5,7}, SLP³, SON^{1,2,3,5,6,15,54}

12. Scaritini

0287. *Scarites anthracinus* Dejean, 1831
CHIH⁴, TAB⁴, TAMPS⁴, VER⁴; **note:** most likely
mis-identified or mis-labelled; type locality [English
translation]: “It is located in the vicinity of Buenos Ai-
res and in the southern parts of Brazil.”, Dejean 1831:
492; known only from Argentina (Erwin, 2011a: 77)

0288. *Scarites granulipygus* (Bates, 1891)
TAB¹⁹, VER^{1,19}

0289. *Scarites illustris* Chaudoir, 1881
VER⁴, (Erwin, 2011a: 83)
0290. *Scarites lissopterus* Chaudoir, 1880
COAH^{1,2,3}, SLP⁵, TAMPS^{1,2,3}
0291. *Scarites marinus* Nichols, 1986
CAMP^{1,2,3}, QROO^{1,2,3}
0292. *Scarites octocoelus* (Chaudoir, 1855)
CAMP^{1,2,3}, CHIS^{1,2,3}, JAL^{1,2,3}, TAB^{1,3}, VER^{1,2,3}, YUC⁵⁹
0293. *Scarites subterraneus* Fabricius, 1775
BCN^{3,5}, BCS⁵, COAH^{1,2,3}, DGO^{1,2,3}, HGO³, MEX³,
NAY³, SIN¹, SLP^{1,2,3}, SON^{1,2,5,54}, VER^{3,4}

13. Clivinini

Note: All states with a superscript ^H for *Ardistomis* species refer to Erwin 2011a.

0294. *Ardistomis convexa* Putzeys, 1867
CHIS^{2,3,7}, NAY⁵, VER⁵⁹
0295. *Ardistomis educta* Bates, 1881
CHIS^{2,3}, CDMX⁵, GRO^{2,3}, NAY⁵, SIN⁵, TAB^H
0296. *Ardistomis mannerheimi* Putzeys, 1846
JAL^H, SIN^H
0297. *Ardistomis muelleri* (Kult, 1950)
CHIS^H, HGO^H, OAX^v, PUE^H
0298. *Ardistomis quixotei* Valdés, 2007
QRO^H, TAB^H, VER^H
0299. *Ardistomis rotundipennis* Putzeys, 1867
CDMX⁴; **note:** most likely mis-identified or mis-labeled; type locality: “Amerique centrale”, Putzeys 1866: 207; “St. Vincent and Grenadines”, Valdés 2009: 71
Note: All states with a superscript ^H for *Aspidoglossa* species refer to Erwin 2011a.
0300. *Aspidoglossa curta* Putzeys, 1866
CHIS^H; (type locality: “province de Ste-Catherine, colonie de Blumenau”, Putzeys 1866: 205)
- 0301[#]. *Aspidoglossa distincta* Putzeys, 1867
CAMP^H, YUC^H
0302. *Aspidoglossa mexicana* (Chaudoir, 1837)
CHIS^{2,3,7}, COL^{2,3}, GRO^{2,3}, JAL^{2,3}, NAY^{2,3}, NL⁷, OAX^{2,3},
PUE^{2,3,5}, SIN^{2,3}, SLP⁷, TAB^{2,3,4}, TAMPS⁴, VER^{3,4},
YUC^H
0303. *Aspidoglossa subangulata* (Chaudoir, 1843)
CHIS^H, CHIH^H, COAH^{2,3}, COL^H, GRO^{2,3,5}, MEX⁵,
MICH^{2,3}, MOR^{2,3,5}, NAY⁵, NL^{2,3}, OAX⁵, PUE^{2,3},
QRO^{2,3}, QROO^H, SIN^H, SLP^{2,3,5}, SON^{5,54}, TAMPS^{2,3},
VER^{2,3}
0304. *Aspidoglossa torrida* Putzeys, 1846
CHIS^H, GRO^H, NAY^H, SLP^H, TAMPS^H
Note: All states with a superscript ^H for *Clivina* species refer to Erwin 2011a.
0305. *Clivina adstricta* Putzeys, 1866
CAMP^{2,3}, GTO⁵, HGO^{2,3}, SLP^{2,3}
0306. *Clivina biguttata* Putzeys, 1866
CHIS^H; (type locality: “Havana”, Putzeys 1866: 158)
0307. *Clivina bipustulata* (Fabricius, 1798)
JAL⁶, SIN⁵, SON⁶, TAB^H, VER^H
0308. *Clivina bituberculata* Putzeys, 1866
Type locality: “Mexico”, Putzeys 1866: 161
0309. *Clivina bolivari* Barr, 1967
TAMPS^{1,3}

0310. *Clivina brunnea* Putzeys, 1846
CHIS^{2,3}, GRO^{2,3}, GTO⁴, JAL^{2,3}, MOR^{2,3}, NAY^{2,3},
OAX^{2,3}, TAB^{2,3}, VER⁴
0311. *Clivina brunneipennis* Putzeys, 1846
GRO^H, MOR^H, VER⁴
0312. *Clivina cruciata* Putzeys, 1866
SLP⁷, VER^{59,H}
0313. *Clivina dentifemorata* Putzeys, 1846
CHIS^{2,3,7}, COL^H, GRO^H, GTO^H, JAL^H, NL⁷, OAX^{2,3},
PUE⁴, SLP^{2,3,7}, TAB^{2,3}, TAMPS^{2,3}, VER^{2,3,4}
0314. *Clivina dentipes* Dejean, 1825
CHIS^{2,3}, GTO^{2,3}, JAL^{2,3,6}, MOR^{2,3,6}, NAY^{2,3,5}, OAX^{2,3},
PUE^{2,3}, SLP^{2,3}, SON^{3,6}, TAMPS^{2,3}
- 0315[#]. *Clivina dilutipennis* Putzeys, 1866
VER^{59,H} “México” (Blackwelder, 1944:24; Erwin et al., 1977:4.11)
0316. *Clivina distigma* Putzeys, 1866
VER^{3,4}
0317. *Clivina fasciata* Putzeys, 1846
CAMP^H, OAX^H, TAB⁴, TAMPS^{2,3}, VER^{2,3}, YUC⁵⁹
0318. *Clivina ferrea* LeConte, 1857
BCN^{3,6}, BCS^{3,5,6}, PUE^H, SON^{3,6}, VER^{2,3}
0319. *Clivina limbipennis* Jacquelin du Val, 1857
COAH^H, NL⁷, SLP^H, TAMPS^H, VER²
0320. *Clivina lucida* Putzeys, 1866
CHIS^{2,3}, COAH^{2,3}, OAX^{2,3}, SON^{2,3,54}, TAB^{2,3}
0321. *Clivina marginipennis* Putzeys, 1846
CHIS^{2,3}, COL^{2,3}, GTO^{2,3}, JAL^{2,3}, MICH^{2,3}, MOR^{2,3},
VER^{2,3}, YUC^{59,H}
0322. *Clivina parvidens* Putzeys, 1866
CAMP^{2,3}, CHIS^{2,3}, VER^{59,H}
0323. *Clivina planicollis* LeConte, 1857
COL³, JAL^H, MOR³, NAY^H, NL³, OAX³, QRO³, SLP³,
TAMPS³, VER³
- 0324[#]. *Clivina planulata* Putzeys, 1866
Type locality: “México”, Putzeys 1866: 140; “México” Blackwelder 1944: 26; Erwin et al. 1977: 4.13); “México”^H
Note: All states with a superscript ^H for *Halocoryza* species refer to Erwin 2011b.
0325. *Halocoryza acapulcana* Whitehead, 1966
GRO^{2,3,5}, OAX^H, QROO^{2,3}
0326. *Halocoryza arenaria* Darlington, 1939
QRO^H, YUC^H
0327. *Halocoryza whiteheadiana* Erwin, 2011
BCS⁵⁹; type locality: “Baja California Sur, Isla Carmen, north end” (Erwin 2011b: 7)
0328. *Oxydrepanus mexicanus* Putzeys, 1866
CHIS^{2,3}
Note: Erwin’s superscript ¹ for *Schizogenius* species refers to Whitehead 1972.
0329. *Schizogenius auripennis* Bates, 1881
BCN^{1,3,6,55}, BCS^{1,5}, CHIS^{1,55}, HGO²⁷, JAL^{1,2,3,55},
NAY^{1,5,55}, PUE^{1,2,3,55}, SIN^{1,5,55}, SON^{1,2,3,5,6,54,55}, VER³
0330. *Schizogenius brevisetosus* Whitehead, 1972
COAH^{1,55}
0331. *Schizogenius chiricahuensis* Whitehead, 1972
CHIH^{3,6}

0332. *Schizogenius crenulatus crenulatus* LeConte, 1852
JAL^{2,3}, NAY^{1,55}, SIN^{1,2,3,5,55}, SON^{1,55}
0333. *Schizogenius crenulatus chiapatecus* Whitehead, 1972
CHIS^{1,2,3,55}, GRO^{1,3,55}, JAL¹, OAX^{1,55}
0334. *Schizogenius depressus* LeConte, 1852
CHIH^{1,2,3,55}, GTO^{1,55}, MEX^{4,55}, ZAC^{1,2,3,55}
0335. *Schizogenius dilatus* Whitehead, 1972
NL^{1,2,3,5,55}, TAMPS^{1,55}
0336. *Schizogenius falli* Whitehead, 1972
BCN^{1,2,3,5,6,55}, BCS^{2,3,5}, CHIH^{1,2,3,55}, COAH^{1,2,3,55},
COL^{1,2,3,55}, DGO^{1,2,3,55}, NAY^{1,2,3,5,55}, NL^{1,2,3,5,8,55},
SIN^{1,2,3,5,55}, SON^{1,2,3,5,6,8,54,55}, TAMPS^{1,55}, ZAC^{1,2,3,5,55}
0337. *Schizogenius ferrugineus* Putzeys, 1846
SON^{3,6}
0338. *Schizogenius kulti* Whitehead, 1972
CHIS^{1,2,3,5,55}, GRO^{1,2,3,55}, JAL^{1,2,3,55}, NAY^{1,2,3,55},
OAX^{1,2,3,5,55}, TAB⁵⁵, VER^{1,3,5,55}, ZAC^{1,2,3,55}
0339. *Schizogenius lineolatus* (Say, 1823)
TAMPS^{1,2,3,55}
0340. *Schizogenius longipennis* Putzeys, 1866; Figure 1F
CHIS^{1,2,3,55}, COAH⁵, COL^{1,2,3,5,55}, GRO^{1,2,3,55}, JA^{1,2,3,5,6,55},
MICH^{1,2,3,55}, MOR^{1,2,3,4,55}, NAY^{1,2,3,5,55}, NL^{1,2,3,55},
OAX^{1,2,3,4,55}, PUE^{1,2,3,55}, QRO^{1,2,3,4,55}, SIN^{1,2,3,4,5,55},
SLP^{1,2,3,55}, SON^{1,2,3,5,54,55}, TAB^{1,2,3,55}, TAMPS^{1,2,3,55},
VER^{1,2,3,5,55}
0341. *Schizogenius multisetosus* Bates, 1891
BCS^{3,5}, GRO^{1,3,5,55}, HGO^{2,3,55}, MOR^{1,2,3,7,19,55},
OAX^{1,2,3,5,19,55}, PUE^{1,2,55}, SLP^{1,2,3,5,55}
0342. *Schizogenius neovalidus* Whitehead, 1972
SON^{2,3,6,54}
0343. *Schizogenius optimus* Bates, 1881
CHIS^{1,2,3,5,55}
0344. *Schizogenius pacificus* Whitehead, 1972
GRO^{1,3,5,55}, JAL^{1,2,3}, NAY^{1,2,3}, OAX^{1,2,3,4,5,55}, SIN^{1,2,3,5,55}
0345. *Schizogenius pluripunctatus* LeConte, 1852
DGO^{1,2,3,55}, NAY^{1,2,3,5,55}, SIN^{1,5,55}, SON^{1,2,3,54,55}
0346. *Schizogenius plurisetosus* Whitehead, 1972
BCN², NL^{1,2,55}, TAMPS^{1,5,55}
0347. *Schizogenius pygmaeus* Van Dyke, 1925
AGS^{1,2,3,55}, BCN^{1,6,55}, BCS⁵, CHIH^{1,55}, CHIS^{1,2,3,4,55},
COAH¹, COL^{1,2,3}, DGO^{1,5,55}, GRO^{1,2,3,55}, GTO^{1,2,3,55},
HGO^{1,2,3,55}, JAL^{1,2,3,5,55}, MEX^{1,55}, MICH^{1,2,3,5,55}, MOR^{1,55},
NAY^{1,2,3,55}, NL^{1,2,3,55}, OAX^{1,2,3,55}, PUE^{1,2,3,5,55}, QRO^{1,2,3,55},
SIN^{1,2,3,5,55}, SLP^{1,2,3,4,5,55}, SON^{1,2,3,5,6,54,55}, TAMPS^{1,2,3,55},
VER^{1,2,3,55}, ZAC^{1,2,3,55}
0348. *Schizogenius scopaeus* Whitehead, 1972
NL^{1,2,3,4,55}, TAMPS^{1,2,3,4,55}
0349. *Schizogenius sculptilis* Whitehead, 1972
CHIS^{1,2,5,3,55}, TAB^{1,55}
0350. *Schizogenius seticollis seticollis* Fall, 1901
BCN¹, BCS^{1,2}
0351. *Schizogenius seticollis vandykei* Whitehead, 1972
BCN^{5,55}, BCS⁵
0352. *Schizogenius tenuis* Bates, 1881
CHIS^{1,2,55}, DGO^{1,55}, JAL^{1,2,55}, NAY^{1,5,55}, OAX^{1,2,5,55},
PUE^{1,2,55}, SIN⁵⁵, SLP^{1,3,55}, SON^{1,55}, TAB^{1,55}, VER^{1,2,3,55}
0353. *Schizogenius tibialis* Whitehead, 1972
CHIH¹, CHIS^{1,55}, COAH⁵⁵, GRO^{1,55}, JAL^{1,2,55}, NL^{1,55},
OAX^{1,2,7,55}, SLP^{1,2,3,5,7,55}, TAMPS^{1,2,55}, VER^{1,55}
0354. *Schizogenius tristriatus* Putzeys, 1846
QRO^{1,2,55}, SLP⁵⁵, VER^{1,2,5,55}
0355. *Schizogenius truquii* Putzeys, 1846
JAL^{1,2,5,55}, MICH⁵⁵, MOR^{1,55}, SIN^{1,55}
0356. *Semiardistomis cyaneolimbatus* (Chevrolat, 1863)
CHIS⁷, DGO^{1,2}
0357. *Semiardistomis labialis* (Chaudoir, 1837)
CDMX⁵, CHIS^{1,2,5}, COAH⁵, COL¹, GRO^{1,5}, JAL^{1,5,7},
MICH^{1,2}, MOR^{1,2}, NAY^{1,2,5}, NL^{1,2}, OAX^{1,2}, PUE^{1,2,4},
QRO^{1,2}, SIN^{1,5}, SLP^{1,2,5}, TAB¹, TAMPS¹, VER^{1,3,4,5}
0358. *Semiardistomis muelleri* (Kult, 1950)
HGO²⁴, PUE⁵⁹
0359. *Semiardistomis pallipes* (Dejean, 1831)
CHIS^{1,2}
0360. *Semiardistomis propinquus* (Putzeys, 1866)
AGS^{1,2}, CDMX⁵, CHIH⁵, DGO², GTO^{1,2}, GRO^{1,2},
JAL^{1,2,5}, MEX^{1,2}, MICH^{1,2,5}, MOR^{1,2,5}, NAY⁵, OAX^{1,2,5},
PUE^{4,5}, SON^{1,54}
0361. *Semiardistomis viridis* (Say, 1823)
BCS³, CHIS³, GRO³, JAL³, MOR³, NL³, OAX³,
SLP^{1,2,3,5,7}, TAMPS^{1,2,3}, VER^{1,2,3}, YUC³

14. Dyschiriini

0362. *Akephorus marinus* (LeConte, 1852)
BCN^{5,21,26}
0363. *Dyschirius abbreviatus* Putzeys, 1846
CHIS¹, COAH^{1,2}, SLP⁵, VER³, YUC⁵⁹
0364. *Dyschirius affinis* Fall, 1901
NL^{1,2}, SLP³, TAMPS^{1,2}
0365. *Dyschirius analis* LeConte, 1852
BCS³, CHIH^{1,2}, CHIS^{1,2}, COAH⁵, GTO^{1,2}, HGO^{1,2},
MOR^{1,2}, NAY^{1,2}, OAX^{1,2}, SIN^{1,2,5}, SON^{1,2,3,5,6,54},
TAMPS^{1,2}, VER¹
0366. *Dyschirius aratus* LeConte, 1852
CHIS¹, GTO^{1,2}, NAY⁵, SIN^{1,2,5}, SON^{5,54}
- 0367*. *Dyschirius darlingtoni* Kult, 1950
VER¹; (type locality: "Goias, Brazil", Kult 1950: 311)
- 0368*. *Dyschirius erythrocerus* LeConte, 1857
Type locality: "Ohio; Pennsylvania", LeConte 1857:
78; "México, Cuba" (Erwin et al. 1977: 4.12)
0369. *Dyschirius interior* Fall, 1922
COAH³
0370. *Dyschirius larochellei* Bousquet, 1988
YUC^{1,2}
0371. *Dyschirius nigricornis* Motschulsky, 1844
BCN^{3,6}, BCS^{3,6}, SON^{3,6}
0372. *Dyschirius patruelis* LeConte, 1852
BCN²⁶; type locality: "San Diego, sat frequens" Le-
conte 1852: 196
- 0373#. *Dyschirius planiusculus* Putzeys, 1866
Type locality: "México", Putzeys 1866: 39; "México"
(Blackwelder 1944: 26; Erwin et al. 1977: 4.12)
0374. *Dyschirius politus* (Dejean, 1825)
SON^{5,54}
- 0375*. *Dyschirius sublaevis* Putzeys, 1846
YUC¹; (type locality: "Texas", Putzeys 1846b: 563);
"Cuba, U.S.A." (Blackwelder 1944: 26); "México"
(Erwin et al. 1977:4.12)

0376. *Dyschirius terminatus* LeConte, 1848
CHIS^{1,2}, GRO³, JAL^{1,2,3}, NAY⁵, OAX^{1,2,3}, SIN^{1,2}, SLP^{1,2},
SON^{5,54}
- 0377*. *Dyschirius truncatus* LeConte, 1857
BCN¹; (type locality: “Illinois”, LeConte 1857: 78)

15. Broscini

0378. *Rawlinsius papillatus* Davidson and Ball, 1998
GRO^{1,2,4}

16. Trechini

0379. *Chiapadytes bolivari* Vigna Taglianti, 1977
CHIS^{1,14}
0380. *Mexaphaenops elegans* Barr, 1967
QRO^{3,14}
0381. *Mexaphaenops febriculosus* Barr, 1982
TAMPS¹⁴
0382. *Mexaphaenops fishi* Barr, 1967
SLP^{3,14}; **note:** Lorenz (2005: 180, and 2021) lists this species as *fiski*, but that is incorrect, as Barr (1967:162) spelled the species name as *fishi*, naming the species after one of the collectors – John Fish.
- 0383#. *Mexaphaenops intermedius* Barr, 1971
TAMPS^{1,14}
- 0384#. *Mexaphaenops jamesoni* Barr, 1982
TAMPS¹⁴
0385. *Mexaphaenops mackenziei mackenziei* Barr, 1982
NL⁵¹, TAMPS¹⁴
0386. *Mexaphaenops mackenziei dulcinominis* Barr, 1982
NL^{14,51}
0387. *Mexaphaenops mackenziei gracilis* Barr, 1982
NL⁵¹, TAMPS¹⁴
0388. *Mexaphaenops prietoi* Bolívar y Pieltain, 1942
NL^{14,53}
- 0389#. *Mexaphaenops sulcifrons* Barr, 1982
TAMPS¹⁴
- 0390#. *Mexitrechus coarctatus* (Bates, 1882)
OAX^{1,14}
0391. *Mexitrechus michoacanus* (Bolívar y Pieltain, 1941)
AGS³, GRO¹⁴, MEX¹⁴, MICH^{1,4,14}
- 0392#. *Mexitrechus mogotensis* Barr, 1982
GRO^{14,51}
- 0393#. *Mexitrechus occidentalis* (Mateu, 1974)
DGO^{1,14}
0394. *Mexitrechus quirogai* (Bolívar y Pieltain, 1943)
HGO⁴, MICH^{1,4,14}
0395. *Mexitrechus tepoztilanensis* (Bolívar y Pieltain, 1943)
GRO^{1,14}, MOR^{1,4,14}, MEX¹
- 0396#. *Paratrechus altitudinis* Mateu, 1974
HGO²⁷, MEX^{1,14}
- 0397#. *Paratrechus balli* Barr, 1982
OAX¹⁴
0398. *Paratrechus batesi* Mateu, 1974
VER^{1,2,3,14}
- 0399#. *Paratrechus beltrani* Mateu, 1974
MEX^{1,14}
0400. *Paratrechus bifoveatus* (Jeannel, 1920)
CHIS^{1,14}

- 0401#. *Paratrechus bolivari* Mateu, 1974
MICH^{1,14}, MEX¹
- 0402#. *Paratrechus cataractae* Barr, 1982
VER¹⁴
- 0403#. *Paratrechus contrarius* Barr, 1982
QRO¹⁴
0404. *Paratrechus erwini* Barr, 1982
VER^{3,14}
0405. *Paratrechus hoegei* (Jeannel, 1920)
OAX^{1,4,14}
- 0406#. *Paratrechus jeanneli* Barr, 1982
OAX¹⁴
- 0407#. *Paratrechus laticeps* Barr, 1982
TAMPS¹⁴
0408. *Paratrechus mexicanus* (Putzeys), 1870
VER^{1,14}
- 0409#. *Paratrechus oaxaquensis* Barr, 1982
OAX¹⁴
0410. *Paratrechus osorioi* Bolívar y Pieltain, 1943
HGO^{1,4,14}
0411. *Paratrechus pallescens* (Barr, 1967) (**note:** GBIF has Bolívar y Pieltain as the author)
QRO¹⁴
- 0412#. *Paratrechus propior* Barr, 1982
HGO¹⁴
- 0413#. *Paratrechus putzeysi* Barr, 1982
OAX¹⁴
- 0414#. *Paratrechus reddelli* Barr, 1982
VER¹⁴
- 0415#. *Paratrechus sylvarum* Mateu, 1974
MEX^{1,14}
0416. *Paratrechus sylvaticus* (Bolívar y Pieltain, 1941)
CDMX¹, MOR^{1,4,14}
- 0417#. *Paratrechus tacana* Barr, 1982
CHIS¹⁴
- 0418#. *Paratrechus totontepec* Barr, 1982
OAX¹⁴
0419. *Trechus aztec aztec* Jeannel, 1920
CDMX⁴, GRO^{1,4}, MEX^{1,4}, MOR⁴
- 0420#. *Trechus aztec rotgeri* Mateu, 1974
MEX^{1,14}
- 0421#. *Trechus aztec zempoala* Mateu, 1974
MOR¹; (type locality: “Parque Nacional Lagunas de Zempoala, [MOR⁴⁷ and MEX⁴⁷], México”, Mateu 1974b: 188)
- 0422#. *Trechus hendrichsi* Mateu, 1974
MEX^{1,14}
- 0423#. *Trechus moctezuma moctezuma* Mateu, 1974
MEX^{1,14}
- 0424#. *Trechus moctezuma lachicai* Mateu, 1974
MEX⁴⁷, PUE¹; (type locality: “Salazar, [Mexico state, México]”, Mateu 1974b: 189)
0425. *Trechus tolucensis* Bolívar y Pieltain, 1941
MEX^{1,4,14}

17. Bembidiini

0426. *Bembidion affine* Say, 1823
SON³

0427. *Bembidion aratum* (LeConte, 1852)
CHIH², SON⁵⁴
0428. *Bembidion championi* Bates, 1882
CHIS¹, DGO², GUER², JAL², MEX^{1,2}, MICH^{1,2}, MOR¹, SON²
0429. *Bembidion cheyennense* Casey, 1918
CHIH², JAL², SON⁵⁴
- 0430#. *Bembidion citulum* Casey, 1918
CHIH⁵⁹; (type locality: "México (Colonia Garcia, Sierra Madre Mts., Chihuahua)", Casey 1918a: 66)
0431. *Bembidion ciudadense* Bates, 1891
CHIH², DGO¹⁹
0432. *Bembidion cognatum* Dejean, 1831
AGS², CHIS², COAH⁷, DGO², GRO², GTO^{2,4}, HDO², JAL², MEX², MICH², MOR², NL⁷, OAX², QRO², SLP², SON⁵⁴, TLAX², ZAC²
0433. *Bembidion coloradense* Hayward, 1897
DGO²; (type locality: "mountains of southwestern Colorado", Hayward 1897: 98); (Lectotype, "vicinity of Rico, Dolores County, Colorado" (Erwin 1984: 169))
0434. *Bembidion consanguineum* Hayward, 1897
BCN^{21,26}
0435. *Bembidion constricticollae* Hayward, 1897
CHIH², COAH³
0436. *Bembidion constrictum* LeConte, 1848
COAH², TAMPS², YUC²
0437. *Bembidion cortes* Erwin, 1982
VER (Erwin, 1982: 481); (type locality: "La Lima, Cortes, Honduras", Erwin 1982: 481)
0438. *Bembidion cyclodes* Bates, 1884
DGO², GTO², MEX², MOR¹, OAX^{1,2,18}, PUE²
- 0439#. *Bembidion dannieae* Perrault, 1982
OAX^{1,2}; (type locality: "Sierra Madre del Sur, Oaxaca, [México]", Perrault 1982: 102)
0440. *Bembidion durangoense* Bates, 1891
DGO^{1,19}, SIN²
0441. *Bembidion egens* Casey, 1918
BCS², CHIS², COAH², GRO², JAL², MEX², MOR², NAY², OAX², PUE², QRO², SIN², SLP², SON^{2,54}, ZAC²
- 0442#. *Bembidion elescarabajo* Maddison, 2023; Figure 1G
OAX²; (type locality: "México: Oaxaca: 28 miles (45 km) south of Valle Nacional (San Juan Bautista Valle Nacional along route 175; this is approximately 17.594°N 96.427°W", Maddison 2023: 443)
0443. *Bembidion ephippigerum* LeConte, 1852
BCN²
- 0444#. *Bembidion erwini* Perrault, 1982
OAX¹; (type locality: "Juchatengo, Oaxaca (Mexico)", Perrault 1982: 103)
0445. *Bembidion flohri* Bates, 1878
PUE⁴; (type locality: "México, near the capital", Bates 1878: 602)
0446. *Bembidion franiae* Erwin, 1982
CHIS^{2,3}
0447. *Bembidion georgettae* Perrault, 1982
GRO², MEX², MICH², SON², VER^{1,2}
0448. *Bembidion horni* Hayward, 1897
BCN^{2,5}
0449. *Bembidion impotens* Casey, 1918
AGS², BCN², CHIH^{2,3,6}, COAH^{2,3}, DGO², GTO², HGO^{2,4}, JAL², MICH², NL², QRO², SLP², TAMPS², VER^{2,3}
0450. *Bembidion insulatum* (LeConte, 1852)
BCN^{21,26}
0451. *Bembidion integrum* Casey, 1918
NL²
0452. *Bembidion jucundum* G. Horn, 1895
BCS^{3,5}
0453. *Bembidion lacunarium* Zimmermann, 1869
COAH²; (type locality: "Middle States", Zimmermann 1869: 248)
0454. *Bembidion lecontei* Csiki, 1928
GTO²; *Bembidion lecontei* Csiki 1928: 62 is a replacement name for *B. grandicollae* LeConte 1851: 189, (type locality: "San Diego, California"), [nec. Motschulsky 1850: 14]
0455. *Bembidion levigatum* Say, 1823
CHIH⁴
0456. *Bembidion lugubre* LeConte, 1857
SON^{2,54}
0457. *Bembidion luculentum* Casey, 1918
VER²; (type locality: "Florida (Indian River)", Casey 1918a: 122)
0458. *Bembidion macrogonum* Bates, 1891
CHIS², GRO², MICH², OAX², VER^{2,3,19,52}
0459. *Bembidion mexicanum* Dejean, 1831
BCN¹, JAL⁴, NL⁷, PUE⁴, SON^{2,3,6,54}, VER^{3,52}
0460. *Bembidion nahuala* Erwin, 1982
CHIS^{2,3}, GRO², HGO², JAL², MEX², MICH², OAX², PUE², VER²
0461. *Bembidion nevadense* Ulke, 1875
BCN²
0462. *Bembidion nogalesium* Casey, 1924
SON⁵⁴; (type locality: "Nogales, Arizona", Casey 1924: 42); "México"²¹
- 0463*. *Bembidion nubiculosum* Chaudoir, 1868)
BCN¹; (*Bembidion nubiculosum* Chaudoir 1868: 244 is a replacement name for *B. laticollae* LeConte 1852: 187 (type locality: "Colorado [River, California]"))
0464. *Bembidion perspicuum* LeConte, 1848
SON^{2,3,54}
0465. *Bembidion placitum* Bates, 1878
CDMX⁴, PUE², TLAX²
0466. *Bembidion poculare* Bates, 1884
AGS¹, COAH³, COL^{1,2}, JAL², MICH^{1,2}, OAX^{1,2}, PUE¹, SLP^{1,2}, SON^{2,54}
- 0467*. *Bembidion quadrimaculatum dubitans* (LeConte, 1852)
BCN¹; (type locality: "Ad San Francisco [California]", LeConte 1852: 189)
0468. *Bembidion quadrulum* LeConte, 1861
CHIH², SON²
0469. *Bembidion rapidum* (LeConte, 1847)
COAH^{2,3,7}, DGO², GTO², NL^{2,7}, QRO², SLP², TAMPS²
0470. *Bembidion rupicola* (Kirby, 1837)
COAH², NL², PUE², ZAC²

0471. *Bembidion scintillans* Bates, 1882
CHIH², CHIS², JAL², NAY², OAX², PUE², SIN²
0472. *Bembidion semifasciatum* Say, 1834
DGO², JAL², MEX², MICH², NAY², SIN², SLP², SON²
- 0473*. *Bembidion semiopacum* Casey, 1924
Type locality: “California (San Joaquin Co.”, Casey 1924: 39; “known only from California central valley (Erwin 1984a: 182)” (Bousquet 2012: 617)
0474. *Bembidion sordidum* Kirby, 1837
CHIH²
0475. *Bembidion sparsum* Bates, 1882
AGS², CAMP², CHIS², COAH², COL², GRO², JAL², MEX⁴, NAY², OAX², QRO², SIN², SLP², SON⁴, TAB^{2,4}, VER^{2,3,16}
0476. *Bembidion sphaeroderum* Bates, 1882
CHIH², DGO^{1,2}, HGO¹, MEX^{1,2}, NL^{1,2}, OAX¹, PUE⁴, QRO^{1,2}, SLP¹, VER¹⁶
0477. *Bembidion sphaeruliferum* Bates, 1891
CHIH¹, DGO^{1,2,19}, JAL^{1,2}, MEX^{1,2}, MICH^{1,2}, TLAX², VER⁶
0478. *Bembidion spretum* Dejean, 1831
PUE^{4,7}, TAMPS², VER²
0479. *Bembidion stephensii* Crotch, 1866
SON⁶
0480. *Bembidion subangustatum* Hayward, 1897
CHIH², DGO², MEX², MICH², NL², OAX²
0481. *Bembidion submaculatum* Bates, 1878
CDMX¹⁶, CHIH², DGO², GRO², HGO^{2,16}, JAL², MEX², MICH², OAX^{2,16}, QRO², TLAX², VER^{2,3,16}
0482. *Bembidion texanum* Chaudoir, 1868
HGO², NL², QRO², SLP², TAMPS²
0483. *Bembidion transversale* Dejean, 1831
BCN², CHIH², DGO², SON²
0484. *Bembidion vernale* Bates, 1882
CHIS², GRO², HGO², PUE^{2,4}, QRO², VER²
- 0485*. *Bembidion vile* (LeConte, 1852)
“México”¹; (type locality: “San Diego [California]”, LeConte 1852: 189)
0486. *Bembidion viridicolle* (LaFerté-Sénéctère, 1841)
COAH³
0487. *Elaphropus conjugens* (Notman, 1919)
SON⁵⁴; (type locality: “Rincon Mts., Ariz.”, Notman 1919: 229)
0488. *Elaphropus cruciatus* (Chaudoir, 1868)
MICH⁴, PUE⁴, TAB⁴, VER²
0489. *Elaphropus dolosus* (LeConte, 1848)
BCS^{21,26}
0490. *Elaphropus mellitus* (Casey, 1918)
BCS⁵, COAH⁵, COL⁵, DGO⁵⁹, GRO⁵, JAL⁵, NAY⁵, NL⁵, PUE⁵, SIN⁵, SON^{5,54}, VER⁵
0491. *Elaphropus microspilus* (Bates, 1882)
VER³
0492. *Elaphropus mundulus* (Bates, 1882)
SLP³, TAMPS³, VER^{3,16}
0493. *Elaphropus nebulosus* (Chaudoir, 1868)
SLP⁵, TAMPS³, VER⁵
0494. *Elaphropus obesulus* (LeConte, 1852)
SON^{5,6,54}, TAMPS³
0495. *Elaphropus obtusellus* (Bates, 1882)
VER¹⁶
0496. *Elaphropus parvulus* (Dejean, 1831)
MOR⁵
0497. *Elaphropus pericallis* (Bates, 1882)
CHIS³, NL^{3,5}, SLP^{3,5}, VER²
0498. *Elaphropus purgatus* (Bates, 1882)
GRO⁴, PUE^{4,16}, VER^{3,16}
- 0499#. *Elaphropus unistriatus* (Bilimek, 1867)
GRO⁵⁹, MEX¹; (type locality: “Auf trockenen Stalagmiten schnell herumlaufend in der Höhle Cacahuamilpa in México” [= running on dry stalagmites in cave of Cacahuamilpa in México [= Guerrero]], Bilimek 1867: 902)
0500. *Elaphropus xanthopus* (Dejean, 1831)
TAB², VER³
- 0501*. *Erwiniana ampliata* (Bates, 1884)
GRO¹; (type locality: “Panama, Bugaba”, Bates 1884: 290)
- 0502*. *Erwiniana belti* (Bates, 1878)
VER¹; (type locality: “Chontales, Nicaragua”, Bates 1878: 601)
- 0503#. *Geocharidius zullinii* Vigna Taglianti, 1973
CHIS^{1,59}
0504. *Lionepha erasa* (LeConte, 1859)
CHIH⁴
0505. *Meotachys amplicollis* (Bates, 1882)
TAB¹⁶, VER^{3,16}
- 0506#. *Mexanillus sbordonii* Vigna Taglianti, 1973
CHIS^{1,59}
0507. *Mioptachys autumnalis* (Bates, 1882)
OAX¹⁶, VER¹⁶
0508. *Mioptachys oopertoides* Bates, 1882
CHIS³
- 0509*. *Mioptachys xanthura* (Bates, 1871)
Type locality: “Rio de Janeiro” [Brazil], Bates 1871: 12; “México” (Blackwelder 1944: 31)
Note: Erwin’s superscript ¹ for *Pericompsus* species refers to Erwin 1974.
0510. *Pericompsus brasiliensis* (R.F. Sahlberg, 1844)
CAMP^{1,2,30}, TAB^{1,2,30}
0511. *Pericompsus ephippiatus* (Say, 1834)
CHIS^{2,30}, GRO^{2,30}, JAL^{2,30}, OAX^{2,30}, SLP^{5,30}, TAMPS³
0512. *Pericompsus gongylus* Erwin, 1974
CAMP^{1,2}
0513. *Pericompsus gracilior* (Bates, 1884)
CHIS^{1,2}, CDMX³⁰, GRO^{1,2,3}, JAL^{1,2,3}, MOR^{18,30}, OAX^{1,2}, SIN^{2,3,30}, SLP^{1,2}, TAB³⁰, VER^{5,30}
0514. *Pericompsus immaculatus* Bates, 1871
VER^{1,2}
0515. *Pericompsus jucundus* Schaum, 1859
SIN², SLP²
0516. *Pericompsus laetulus* LeConte, 1852
DGO^{1,2}, SON^{1,5,54}
- 0517#. *Pericompsus leechi* Erwin, 1974
NAY¹; (type locality: “Five miles south of Rio Grande de Santiago Ferry, Nayarit, México”, Erwin 1974: 46)
0518. *Pericompsus leucocareus* Erwin, 1974
MEX¹, MICH¹, SLP², TAMPS^{1,2}, VER^{1,2}

0519. *Pericompsus longulus* Bates, 1878
CDMX¹, DGO¹, MEX^{1,2}, OAX¹
- 0520#. *Pericompsus pauli* Erwin, 1974
CHIS¹; (type locality: “Near Pijijiapan, Chiapas, México”, Erwin 1974: 47)
0521. *Pericompsus reichei* (Putzeys, 1845)
CHIS^{1,2}, SLP^{1,2,5}, TAB¹, VER^{1,5}
0522. *Pericompsus sigma* Erwin, 1974
CHIS^{1,2}, COL^{1,2}, GRO^{1,2}, VER^{1,5}
0523. *Pericompsus sellatus* LeConte, 1852
NAY⁵, SIN³, SON^{2,54}
0524. *Pericompsus tlaloc* Erwin, 1974
CHIS^{1,2}, COL^{1,2}, JAL^{1,2}, NAY^{1,2,5}, OAX^{1,2}, SIN^{1,2,5}, SLP^{1,2}, SON^{1,3,5,6,54}, VER^{1,5}, ZAC²
- 0525*. *Tachys antiqua* Erwin, 1971
“Chiapas amber (Upper Oligocene or Lower Miocene), México” (Erwin 1971: 234)
0526. *Tachys corax* LeConte, 1852
BCS^{21,26}
- 0527*. *Tachys latalatus* (Csiki, 1928)
“California” (Csiki 1928: 185); “México”¹, (Erwin et al. 1977: 4.15) (*Tachys latalatus* Csiki 1928: 185, is a replacement name for *T. latipennis* Hayward 1900: 277, type locality “San Estaban, Lower California, México” [= BCS])
0528. *Tachys misellus* (LaFerté-Sénéctère, 1841)
SON⁵⁴; (type locality: “Texas”, LaFerté-Sénéctère 1841: 48); “Texas; Guatemala” (Erwin et al. 1977: 4.15),
0529. *Tachys monostictus* Bates, 1871
VER^{2,3}
0530. *Tachys pallidus* Chaudoir, 1868
BCS³, NL³
0531. *Tachys proximus* (Say, 1823)
COAH⁵³
0532. *Tachys rufotestaceous* Hayward, 1900
BCS⁵, SIN⁵, VER⁵
0533. *Tachys scitulus* LeConte, 1848
VER²
0534. *Tachys ventricosus* LeConte, 1863
TAMPS³
0535. *Tachys virgo* LeConte, 1852
SON^{3,6,54}
0536. *Tachys vittiger* LeConte, 1852
BCS^{3,6}, COAH³, JAL³, SON^{6,54}
- 0537*. *Tachys vorax* (LeConte, 1852)
BCN¹; (type locality: “Ad fluminis Gilae et Colorado ripas” [= confluence of Gila and Colorado Rivers], LeConte 1852: 194)
Note: All states with superscript ¹ for *Tachyta* species refer to Erwin 1975.
0538. *Tachyta angulata* Casey, 1918
DGO¹, NL¹, VER¹
0539. *Tachyta nana inornata* (Say, 1823)
CHIH³, CHIS^{1,3}, COL^{1,5}, HGO¹, MEX¹, NAY⁵, NL^{1,5}, OAX¹, QRO³, SLP³, SON^{2,3,54}, TAMPS³, VER¹

18. Pogonini

0540. *Diplochaetus emaciatus* (Bates, 1891)
BCN¹, SIN^{1,19}
- 0541*. *Diplochaetus rutilus* (Chevrolat, 1863)
“México”²¹; (type locality: “Cuba”, Chevrolat 1863: 197)
- 0542*. *Thalassotrechus barbarae* (G. Horn, 1892)
BCN¹; (type locality: “Santa Barbara, Cal.”, G. Horn 1892: 41)

19. Ozaenini

Note: Erwin’s superscript ¹ for Ozaenini refers to Ball and McCleve 1990.

0543. *Entomoantyx cyanipennis* (Chaudoir, 1852)
CHIS^{1,3,5}, VER^{1,2,4,5}
0544. *Goniotropis kuntzeni kuntzeni* (Bänninger, 1927)
DGO⁵⁹; type locality: “Canelas, Durango, México”, Bänninger 1927: 205; SON^{1,2,54}
0545. *Goniotropis parca* (LeConte, 1884)
BCN¹, BCS⁵, DGO¹, SON^{1,2,3,54}, TAMPS³
0546. *Ozaena lemoulti* Bänninger, 1932
CHIS⁷, MOR⁵, SIN⁵, VER¹
0547. *Pachyteles elongatus* (Chaudoir, 1854)
CHIH¹, CHIS^{2,3}, COL³, GRO³, JAL^{1,2,3}, NAY^{1,2}, SLP¹, TAMPS^{1,2}, VER^{1,2}
0548. *Pachyteles enischnus* Ball and McCleve, 1990
CHIS³, GRO³, HGO²⁴, JAL^{1,2}, NAY⁵, SON^{1,3}
0549. *Pachyteles gyllenhali* (Dejean, 1825)
BCN^{1,5}, BCS^{1,3,5}, CHIH¹, CHIS^{1,2,5}, COL^{1,5}, JAL^{1,2,4}, NAY^{1,2} (Tres Marias Islands), OAX³, PUE²⁷, SIN¹, SON^{1,2,5,54}, TAB^{1,2,3}, VER^{1,2,5}
0550. *Pachyteles mexicanus* (Chaudoir, 1848)
CHIS^{1,2,4}, OAX^{1,2}, PUE^{1,2}, SLP^{1,4,5,7}, TAB⁵, TAMPS^{3,4}, VER^{1,2,3,4,5,7,52}
0551. *Pachyteles oxyomus* Chaudoir, 1868
TAB⁵⁹, VER², YUC⁵⁹
0552. *Pachyteles porrectus* Chaudoir, 1868
CHIS^{1,2}, PUE⁶, SLP⁷
0553. *Pachyteles punctulatus* Chaudoir, 1868
CHIS^{1,2}, SLP⁷, YUC⁵⁹
0554. *Physeia hirta* LeConte, 1853
CHIS^{1,2}, GRO³, OAX^{1,2}, SLP^{1,5}, VER^{1,4}
0555. *Physeia latipes* Schaum, 1864
COL^{1,2}, JAL^{1,2,4}, SIN^{1,5}, VER⁵
0556. *Physeia setosa* Chaudoir, 1868
SLP⁷

20. Paussini

0557. *Eohomopterus simojovelensis* Kraemer, 2006
CHIS³; “Miocene amber, Simojovel area, Chiapas, México” (Kraemer 2006: 108)
- 0558*. *Homopterus honduriensis* Darlington, 1937
SLP¹; (type locality: “Lancetilla, Honduras”, Darlington 1937: 57)
- 0559*. *Homopterus steinbachi* Kolbe, 1920
TAB¹; (type locality: “Ost-Bolivien, Provinz Sara” [= Province Sara, Bolivia], Kolbe 1920: 153)

21. Brachinini

Note: Erwin's superscript ¹ for *Brachinus* species refers to Erwin, 1970 and states with a superscript ^H are from Erwin, 2011a.

- 0560.** *Brachinus aabaaba* Erwin, 1970
SLP^{1,2,3}
- 0561.** *Brachinus adustipennis* Erwin, 1969
AGS^{1,2,9}, NAY^{5,29}, SIN^{2,3,5,29}, SON^{1,2,3,29,54}, TAB^{1,29}, TAMPS^{1,2,29}, VER^{1,2,3,29}, YUC^{1,2,29}
- 0562.** *Brachinus aeger* Chaudoir, 1876
BCN^H, CHIS^{1,2,29}, GRO^{1,2,29}, JAL², MOR^H, NAY^{1,2,29}, OAX^{1,2,9}, SIN^{1,29}, SLP^{1,29}, SON^{1,29}, TAB^{1,29}, VER^{1,29}
- 0563.** *Brachinus alternans* Dejean, 1825
BCN^{26,H}, CHIH^H, SIN^H, SLP^H
- 0564.** *Brachinus arboreus* Chevrolat, 1834
JAL^{1,29}, MOR^{1,29}, NAY²⁹, SIN^{1,29}, TAMPS^H, VER^{1,4,29}
- 0565.** *Brachinus atramentarius* von Mannerheim, 1837
CAMP^H, VER^H
- 0566.** *Brachinus azureipennis* Chaudoir, 1876
AGS^{1,29}, CDMX^{1,5,29}, CHIH²⁹, DGO^{2,3,29}, GRO^{1,29}, JAL^{1,29}, MEX^{1,29}, MICH^{1,4,29}, PUE^{1,4,29}, SLP^{1,29}, SON⁵⁴, ZAC^{1,29}
- 0567.** *Brachinus chalchihuitlicue* Erwin, 1970
COL^H, GRO^{1,3,29}, NAY^{1,2,3,5,29}, SIN^{1,3,5,29}, VER³
- 0568.** *Brachinus chirriador* Erwin, 1970
CHIS^{1,2,3,29}, HGO^{3,24}, JAL^{1,29}, NAY^{1,29}, SLP^{1,3,29}, TAMPS^{1,2,3,29}, VER^{1,3,29}
- 0569.** *Brachinus cibolensis* Erwin, 1970
CHIH³, DGO^{1,2,5,29}, SON^{2,54}
- 0570.** *Brachinus cinctipennis* Chevrolat, 1835
AGS^{1,29}, CDMX^{1,29}, JAL^{1,2,3,5,29}, MEX^{1,2,4,29}, MICH³, PUE^{1,2,29}, SLP^{1,29}
- 0571.** *Brachinus consanguineus* Chaudoir, 1876
GRO^{1,2,29}, MEX²⁹, MICH^H, SIN^{1,29}, SON^{1,2,5,29,54}
- 0572.** *Brachinus costipennis* Motschulsky, 1859
BCN^{1,5,29}, DGO^{1,2,29}, GTO⁴, JAL^{5,29}, OAX^{1,2,5,29}, PUE^{1,2,3,29}, SIN^{1,5,29}, SLP^{1,2,5,29}, SON^{2,3,5,6,29,54}, VER^{1,2,3,4,29}, ZAC^{1,2,29}
- 0573.** *Brachinus elongatulus* Chaudoir, 1876
AGS^{1,2,3,29}, BCN^{1,3,5,6,29}, BCS⁵, CAMP⁴, CDMX^{1,29}, CHIH^{1,2,3,5,6,29}, CHIS^{4,7}, DGO^{1,2,3,4,7,29}, GTO^{1,2,3,29}, GRO^{1,2,3,4,7,29}, HGO^{1,2,29}, JAL^{1,2,3,4,5,6,29}, MEX^{1,2,4,5,29}, MICH^{1,2,3,5,29}, MOR^{1,2,3,4,5,6,29}, NAY^{1,2,3,4,5,6,29}, OAX^{1,2,3,4,7,29}, PUE^{1,2,4,5,29}, QRO^{1,2,3,29}, QROO^H, SIN^{1,2,3,5,29}, SLP^{1,3,29}, SON^{1,2,3,5,6,29,54}, TAMPS^{1,2,29}, TLAX²⁹, VER^{2,3,4}, YUC⁴, ZAC^{1,2,29}
- 0574.** *Brachinus explosus* Erwin, 1970
HGO², SLP^{1,3,29}, SON^{2,54}
- 0575.** *Brachinus favicollis* Erwin, 1965
BCN^{1,5,29}, SON^{2,54}
- 0576.** *Brachinus fumans* (Fabricius, 1781)
BCN^H, CHIH^{1,29}, COAH^{1,3,29}, COL^H, HGO³, NL^{1,2,5,29}, PUE³, SIN^{1,2,5,29}, SLP^{2,29}, SON^{1,2,3,5,6,29,54}, TAB^H, TAMPS^{1,2,29}
- 0577.** *Brachinus galactoderus* Erwin, 1970
GRO^{1,2,3,29}, NAY^{1,2,3,5,29}, OAX^{1,2,29}, SIN^{1,2,3,5,29}, SON^{1,3,5,29,54}
- 0578.** *Brachinus gebhardis* Erwin, 1965
BCN^{1,3}, BCS^{3,5,29}, SON^{2,3,6,54}

- 0579.** *Brachinus geniculatus* Dejean, 1831
BCN^H, CHIS^{1,2,3,29}, DGO^{1,29}, GRO^{2,4}, HGO^{1,29}, JAL³, MICH^{1,29}, MOR³, NAY², NL^{1,2,3,29}, OAX^{1,2,3,29}, PUE^{1,3,4,29}, QRO^{1,2,3,29}, QROO^H, SIN^H, SLP^{1,2,29}, SON^H, TAB², TAMPS^{1,2,3,29}, VER^{1,2,3,29}, YUC^{1,2,3,29}, ZAC^H
- 0580.** *Brachinus grandis* Brulle, 1837
SIN^{1,5,29}, SLP²⁹, TAB^{1,29}
- 0581.** *Brachinus hirsutus* Bates, 1884
CHIH^{18,29}, DGO^{1,2,29}, GTO², HDO^{1,2,29}, JAL^{1,2,29}, SLP^{1,2,29}, SON^{1,2,3,6,29,54}, ZAC^{1,2,29}
- 0582.** *Brachinus imperialensis* Erwin, 1965
DGO^{1,29}, SIN^{1,5,29}, SLP^{1,29}, SON^{1,5,29,54}
- 0583.** *Brachinus javalinopsis* Erwin, 1970
COAH²
- 0584.** *Brachinus kavanaughi* Erwin, 1969
NL^{1,2,29}, TAMPS^{1,29}
- 0585.** *Brachinus lateralis* Dejean, 1831
BCN^{1,5,29}, BCS⁵, CHIS^{1,29}, DGO^{1,29}, GRO^{1,2,29}, GTO^{1,2,3,29}, JAL^{1,2,3,6,29}, MICH^{1,2,4,29}, MOR^{1,2,3,29}, NAY^{1,3,5,6,29}, OAX^{1,2,29}, PUE^{1,2,3,29}, SIN^{1,3,5,29}, SON^{1,2,3,5,29,54}, VER^{1,29}
- 0586.** *Brachinus medius* T.W. Harris, 1828
TAMPS^{1,2,29}
- 0587.** *Brachinus melanarthrus* Chaudoir, 1876
CHIS², SLP^{1,29}, TAB^H, VER^{1,2,29}
- 0588.** *Brachinus mexicanus* Dejean, 1831
AGS^{1,2,29}, BCN^{1,3,5,6,29}, BCS^{5,29}, CDMX^{1,4,5,29}, CHIH^{1,2,3,6,29}, CHIS^{2,3,29}, COAH^{1,2,29}, COL^H, DGO^{1,2,29}, GRO^{1,2,29}, GTO^{1,2,4,29}, HGO^{1,2,29}, JAL^{1,2,3,5,6,29}, MEX^{2,3,5,29}, MICH^{1,2,3,4,29}, MOR^{1,5,29}, NAY^{2,3,5,6,29}, NL^{1,2,3,5,29}, OAX^{1,2,5,29}, PUE^{1,2,3,4,5,29}, QRO^{1,2,29}, SIN^{1,29}, SLP^{1,2,5,56}, SON^{1,2,3,5,6,29,54}, TAMPS², VER^{1,2,3,29}, ZAC^{1,2,29}
- 0589.** *Brachinus oaxacensis* Erwin, 1970; Figure 1H
CHIH³, GRO^{1,29}, OAX^{1,2,29}, SON²⁹
- 0590.** *Brachinus phaeocerus* Chaudoir, 1868
CHIH^{1,29,H}, (type locality: "l'Amérique septentrionale", Chaudoir 1868: 300); "Texas" (lectotype, type locality, Erwin 1970: 111)
- 0591.** *Brachinus quadripennis* Dejean, 1825
COAH³, DGO^H
- 0592.** *Brachinus sallei* Chaudoir, 1876
CAMP^{1,2,29}, CHIS^{1,29}, QROO (Cozumel Island)^{1,29}, SLP^{1,29}, TAB^{1,29}, VER^{1,29}
- 0593.** *Brachinus sonorous* Erwin, 1970
SIN^{1,29}, SON^{1,3,5,29,54}
- 0594.** *Brachinus sublaevis* Chaudoir, 1868
TAMPS³
- 0595.** *Pheropsophus aequinoctialis* (Linnaeus, 1763)
VER⁴, YUC^{1,4,17}
- 0596.** *Pheropsophus biplagiatus* Chaudoir, 1876
JAL⁴, OAX^{1,2,4,17}

22. Morionini

- 0597.** *Morion aridus* Allen, 1968
BCN⁵, BCS^{2,3,5}, CHIS², SIN^{2,5}, SON^{2,3,5,54}
- 0598.** *Morion cordatus* (Chaudoir, 1837)
CHIS^{2,3,5}, COL², GRO^{2,3,5}, HGO²⁴, JAL^{2,3,5}, NAY^{2,5} (Tres Marias Islands), OAX^{2,3}, SLP^{2,7}, SON^{2,54}, TAMPS⁵, VER^{2,3,5}, YUC²

0599. *Morion lafertei* Guérin-Méneville, 1844
CHIS^{3,7}, HGO²⁴, JAL^{3,5}, OAX¹⁶, SIN⁵, SON^{3,6,16,54},
TAMPS³, VER^{3,4,5,16}

0600. *Morion monilicornis* (Latreille, 1805)
BCS³, CHIS⁴, COL⁴, DGO¹⁶, MOR⁴, PUE⁴, SON^{3,6},
TAMPS⁴, VER^{4,16}

0601. *Morion simplex* Dejean, 1826
CAMP², CHIS², OAX², VER^{2,5}, YUC¹⁶

23. Abecetini

0602. *Oxycrepis anthracina* (Allen and Ball, 1980)
CHIS^{2,10}

0603. *Oxycrepis balli* (Allen, 1972)
CHIS^{2,9,10}, TAMPS³, VER^{2,3,9,10}

0604. *Oxycrepis binotata binotata* (Allen and Ball, 1980)
CHIS^{3,7,10}, TAB¹⁰

0605. *Oxycrepis binotata lacondonensis* (Allen and Ball, 1980)
CHIS¹⁰

0606. *Oxycrepis binotata tuxtensis* (Allen and Ball, 1980)
VER¹⁰

0607. *Oxycrepis celeris* (Dejean, 1828)
CAMP^{2,9}, CHIS^{2,9}, COAH^{2,3}, COL^{2,9}, SLP^{2,9}, TAB²,
TAMPS², VER^{2,9}

0608. *Oxycrepis cubana* (Tschitschérine, 1901)
JAL^{2,9}, NAY^{2,9}, OAX^{2,9}, SIN⁹

0609. *Oxycrepis elnae* (Allen, 1972)
CHIS^{2,3,9}, COL², GRO², MOR^{2,9}, OAX^{2,9}

0610. *Oxycrepis elongata* (Allen, 1972)
CHIS^{2,9}, OAX², VER^{2,3,9}

0611. *Oxycrepis fraus* (Allen, 1972)
CHIS^{2,9}, MOR^{2,3,9}, NAY^{2,9}, OAX^{2,9}, PUE²,

0612. *Oxycrepis infima* (Bates, 1882)
CHIS², COL^{2,9}, GRO², MICH^{2,3,9}, MOR^{2,9}, NL⁷,
QROO^{2,3}, SIN⁹, SLP², SON^{2,5,9,54}, TAB^{4,9}, TAMPS^{2,9},
VER¹⁶

0613. *Oxycrepis intercepta* (Chaudoir, 1873)
CHIS¹⁰, NAY^{3,10}, OAX^{3,10}, SLP⁷, TAMPS^{3,10}, VER^{4,10,16},
YUC¹⁶

0614. *Oxycrepis lepida* (Allen, 1972)
VER^{3,9}

0615. *Oxycrepis littleorum* (Allen and Ball, 1980)
CHIS^{2,7,10}, TAB², VER¹⁰

0616. *Oxycrepis omiltemi* (Allen and Ball, 1980)
GRO^{2,10}

0617. *Oxycrepis pactinullus* (Allen, 1972)
COL^{2,9}, MICH^{2,3,9}, MOR^{2,9}, SON^{2,9,54}

0618. *Oxycrepis rectangula* (LeConte, 1878)
CAMP^{2,3,9}, CHIS^{2,9}, JAL^{2,3}, NAY^{2,3}, TAB², VER^{2,3,9}

0619. *Oxycrepis rubricata* (Bates, 1891)
GRO¹⁹

0620. *Oxycrepis sculptilis* (Bates, 1884)
CHIS^{2,3,9}, COL^{2,3,9}, GTO¹⁸, JAL^{2,9}, MICH¹⁰, MOR^{2,3,4,9},
NAY², OAX^{9,18}, SIN⁹, SLP¹⁰, SON^{2,54}, TAMPS^{2,3,9},
VER^{2,3,18}

0621. *Oxycrepis stockwelli* (Allen and Ball, 1980)
CHIS¹⁰

0622. *Oxycrepis whiteheadi* (Allen, 1972)
CHIS^{3,9}, NAY¹⁰, OAX⁹, VER⁹

0623. *Oxycrepis yeariani* (Allen, 1972)
COL^{3,9}

24. Pterostichini

0624. *Abaris aequinoctialis* Chaudoir, 1852
CHIS^{2,3,58}, HGO^{3,24,58}, OAX^{2,58}, SLP^{2,58}, TAMPS^{2,58},
VER^{2,3,58}

0625. *Abaris aquilonaria* Will, 2002
CHIS^{7,58}, VER⁵⁸

0626. *Abaris bigenera* Bates, 1882
CHIS^{2,58}, GRO^{2,58}, JAL^{2,58}, MEX², MICH^{2,58}, OAX^{2,16,58},
SLP^{2,58}, SIN^{2,58}, VER²

0627. *Abaris splendidula* (LeConte), 1863
BCS^{2,3,58}, SIN², SON^{2,54,58}

Note: Erwin's superscript ¹ for *Allotriopus* species refers to Ball and Roughley 1982.

0628. *Allotriopus ashei* (Ball and Roughley, 1982)
OAX^{1,3}

0629. *Allotriopus brachypterus brachypterus* (Chaudoir, 1878)
OAX^{1,3}

0630[#]. *Allotriopus brachypterus greenwoodorum* (Ball and Roughley, 1982)

OAX¹; (type locality: "Llano Verde and vicinity, e. San Pedro Nolasco (n.e. Capulalpam), Oaxaca, México", Ball and Roughley 1982: 369)

0631. *Allotriopus hallbergi* (Ball and Roughley, 1982)
OAX^{1,3}

0632. *Allotriopus hemingi* (Ball and Roughley, 1982)
OAX^{1,2,3}

0633. *Allotriopus oscitans* (Tschitschérine, 1900)
OAX¹; (type locality: "México", Tschitschérine 1900: 465); type locality restricted to Oaxaca, Ball and Roughley 1982: 350.

0634. *Allotriopus serratipes* (Chaudoir, 1878)
OAX^{1,3}

0635[#]. *Allotriopus shpeleyi* (Ball and Roughley, 1982)
OAX¹; (type locality: "16.6 km. s. Totontepec, Oaxaca, México", Ball and Roughley 1982: 362)

0636. *Allotriopus taeniola* (Bates, 1882)
CHIS¹; (type locality: "Quiche Mountains, Guatemala", Bates 1882: 82)

0637. *Allotriopus triunfo* (Ball and Roughley, 1982)
CHIS^{1,3}

0638[#]. *Allotriopus whiteheadi* (Ball and Roughley, 1982)
OAX¹; (type locality: "35.7 mi. s. Valle Nacional, Oaxaca, México", Ball and Roughley 1982: 352)

0639. *Cratocerus indupalmensis* Grzymala and Will, 2014
MEX³

0640. *Cratocerus sulcatus* Chaudoir, 1852
SLP⁷, VER²

0641. *Cratocerus tanyae* Grzymala and Will, 2014
CHIS³

0642. *Cyclotrachelus substriatus* (LeConte, 1848)
CHIH^{2,3}, DGO², SON^{2,54}

Note: Erwin's superscript ¹ for *Cyrtolaus* species refers to Whitehead and Ball 1975.

0643[#]. *Cyrtolaus brevispina* Whitehead and Ball, 1975
CHIS¹; (type locality: "Yerba Buena Hosp., 1.5 mi. n.

- Pueblo Nuevo, Chiapas, México”, Whitehead and Ball 1975: 608)
0644. *Cyrtolaus furculifer* Bates, 1882
CHIS^{1,2,3}
0645. *Cyrtolaus grumifer* Whitehead and Ball, 1975
CHIS^{1,2,5}
- 0646#. *Cyrtolaus newtoni* Whitehead and Ball, 1975
OAX¹; (type locality: “22.4 mi. s. Valle Nacional, Oaxaca, México”, Whitehead and Ball 1975: 609)
0647. *Cyrtolaus oaxacana* Ball, 1991
OAX^{1,2,3}
- 0648#. *Cyrtolaus orizabae* (Csiki, 1930)
VER¹ (*Cyrtolaus* (*Ithytolus*) *orizabae* Csiki 1930: 585 is a replacement name for *Ithytolus anomalus* Bates 1884: 278 (type locality: “Orizaba, Veracruz, México”), [nec. *Pterostichus* (*Ophryogaster*) *anomalus* Chaudoir, 1878])
0649. *Cyrtolaus ricardo* Whitehead and Ball, 1975
CHIS^{1,2,3,5}
0650. *Cyrtolaus subiridescens* Whitehead and Ball, 1975
CHIS^{1,2}
0651. *Cyrtolaus whiteheadi* Ball, 1991
OAX^{1,2}
0652. *Euchroa atoyac* Frania and Ball, 2007
GRO^{2,3,32}
0653. *Euchroa carbonera* Frania and Ball, 2007
OAX^{2,3,32}
0654. *Euchroa chrysophana* Bates, 1891
GRO^{2,3,19,32}
- 0655#. *Euchroa citlaltepetl* Frania and Ball, 2007
VER³²; (type locality: “E. Citlaltepetl, Veracruz, México”, Frania and Ball 2007: 55)
0656. *Euchroa cuiyachapa* Frania and Ball, 2007
VER^{2,32}
0657. *Euchroa dimidiata* Chaudoir, 1874
GRO^{2,32}, MOR^{2,3,32}, OAX^{2,3,32}, VER²
0658. *Euchroa filodecaballo* Frania and Ball, 2007
GRO^{2,3,32}
0659. *Euchroa flohri* Bates, 1882
PUE^{16,32}, VER^{2,3,32} **note:** Frania and Ball (2007: 52) questioned the validity of the Puebla locality
0660. *Euchroa harrisoni* Frania and Ball, 2007
HGO^{2,3,24}, QRO^{2,3}, SLP², TAMPS^{2,3,32}
0661. *Euchroa huautla* Frania and Ball, 2007
HGO², OAX^{2,32}, QRO², SLP², TAMPS²
0662. *Euchroa ixtapa* Frania and Ball, 2007
GRO^{2,32}
0663. *Euchroa jalisco* Frania and Ball, 2007
JAL^{2,3,32}
0664. *Euchroa juchatengo* Frania and Ball, 2007
OAX^{2,32}
0665. *Euchroa lasvigas* Frania and Ball, 2007
VER^{2,32,52}
0666. *Euchroa miahuatlan* Frania and Ball, 2007
OAX^{2,3,32}
0667. *Euchroa nitidipennis* Putzeys, 1846
OAX³², VER^{2,3,4,7,32}
0668. *Euchroa nizavaguiuti* Frania and Ball, 2007
OAX^{2,3,32}
0669. *Euchroa perote* Frania and Ball, 2007
HGO²⁴, PUE^{2,3,7}, TLAX², VER^{2,32}
0670. *Euchroa puertogallo* Frania and Ball, 2007
GRO^{2,32}
0671. *Euchroa sallei* Chaudoir, 1874
VER^{2,3,4,32}
0672. *Euchroa santacatarina* Frania and Ball, 2007
OAX^{2,3,32}
0673. *Euchroa soladevega* Frania and Ball, 2007
OAX^{2,32}
0674. *Euchroa suchixtepec* Frania and Ball, 2007
OAX^{2,32}
0675. *Euchroa tenancingo* Frania and Ball, 2007
MEX^{2,3,32}
0676. *Euchroa teotitlan* Frania and Ball, 2007
OAX^{2,32}, PUE^{2,32}
0677. *Euchroa yucuyacua* Frania and Ball, 2007
OAX^{2,3,32}
0678. *Euchroa zempoaltepetl* Frania and Ball, 2007
OAX^{2,3,32}
- 0679#. *Euchroa zongolica* Frania and Ball, 2007
VER³²; (type locality: “Five km. s.w. of San Andrés de Tenejapa, State of Veracruz, México. The designation of the type locality is based upon information that Sótano de Milpa is located 5 km southwest of San Andrés de Tenejapa in the Municipio de Tenejapa (Redell 1981”, Frania and Ball 2007: 48).
0680. *Hybothecus anomalus* (Chaudoir, 1878)
VER²
0681. *Hybothecus flohri* (Bates, 1882)
BCS^{2,26}, CHIS^{2,3}, COL², GRO², MOR², NAY², NL², SON^{2,54}, VER²
0682. *Percolaus championi* Bates, 1882
CHIS^{2,5}
0683. *Percolaus guillermo* (Ball and Roughley, 1982)
CHIS²
- 0684*. *Piesmus submarginatus* (Say, 1823)
No type locality by Say 1823: 45; (type locality: “Hope, Hempstead County, Arkansas”, designated by Lindroth and Freitag, 1969: 340); “North America, Southern States and México” (Bates 1882: 83)
- 0685#. *Poecilus aztecus* (Tschitschérine, 1897)
Type locality: “México”, Tschitschérine 1897: 349
0686. *Poecilus chalcites* (Say, 1823)
COAH², COL², GTO², MICH², MOR², QRO², SLP², TAMPS^{2,3}, VER²
0687. *Poecilus cyanicolor* Chaudoir, 1876
CDMX⁴, SLP⁴, VER⁴
0688. *Poecilus diplophryus* Chaudoir, 1876
BCS⁴, CHIH³, SIN², SON^{2,54}
0689. *Poecilus mexicanus* Chaudoir, 1876
GTO⁴, MICH^{3,4}
0690. *Poecilus scitulus* LeConte, 1848
CHIH², DGO², GTO², SIN⁷, SLP², SON^{2,54}, TAMPS²
0691. *Pseudabarys mexicanus* Chaudoir, 1873
CHIS⁵⁷, VER⁵⁷
0692. *Pseudabarys substriata* Chaudoir, 1873
GRO⁴, JAL⁵⁷, MICH⁵⁷, MOR⁴, NAY⁵⁷, OAX³, PUE⁵⁷, SIN⁵⁷, SON², VER⁵⁷

- Note:** Kipling Will, Essig Museum, Berkeley, California, is currently revising the Mexican species of *Pseudabarys*, and is describing five new species, three of which are from Jalisco and the remaining two from Nayarit.
0693. *Pterostichus brevicornis* (Kirby, 1837)
VER⁴; **note:** most likely mis-identified or mis-labelled; type locality: “Lat. 65° [= apparently region of Great Bear Lake, Northwest Territories]”, Kirby 1837: 31; type locality restricted to “Good Hope, Northwest Territories” by Ball (1966: 105); known only from Canada and northeastern USA (Ball and Currie 1997: 459, Fig. 4)
- 0694*. *Pterostichus hornii* LeConte, 1873
Type locality: “S.E. Sierras of California”, LeConte 1873: 313, restricted to Fort Tejon, Kern County, California (Van Dyke 1927: 196); “record from Baja California (Horn 1894: 308) needs confirmation” (Bousquet 2012: 840)
0695. *Pterostichus illustris* LeConte, 1851
BCN^{2,3,26}
0696. *Pterostichus jacobinus* Casey, 1913
BCN^{2,26}
- 0697*. *Pterostichus protractus* LeConte, 1860
Type locality: “Jasper House, [Alberta, Canada]”, LeConte 1860: 319; “Baja Calif” (Blackwelder 1944: 37); “México” (Erwin et al. 1977: 4.37)
0698. *Pterostichus tropicalis* Bates, 1882
CHIS^{2,3,7}, DGO², OAX^{2,16}, VER¹⁶
- 0699#. *Stomis granulatus* Say, 1834
Type locality: “México”, Say 1834: 427
- ### 25. Zabryni
0700. *Amara aenea* (DeGeer, 1774)
BCN³
0701. *Amara apachensis* Casey, 1884
CHIH¹, SON^{1,5}
0702. *Amara aurata* Dejean, 1828
BCN^{5,26}
0703. *Amara basillaris* (Say, 1823)
COL⁷
0704. *Amara californica* Dejean, 1828
AGS², BCN^{5,26}, CDMX^{4,5}, CHIH^{2,5}, CHIS^{2,5}, COAH², DGO^{2,5}, HGO^{2,4,5,24}, JAL², MEX^{2,3,4,5}, MICH², NL², OAX^{1,2}, PUE^{2,4}, QRO², SLP², SON^{2,3,5,54}, TAB⁵, TLAX², VER², ZAC²
0705. *Amara chihuahuae* (Casey, 1918); Figure 1I
CHIH⁵⁹, NL²
0706. *Amara deparca* (Say, 1830)
BCN^{5,26}, CHIH^{2,5}, DGO^{2,3,4,5}, MEX^{2,4,5}, MICH², MOR⁵, PUE², SON^{2,54}, VER^{2,3}
0707. *Amara dolosa* Say, 1834
CDMX², COAH², DGO^{2,5}, GTO², HGO^{2,5,24}, MEX^{2,4}, NL^{2,3,7}, OAX^{2,5}, PUE^{2,4,5}, QRO², SLP², TAB⁵, TLAX², VER^{2,3}, ZAC²
0708. *Amara durangensis* Van Dyke, 1943; Figure 1J
DGO^{2,3,5}
- 0709#. *Amara habilis* Casey, 1918
MEX; (type locality: “México (Salazar, Mex. [state])”, Casey 1918b: 255)
0710. *Amara impuncticollis* (Say, 1823)
CHIH², NL^{1,2}, TLAX²
0711. *Amara insignis* Dejean, 1831
BCN^{5,26}
- 0712*. *Amara jacobina* LeConte, 1855
Type locality: “San Diego, California”, LeConte 1855: 346; “México” (Erwin et al. 1977: 4.40; Bousquet, 2012: 894)
0713. *Amara littoralis* von Mannerheim, 1843
DGO³
0714. *Amara lugens* (Zimmermann, 1832)
CDMX², CHIH², DGO^{1,2,3,5}, HGO³, MEX^{2,4}, MOR⁵, OAX^{1,2}, QRO², TLAX², VER^{1,2,3,5}
0715. *Amara moerens* (Zimmermann, 1832)
AGS², CDMX^{2,5}, HGO², JAL², MEX^{1,2,5}, MICH², MOR⁵, NAY⁵, NL², OAX², PUE², QRO², TLAX², VER^{2,3}
0716. *Amara obesa* (Say, 1823)
VER³
- 0717#. *Amara parata* Casey, 1918
MEX; (type locality: “México (Salazar, Mex. [state])”, Casey 1918b: 256)
0718. *Amara pseudobrunnea* Lindroth, 1968
VER³; **note:** most likely mis-identified or mis-labelled; type locality: “Swift River, Yukon Territory”, Lindroth 1968: 706; known only from Canada and Idaho and Montana in USA (Bousquet 2012: 918)
0719. *Amara pterostichina* Hayward, 1908
CHIH^{2,3,5}, CDMX^{2,5}, DGO^{2,5}, GTO², MEX², NL², OAX², PUE^{2,4}, SLP², TLAX², ZAC²
0720. *Amara rectangula ciudadensis* Bates, 1891
CHIH^{1,2}, COAH², DGO^{1,2,5}, NL^{1,2}, SON^{1,2,54}
0721. *Amara sera* Say, 1830; Figure 1K
AGS², CDMX^{4,5}, CHIH², CHIS⁵, COAH², DGO², HGO^{2,4}, JAL², MEX², MICH², NL², OAX^{1,2}, QRO², SON^{2,54}, TLAX^{2,7}, VER^{2,3}, ZAC²
0722. *Amara sublustris* Tschitscherine, 1898
CDMX⁴, MEX⁴, MICH⁴; **note:** most likely mis-identified or mis-labelled; type locality: “Turkestan russe: monts d’Alexandre”, Tschitscherine 1898: 221.
0723. *Amara tenebrionella* (Bates, 1882)
CDMX^{2,7}, CHIH^{2,5}, DGO^{1,2,3,4,5}, GRO¹, HGO⁵, MEX², TLAX², VER^{2,3,16}
- ### 26. Oodini
0724. *Anatrichis minuta* (Dejean, 1831)
COAH³
0725. *Anatrichis oblonga* G. Horn, 1891
CAMP^{1,2}, CHIS^{1,2}, QRO^{1,2}, TAB^{1,2}, TAMPS^{1,2}, VER^{1,2}
0726. *Dercylus mexicanus* (Bates, 1891)
CHIS^{2,19}, NAY²
- 0727*. *Oodes parallela* (Say, 1830)
Type locality: “Louisiana”, Say 1830: 8; (type locality restricted by Bousquet 1996: 468 to “New Orleans, Orleans Parish”); (Summers, 1874 noted seeing one specimen “from México, near the Rio Grande”) Bousquet 2012: 955

0728. *Oodinus alutaceus* (Bates, 1882)
CAMP^{1,2}, CHIS^{1,2}, COL^{1,2}, JAL^{1,2}, NAY^{1,2}, OAX^{1,2},
SIN⁵, SLP^{1,2}, SON^{1,2,5,4}, TAB^{1,2}, TAMPS^{1,2,3}, VER^{1,6}
0729. *Oodinus edentulus* Bousquet, 1996
CAMP^{2,20}, CHIS^{2,20}, TAB^{2,20}
- 0730*. *Oodinus piceus* Motschulsky, 1864
Type locality: “Panama”, Motschulsky 1864: 335
0731. *Oodinus similis* Bousquet, 1996
CAMP^{1,2}, CHIS^{1,2}, JAL^{1,2}, NAY^{1,2}, QRO¹, TAB^{1,2}, VER^{1,2}
0732. *Polychaetus dejeani* Chaudoir, 1882
CHIS^{1,2}, GRO^{1,4}, MICH¹, QRO^{1,2}, QROO⁴, VER^{1,2}
0733. *Stenocrepis cuprea* (Chaudoir, 1843)
TAMPS²
0734. *Stenocrepis duodecimstriata* (Chevrolat, 1835)
CAMP^{2,20}, CHIS^{2,20}, COAH^{2,3}, NAY², QRO⁵, TAB^{2,20},
VER^{2,5,20}, YUC^{2,20}
0735. *Stenocrepis elegans* (LeConte, 1851)
CHIH¹, COAH^{5,20}, MOR⁴, NAY^{1,2}, SIN⁵, SON^{1,3,5,6,54}
0736. *Stenocrepis gratiosa* (Bates, 1882)
CHIS^{1,2}, OAX^{1,2}, TAB¹, VER^{4,16}
0737. *Stenocrepis insulana* (Jacquelin du Val, 1856)
CAMP^{1,2}, CHIS^{1,2}, NAY^{1,5}, SIN^{1,5}, SLP^{1,2}, TAB^{1,2},
VER^{1,2}, YUC¹
0738. *Stenocrepis mexicana* (Chevrolat, 1835)
CAMP^{2,20}, CHIS^{2,5,20}, COL^{2,20}, GRO^{2,20}, JAL³, PUE⁴,
SLP²⁰, TAB^{2,20}, TAMPS^{2,20}, VER^{2,4,5}, YUC²⁰
0739. *Stenocrepis olivacea* (Bates, 1878)
CAMP^{1,2}, CHIS^{1,2}, GRO^{1,2}, JAL^{1,2}, NAY^{1,2}, OAX^{1,2},
SIN^{1,2,5}, TAB^{1,2}, VER^{1,2,4,5}
0740. *Stenocrepis tibialis* (Chevrolat, 1834); Figure 1L
CAMP^{1,2}, CHIS^{1,2}, COAH², GRO^{1,2}, NAY¹, NL^{1,7},
OAX^{1,2}, SIN^{1,2}, SLP¹, SON¹, TAB^{1,2}, TAMPS^{1,2}, VER^{1,2,4}

27. Panagaeni

0741. *Coptia armata* Laporte de Castelnau, 1832
VER⁴; type locality: “Cayenne [French Guiana]”, La-
porte de Castelnau 1832: 391.
0742. *Micrixys distinta* (Haldeman, 1852)
AGS³
0743. *Micrixys mexicana* Van Dyke, 1927
DGO⁴, SIN¹
0744. *Panagaenus cruciger* Say, 1823
OAX³
0745. *Panagaenus fasciatus* Say, 1823
CDMX⁴; **note:** most likely mis-identified or mis-la-
belled; type locality: none stated, “North America [in-
ferred from title]”, Say 1823: 70.
0746. *Panagaenus quadrisignatus* Chevrolat, 1835
JAL^{1,4}, VER^{1,3}
0747. *Panagaenus sallei* Chaudoir, 1862
BCN^{1,26}, CDMX^{1,4}, CHIH¹, GRO¹, HGO²⁴, MEX⁴,
PUE^{1,4}, SON^{2,54}, VER^{1,4}

28. Peleciini

Note: Erwin’s superscript ¹ for *Eripus* species refers to Straneo and Ball 1989.

- 0748#. *Eripus breedlovei* Straneo and Ball, 1989
CHIS¹; (type locality: “México, Chiapas, Municipio
Comitan, 8-12 km. N. Mex. Hwy 190 at Laguna Cha-

- mula on logging rd.”, Straneo and Ball 1989: 99)
0749. *Eripus globipennis globipennis* (Chaudoir, 1866)
VER^{1,2}
0750. *Eripus globipennis rotundicollis* Straneo and Ball,
1989
GRO^{1,2,3,5}
0751. *Eripus globipennis whiteheadi* Straneo and Ball,
1989
GRO^{1,3}, JAL^{1,2}, MOR^{1,2}, PUE^{1,2}
- 0752#. *Eripus microphthalmus* (Chaudoir, 1866)
VER¹; (type locality: “México”, Chaudoir 1866a: 108)
0753. *Eripus nitidus* (Chaudoir, 1861)
HGO^{1,2,24}, OAX^{1,2}, PUE^{1,2}, QRO², TAMPS¹, TLAX²,
VER^{1,2,3,5}
0754. *Eripus oaxacanus* Straneo and Ball, 1989
OAX^{1,2,5}
0755. *Eripus scydmaenoides* Dejean, 1829
CHIS^{1,2,3,7}, GRO^{1,2}, JAL^{1,2}, MEX^{1,2}, OAX^{1,2}, PUE¹,
QRO^{1,2}, SLP^{1,2}, VER^{1,4}
0756. *Eripus subcaecus* (Chaudoir, 1866)
OAX^{1,2}
0757. *Eripus suturalis* (Chaudoir, 1861)
CHIS^{1,2,3,7}, GRO^{1,2}, OAX^{1,2}, PUE²⁷, QRO^{1,2},
TAMPS^{1,2,3,7}, TLAX²⁷, VER^{1,2,3}

29. Chlaeniini

0758. *Chlaenius aberrans* Bates, 1882
CHIS⁴, COL⁵, HGO⁵, JAL², MEX², MICH^{2,3}, MOR²⁸
- 0759#. *Chlaenius ampliatus* Bates, 1891
MOR⁵⁹; (type locality: “Cuernavaca, [Morelos, Méxi-
co]”, Bates, 1891: 237)
0760. *Chlaenius aurolimbatus* (LaFerté-Sénectère 1851)
CAMP¹⁶, CHIS², GTO¹⁶, JAL³, MOR², OAX², PUE^{4,16},
VER⁴, YUC¹⁶
0761. *Chlaenius azureus* Chaudoir, 1876
HGO²⁴, NL², OAX², QRO^{2,3}, SLP^{2,3,5}, TAMPS²,
VER^{2,3,4,16,28}
0762. *Chlaenius beatus* Bates, 1891
MOR¹⁹
0763. *Chlaenius brevilabris* LeConte, 1847
HGO²⁴
0764. *Chlaenius caeruleicollis* Chaudoir, 1876
AGS²⁸, CDMX²⁸, CHIH^{2,3,5}, DGO^{2,3,5}, GRO², GTO^{2,4},
HGO^{25,28}, JAL^{2,5}, MEX¹⁶, MICH^{2,4}, MOR²⁸, NAY⁵,
OAX^{2,16}, PUE^{2,16}, SIN^{5,28}, SLP³, SON^{2,54}, VER^{2,16}
0765. *Chlaenius chaudiiri* G. Horn, 1876
CHIH^{3,5,6,28}, COAH²⁸, DGO²⁸, SLP²⁸, SON^{12,3,5,6,54},
TAMPS³
0766. *Chlaenius chlorochrous* Chaudoir, 1876
CHIS^{3,28}, OAX², TAB², VER^{16,28}
0767. *Chlaenius circumcinctus* Say, 1830
VER^{4,16}
0768. *Chlaenius cordifer* Bates, 1891
CHIS²⁸, GRO², JAL^{2,5,7}, NAY^{2,5}, OAX^{2,7,19}, PUE²⁸,
SIN^{2,5}, SON²⁸, ZAC²
0769. *Chlaenius cumatilis* LeConte, 1851
BCN^{3,5,28}, BCS^{3,5,28}, CHIH³, SON^{3,6,28,54}
0770. *Chlaenius cursor* Chevrolat, 1835
AGS²⁸, BCS^{26,28}, CHIH²⁸, COAH⁵, COL², DGO²,

- GRO²⁸, GTO^{2,16}, JAL^{2,3}, MICH^{2,4}, MOR^{2,4,5}, NAY²⁸, OAX^{2,16}, PUE^{4,28}, QRO², SIN², SLP⁵, SON^{2,54}, VER^{2,4,16}
0771. *Chlaenius eurybates* Bates, 1891
GRO^{19,28}
0772. *Chlaenius forreri* Bates, 188; Figure 1M
= *Chlaenius pimalicus* Casey, 1914 **NEW SYNON-
OMY**; comparison of external morphology as well as
male genitalia confirms that these two species are con-
specific.
CHIH^{2,3,6}, DGO¹⁹, NAY⁵, OAX⁵, SIN^{2,3,5,6,54}, SON^{2,3,5,54}
- 0773*. *Chlaenius glaucus* LeConte, 1856
Type locality: "Colorado river, near the junction of the
Gila" [Arizona], LeConte, 1856: 28; "México" (Black-
welder 1944: 44; Erwin et al. 1977: 4.55)
0774. *Chlaenius guatemalensis* Bates, 1882
MOR²⁸, SLP²⁸, TAB²⁸, TAMPS²⁸, VER²⁸
0775. *Chlaenius herbaceus* Chevrolat, 1835
CAMP²⁸, GTO¹⁶, SON^{5,54}, VER²⁸
- 0776#. *Chlaenius koltzei* Grundmann, 1956
VER¹; (type locality: "Yalappa [= Jalapa, Veracruz],
México", Grundmann 1956: 75)
0777. *Chlaenius laticollis* Say, 1823
COAH³
0778. *Chlaenius leucoscelis leucoscelis* Chevrolat, 1834;
Figure 1N
AGS², BCS⁵, CHIH^{2,5,6}, CHIS², COAH^{2,5}, DGO²,
GRO², GTO^{2,16}, HGO², JAL^{2,5}, MOR², NAY², NL^{2,5},
OAX^{2,16}, PUE^{2,4,5}, QRO²⁸, SIN², SLP², SON^{2,6,54}, TAB²,
TAMPS², VER^{2,5,16}, ZAC^{2,28}
0779. *Chlaenius leucoscelis monachus* LeConte, 1851
SON³
0780. *Chlaenius leucoscelis sanantonialis* Casey, 1914
DGO³
0781. *Chlaenius nebraskensis* LeConte, 1856
SON^{5,54}
0782. *Chlaenius obscuripennis* Chevrolat, 1835
DGO⁴, OAX¹⁶, VER^{4,16}
0783. *Chlaenius obsoletus* LeConte, 1851
BCN^{3,5}, BCS^{5,26}, COAH⁵, DGO², OAX^{2,16}, SON^{2,5,54},
ZAC²
0784. *Chlaenius orbus* G. Horn, 1871
CHIH^{5,28}, COAH^{2,3,5,6}, DGO^{2,5}, NL^{2,5}, OAX^{2,16}, QRO²,
SIN²⁸, SLP², SON^{2,3,6,54}, TAMPS^{2,3}, VER³
0785. *Chlaenius planipennis* Chaudoir, 1876
OAX²
0786. *Chlaenius planulatus* Bates, 1884
DGO^{5,28}, SIN^{5,28}
0787. *Chlaenius porphyrius* Bates, 1891
GRO^{2,5,19}, MOR⁴
0788. *Chlaenius purpureus* Chaudoir, 1876
BCS^{3,5,28}, CHIH², CHIS², GRO^{3,28}, JAL⁵, NL³, OAX⁵,
SIN²⁸, SLP^{3,7,28}, SON^{2,3,6,54}, VER^{2,4,5,16}, ZAC^{5,28}
0789. *Chlaenius regularis* LeConte, 1851
SON²⁸
0790. *Chlaenius rodriguezi* Chaudoir, 1876
CHIS^{5,16}
0791. *Chlaenius ruficauda* Chaudoir, 1856
CHIH², DGO²⁸, MOR⁴, NAY^{2,5}, OAX¹⁶, PUE^{4,16},
SIN^{5,28}, SON^{2,3,6,54}
0792. *Chlaenius sallei* Chaudoir, 1876
MICH^{2,28}, VER^{16,28}
0793. *Chlaenius scabricollis* Chevrolat, 1833
VER¹⁶
0794. *Chlaenius sericeus* Forster, 1771
AGS³, CHIH³, COAH^{3,6}, SON³
0795. *Chlaenius soccatus* Say, 1830
CDMX^{4,16}, CHIH³, GTO¹⁶, MEX⁴, OAX¹⁶, PUE¹⁶,
VER¹⁶
0796. *Chlaenius soginoides* Chaudoir, 1876
CDMX⁴, CHIH^{2,3}, DGO², GTO¹⁶, MICH⁴
0797. *Chlaenius sparsus* LeConte, 1863
AGS^{2,5}, BCS⁵, CHIH², COL², DGO^{2,7}, GRO^{2,7}, GTO²,
JAL^{2,5}, MICH^{2,5}, MOR^{2,5}, NAY^{2,5}, OAX^{2,3,5}, PUE^{2,5},
SLP³, SIN⁵, SON^{2,5,54}, TAMPS²
0798. *Chlaenius suppletor* Bates, 1891
VER¹⁹
0799. *Chlaenius texanus* G. Horn, 1876
COAH²
0800. *Chlaenius tomentosus* (Say, 1823)
CHIH^{2,3}, PUE³, SON^{2,3,54}
0801. *Chlaenius tricolor* Dejean, 1826
BCS^{5,26}, CDMX⁴, CHIH^{2,3}, CHIS^{2,3}, DGO², GTO¹⁶,
HGO^{2,4,24}, JAL^{2,3,5}, MEX^{2,3}, MOR^{4,5}, MICH^{2,4}, NAY⁵,
NL², OAX^{4,5,16}, PUE^{2,4,5}, QRO², SIN⁵, SLP³, SON^{2,54},
VER^{2,5,16}
0802. *Chlaenius validus* (Chevrolat, 1835)
VER^{3,4,16}
0803. *Chlaenius variabilipes* Eschscholtz, 1833
BCN⁵, BCS^{5,26}
0804. *Chlaenius viridicollis* Reiche, 1843
OAX¹⁶

30. Licinini

0805. *Badister elegans* LeConte, 1880
COAH², COL¹, TAMPS^{1,2,3}
0806. *Badister flavipes mexicanus* Van Dyke, 1945
CHIS^{1,2}, MOR^{1,2}, NL^{1,2}, OAX^{1,2,5}, PUE^{1,2}, SLP^{1,2},
TAB^{1,2}, TAMPS^{1,2}
0807. *Badister reflexus* LeConte, 1880
MOR⁴, QROO^{1,2}
0808. *Badister vandykei* Ball, 1959
BCN¹, BCS^{3,5,26}, SIN^{1,5}
0809. *Dicaelus chermocki* Ball, 1959
CHIH³
0810. *Dicaelus costatus* LeConte, 1853
COAH², NL^{3,4,5}, TAMPS^{2,4,7}
0811. *Dicaelus franclemonti* Ball, 1992
COAH², COL¹, NL¹; (type locality: "México, state of
Nuevo León, the mountain Cerro Potosí, northwest of
Galeana", Ball, 1992: 358)
0812. *Dicaelus laevipennis laevipennis* LeConte, 1848
CDMX⁴, CHIH^{1,2}, DGO³, MEX⁴, NL⁴, SON³, TAMPS⁴
0813. *Dicaelus laevipennis abbreviatus* Bates, 1891
DGO^{1,2,5,19}, SIN^{1,2}
0814. *Dicaelus laevipennis flohri* Bates, 1878
CDMX⁵⁹, DGO^{1,2}, MEX^{1,2}, ZAC^{1,2}
0815. *Dicaelus suffusus* (Casey, 1913); Figure 1O
CHIH^{1,2}, SON^{1,2,6,54}

31. Harpalini

- 0816.** *Acupalpus lamprotus* (Bates, 1891)
DGO¹⁹
- 0817[#].** *Amblygnathus balli* Shpeley and Van Devender, 2022
SON⁵⁴; (type locality: “México, Sonora, Municipio de Fronteras, Rancho Capulín, 23.5 km (by air) WSW Fronteras, Sierra Buenos Aires, 30.76167°N 109.82361°W”, Shpeley and Van Devender 2022: 87)
- 0818.** *Amblygnathus evansi* Ball and Maddison, 1987
CAMP¹, CHIS^{1,2}, COL², DGO¹, GRO², JAL^{1,2,5}, NAY^{1,2,5}, OAX^{1,2,3}, SIN^{1,2,3,5,8}, SON¹
- 0819.** *Amblygnathus interior* Ball and Maddison, 1987
DGO^{2,12}, JAL^{2,12}, MICH¹², SON^{2,12,54}
- 0820.** *Amblygnathus mexicanus* Bates, 1882
CHIS², NL², TAB¹², VER^{2,3,5,12,16}, YUC¹²
- 0821.** *Amblygnathus subtinctor* (LeConte, 1866)
CAMP³, SLP², TAMPS^{2,3,12}, VER^{2,12}
- 0822.** *Amblygnathus tikal* Ball and Maddison, 1987
QRO¹²
- 0823.** *Amblygnathus whiteheadi* Ball and Maddison, 1987
CHIS¹², OAX^{2,12}, VER²
- 0824.** *Amblygnathus woodruffi* Ball and Maddison, 1987
CHIS^{1,2}, OAX¹, SLP^{1,2}, TAB¹, TAMPS^{1,2,3}, VER^{1,2,5}
- 0825.** *Amphasia interstitialis* (Say, 1823)
SON⁵⁴; (type locality: “Missouri”, Say 1823:57)
- 0826.** *Anisocnemus amblygonus* Shpeley and Ball, 1978
CAMP¹, CHIS^{1,2,3,5}, COL², GRO^{1,2,7}, JAL^{1,2,5,6,7}, MICH², OAX^{1,2,3,5,8}, SIN^{1,4,5}
- 0827.** *Anisodactylus anthracinus* (Dejean, 1829); Figure 1P
CDMX^{4,5}, CHIH², DGO^{2,3,5}, GTO², HGO², JAL², MEX², MICH², OAX², PUE⁴, QRO², SLP², SON^{2,3,6,54}, VER⁴, ZAC⁵
- 0828.** *Anisodactylus californicus* Dejean, 1829
BCN^{1,5,26}, BCS²⁶, JAL^{1,5}
- 0829.** *Anisodactylus consobrinus* LeConte, 1851
BCN^{1,26}, BCS⁵
- 0830.** *Anisodactylus darlingtoni* Noonan, 1973
CHIH², COAH^{3,6}, DGO³, GTO², JAL², MEX^{2,3}, MICH^{2,3}, OAX², PUE², ZAC²
- 0831.** *Anisodactylus harrisii* LeConte, 1863
CHIH²
- 0832.** *Anisodactylus opaculus* (LeConte, 1863)
BCS⁵
- 0833^{*}.** *Anisodactylus porosus* (Motschulsky, 1845)
BCN¹; (type locality: “California”, Motschulsky 1845: 344)
- 0834.** *Anisodactylus rotundangulus* Bates, 1878
CDMX², CHIH², DGO², PUE⁴
- 0835.** *Anisodactylus texanus* Schaeffer, 1919
CHIH², NL²
- 0836.** *Athrostictus punctatulus* (Putzeys, 1878)
OAX³, VER^{2,3}, YUC⁵⁷
- 0837.** *Athrostictus sericatus* Bates, 1878
CHIS², GRO², OAX^{2,4}, SON⁵⁴, TAB², VER^{2,3,4}
- 0838.** *Aztecarpalus hebescens* (Bates, 1878)
OAX²
- 0839.** *Aztecarpalus hemingi* Ball, 1976
OAX²
- 0840.** *Aztecarpalus lectocolus* Ball, 1970
OAX^{2,3}
- 0841.** *Aztecarpalus liolus* (Bates, 1882)
CDMX⁴, MEX⁴, OAX²
- 0842.** *Aztecarpalus marmoreus* Ball, 1970
HGO^{2,3}
- 0843.** *Aztecarpalus platyderus* (Bates, 1882)
PUE², VER^{2,3,16}
- 0844.** *Aztecarpalus schaefferi* Ball, 1970
NL^{2,7}, TAMPS²
- 0845.** *Aztecarpalus trochotrachis* Ball, 1970; Figure 1Q
OAX^{2,3}
- 0846[#].** *Aztecarpalus whiteheadi* Ball, 1976
OAX²; (type locality: “México, Oaxaca, 16.9 mi. s. Valle Nacional”, Ball 1976: 67)
- 0847.** *Barysomus hoepfneri* Dejean, 1829
COL², GRO², GTO³, JAL², MOR⁵
- 0848.** *Barysomus metallicus* Reiche, 1843
CHIS², GRO², MOR⁴, OAX^{2,3}, SIN^{2,3,5}, TAMPS², VER^{2,3,4}
- 0849.** *Bradycellus angulifer* Bates, 1882
GTO⁵⁹, OAX³, VER⁴
- 0850.** *Bradycellus circumdatus* (Bates, 1878)
CHIS², JAL², NAY², OAX², TAB², VER⁷
- 0851^{*}.** *Bradycellus congener* (LeConte, 1848)
Type locality: “Rocky Mountains”, LeConte 1848: 307
- 0852.** *Bradycellus flohri* (Bates, 1878)
CDMX⁵⁹, CHIH², DGO^{2,4}, GTO², NL², TLAX², VER^{2,3}
- 0853.** *Bradycellus intermedius* Fall, 1905
BCN⁵
- 0854.** *Bradycellus limbicollis* Bates, 1882
CHIS^{2,3,7}, GRO², NAY², OAX², SLP², TAB², VER²
- 0855^{*}.** *Bradycellus nebulosus* LeConte, 1853
BCN¹; “Baja California, United States” (Blackwelder 1944: 51); “Georgia” (Erwin et al. 1977: 4.44); (*Bradycellus nebulosus* LeConte 1853: 385, is a replacement name for *B. suturalis* LeConte 1848: 311 (type locality: “Georgia”)), [nec. Stephens 1828; nec. Dejean 1929]
- 0856.** *Bradycellus nigrellus* Bates, 1882
CHIS², HGO²⁴, JAL², MEX², NAY², OAX², PUE², TAB², VER^{2,3,16}
- 0857.** *Bradycellus nitidus* (Dejean, 1829)
BCN¹, BCS^{5,26}, CDMX^{2,5}, CHIH², DGO^{2,5}, HGO^{2,3,24}, JAL^{2,3,6}, MEX², MICH², MOR^{2,5}, NL², OAX^{2,7}, PUE², QRO², SON^{2,54}, TLAX², VER^{2,3}
- 0858.** *Bradycellus nubifer* Leconte, 1858
NL⁷
- 0859.** *Bradycellus obsoletus obsoletus* Say, 1834
CDMX^{4,16}, DGO⁵, MEX^{4,16}, OAX^{4,16}, SLP¹⁶, TLAX¹⁶, VER¹⁶
- 0860[#].** *Bradycellus obsoletus lucidus* Bates, 1878
Type locality: “México”, Bates 1878: 592
- 0861.** *Bradycellus obsoletus melanochrous* Bates, 1882
MEX¹⁶, PUE¹⁶
- 0862.** *Bradycellus obsoletus subobsoletus* Bates, 1878
CDMX¹⁶, OAX¹⁶, SLP¹⁶, VER¹⁶
- 0863^{*}.** *Bradycellus obtusus* Fall, 1905
Type locality: “Southern California (Pasadena, Azusa, Claremont)”, Fall 1905: 176; “México” (Blackwelder

- 1944: 55; Erwin et al. 1977: 4.44)
0864. *Bradycellus rivalis* LeConte, 1858
BCN^{1,2,26}, JAL², OAX²
0865. *Bradycellus rupestris* (Say, 1823)
BCN¹, NL⁷
- 0866[#]. *Bradycellus socors* (Casey, 1914)
BCN⁵⁹, type locality: “Island of Guadalupe, off the coast of Lower California [Norte]”, Casey 1914: 250; “México”³ (Erwin et al. 1977: 4.44)
0867. *Cratacanthus dubius* (Palisot de Beauvois, 1811)
CHIH^{2,3}, COAH², ZAC³
0868. *Dicheirotichus cognatus* (Gyllenhal, 1827)
BCN^{21,26,28}
0869. *Dicheirus dilatatus angulatus* Casey, 1914
BCN^{3,26}
0870. *Dicheirus piceus* (Ménétriés, 1843)
BCN³
0871. *Discoderus aequalis* Casey, 1914
CHIH^{2,49}, COAH^{2,49}, DGO⁵, GTO⁴⁹, NL^{2,49}, SON^{2,49}, TAMPS^{2,49}, VER⁴⁹, ZAC^{2,49}
0872. *Discoderus amblyderus* (Bates, 1878)
CDMX⁵⁹, CHIH⁴⁹, COAH⁴⁹, DGO⁴⁹, GTO^{2,49}, HGO², JAL⁴⁹, MEX⁴⁹, MICH⁴⁹, NL⁴⁹, OAX⁴⁹, PUE¹⁸, QRO⁴⁹, SLP⁴⁹, VER^{18,49}, ZAC⁴⁹
0873. *Discoderus arcuatus* (Putzeys, 1878)
AGS⁴⁹, CHIH⁴⁹, DGO⁴⁹, GTO⁴⁹, HGO⁴⁹, JAL⁴⁹, MEX⁴⁹, MICH⁴⁹, NAY⁴⁹, QRO⁴⁹, SLP⁴⁹, VER⁴⁹, ZAC⁴⁹
- 0874[#]. *Discoderus circulus* Messer and Raber, 2025
CHIH⁴⁹, COAH⁴⁹, DGO⁴⁹, type locality: “México, Chihuahua, w slope Sierra Sta. Eulalia, c. 25 mi NW Guimbaleté”, Messer and Raber 2025: 462
0875. *Discoderus cordicollis* Horn, 1891
BCN^{2,49}, BCS⁵, COAH^{2,3,49}, DGO^{2,5,49}, SIN⁴⁹, SON^{5,49}
0876. *Discoderus crassicollis* Horn, 1891
CHIH^{2,49}, DGO^{2,49}, GTO^{2,49}, MOR⁴⁹, SLP⁴⁹
0877. *Discoderus crassiusculus* (Putzeys, 1878)
CHIH⁴⁹, COL⁴⁹, DGO⁴⁹, GTO⁴⁹, GRO⁴⁹, HGO⁴⁹, JAL⁴⁹, MICH⁴⁹, NOR⁴⁹, NAY⁴⁹, OAX^{16,49}, SON⁴⁹
0878. *Discoderus curvipes* (Putzeys, 1878)
CHIH^{18,49}, NAY⁴⁹, SON^{2,49}
0879. *Discoderus difformipes* Bates, 1878
AGS^{2,49}, GRO⁴⁹, GTO^{2,49}, JAL^{2,49}, MICH^{2,49}, MOR⁴⁹, NAY⁴⁹, OAX^{16,49}, PUE^{2,3,49}, QRO⁴⁹
0880. *Discoderus discoderoides* (Schaeffer, 1910)
DGO^{2,49}, OAX^{2,49}, TAMPS^{2,49}
0881. *Discoderus impotens* (LeConte, 1858)
CHIH⁴⁹, COAH⁴⁹, DGO⁴⁹, NL⁴⁹, SON⁴⁹, ZAC⁴⁹
- 0882[#]. *Discoderus nonarctuatus* Messer and Raber, 2025
SLP^{2,5,49}, type locality: “México, San Luis Potosí, 1.7 mi E C. del Maíz”, Messer and Raber 2025: 470
0883. *Discoderus obsidianus* Casey 1914
BCN^{2,49}, BCS^{2,5,49}, CHIH⁴⁹, COAH^{2,49}, COL^{2,49}, DGO^{2,49}, GRO⁴⁹, MICH^{2,49}, MOR^{5,49}, NAY⁴⁹, OAX^{2,49}, PUE^{2,49}, SIN^{2,49}, SON^{2,5,49}
- 0884[#]. *Discoderus pueblaensis* Messer and Raber, 2025
GRO⁴⁹, OAX⁴⁹, PUE^{2,49}, type locality: “México, Puebla, near Tahuitzingo, Rte. 190”, Messer and Raber 2025: 466
0885. *Discoderus robustus* Horn, 1883
BCN⁴⁹, BCS^{5,49}, CDMX⁵, CHIH^{6,49}, DGO^{18,49}, JAL⁴⁹, MICH⁴⁹, OAX⁴⁹, PUE^{5,4}, SON^{2,6,7,49}
- 0886[#]. *Discoderus shpeleyi* Messer and Raber, 2025; Figure 1R
OAX^{2,49}, type locality: “México, Oaxaca, 27 km SE Oaxaca [city] on Rte. 190”, Messer and Raber 2025: 472
0887. *Discoderus valgus* (Bates, 1882)
CHIS^{2,49}
0888. *Harpalus alienus* Bates, 1878
CDMX⁵, DGO²
0889. *Harpalus amputatus* Say, 1830
HGO^{2,3,5,24}, MEX², NL², OAX⁵, PUE², SLP², ZAC²
0890. *Harpalus aterrimus* Casey, 1914
MEX²
- 0891[#]. *Harpalus atripes* Casey, 1914
CHIH⁵⁹; (type locality: “México (Colonia Garcia in Chihuahua)”, Casey 1914: 95)
0892. *Harpalus caliginosus* (Fabricius, 1775)
CHIH^{2,5}, SON^{2,3,5,54}
0893. *Harpalus desertus* LeConte 1859
SON^{2,54}
0894. *Harpalus durangoensis* Bates, 1891
CHIH², DGO^{2,5,19}
- 0895*. *Harpalus ellipsis* LeConte, 1848
Type locality: “Missouri”, LeConte 1848: 400; “México” (Blackwelder 1944: 48)
0896. *Harpalus erythropus* Dejean, 1829
MICH^{2,3}
0897. *Harpalus fraternus* LeConte, 1852
SON^{2,54}
0898. *Harpalus fuscipalpis* Sturm, 1818
CHIS⁵
0899. *Harpalus gravis* LeConte, 1858
NL²
0900. *Harpalus hebesceus* Bates, 1882
OAX¹⁶
0901. *Harpalus idiotus* Bates, 1889
VER³; **note:** most likely mis-identified or mis-labelled; type locality: “Goorais Valley; Skardo” [Kashmir, India], Bates 1889: 15.
0902. *Harpalus laevipes* Zetterstedt, 1828
CDMX^{3,5}, HGO^{3,5}
0903. *Harpalus liobasis* Chaudoir, 1868
AGS², CHIH², JAL², MEX², QRO²
- 0904*. *Harpalus paratus* Casey, 1924
“Canada and United States”. “According to Ball and Anderson (1962: 76), the species is also found in “México”” (Bousquet, 2012: 1098)
0905. *Harpalus pensylvanicus* DeGeer, 1774
BCN⁵, CHIH^{2,3,6}, COAH⁵, DGO⁵, SON^{2,54}
- 0906*. *Harpalus retractus* LeConte, 1863
Harpalus retractus LeConte 1863: 13, is a replacement name for *Harpalus impiger* LeConte 1854a: 79 (type locality: “Santa Fe, New Mexico” [nec. *Harpalus impiger* Duftschmid, 1812]; “México” (Blackwelder 1944: 48); “The record from “México” (Csiki 1932a: 1186) is probably in error.” (Bousquet 2012: 1104)

- 0907.** *Notiobia brevicollis* Chaudoir, 1837
AGS², BCN⁵, BCS^{5,26}, CDMX^{2,5}, CHIH^{2,3,5,6}, CHIS², COAH², DGO^{2,3}, GRO², HGO^{2,3,5,24}, JAL^{2,5}, MEX^{2,3,4,5}, MICH^{2,3,5}, MOR⁵, NAY⁵, NL^{2,3,4}, OAX², PUE^{2,4,5}, QRO², SIN⁵, SLP², SON^{2,54}, TLAX², VER^{2,3,4}, ZAC²
- 0908.** *Notiobia cooperi* Noonan, 1973
JAL², NAY²
- 0909.** *Notiobia cyanippa* (Bates, 1882)
GRO², JAL^{2,3}, MEX², MICH², MOR^{3,16}, OAX², SIN², SON^{2,54}
- 0910.** *Notiobia disparilis* Bates, 1878
CHIS², VER²
- 0911[#].** *Notiobia ewarti* Noonan, 1973
VER⁵⁹; (type locality: “México, Veracruz, Coyame, Lake Catemaco”, Noonan 1973: 326)
- 0912.** *Notiobia flavicincta* (Erichson, 1847)
CHIS², JAL², OAX², VER²
- 0913.** *Notiobia flebilis* (LeConte, 1863)
BCN^{3,5}, BCS^{2,5}
- 0914.** *Notiobia hilariola* Bates, 1891
GRO^{2,19}
- 0915.** *Notiobia lamprota* (Bates, 1882); Figure 2A
OAX², VER^{2,5,16}
- 0916.** *Notiobia leiroides* Bates, 1878
CHIS^{2,3,7}, COL², GRO², MOR², NAY², OAX^{2,4}, PUE², VER³, ZAC³
- 0917.** *Notiobia maculicornis* (Chaudoir, 1843)
VER⁴
- 0918.** *Notiobia melaena* Bates, 1882
CHIS², OAX², VER^{2,3,5,17,52}
- 0919.** *Notiobia mexicana* Dejean, 1829
BCS^{5,26}, CDMX^{4,5}, CHIH^{2,3,5}, CHIS^{2,3,4,5}, COAH², COL⁴, DGO^{2,3,4,5}, GRO^{2,5}, GTO², HGO^{2,3,5,24}, JAL^{2,3,5}, MEX^{2,3,4,5}, MICH^{2,3,4,5,6,7}, MOR^{2,5}, NAY^{2,3,5,6}, NL^{2,5}, OAX^{2,3,4,5}, PUE^{2,3,4,5}, QRO², SIN², SLP^{2,3,5,7}, SON^{2,3,6,54}, TAMPS⁵, TLAX², VER^{2,4}, ZAC^{2,3,5}
- 0920.** *Notiobia nebrionides* Perty, 1830
CHIS^{2,7}, MOR², QROO², VER²
- 0921.** *Notiobia obscura* Bates, 1882
CHIS², DGO⁵, GRO², JAL², NAY², OAX^{2,16}, VER^{2,7,16}
- 0922.** *Notiobia pallipes* Bates, 1882
CHIS², OAX¹⁶, PUE², TAMPS^{2,3}, VER^{2,16}
- 0923.** *Notiobia purpurascens* (Bates, 1882)
AGS², BCN⁵, BCS⁵, COAH^{2,3}, GRO³, HGO^{2,24}, JAL², MICH², NAY², NL², SLP², SON^{2,54}, VER¹⁶
- 0924.** *Notiobia schlingeri* Noonan, 1973
CDMX⁵, GRO², JAL^{2,3}, MICH², OAX², SON^{2,54}
- 0925.** *Notiobia terminata* (Say, 1823)
BCS⁵, CDMX², CHIH^{2,5}, CHIS^{2,3,5}, COAH², DGO^{2,5}, GRO⁵, GTO², HGO^{2,3}, JAL^{2,5}, MEX², MICH², MOR^{2,3,5}, NAY², NL^{2,3}, OAX^{2,5}, PUE^{2,3,4,5}, QRO², SLP^{2,3,4,5}, SON^{2,54}, TAMPS², VER⁵
- 0926.** *Notiobia virescens* (Dejean, 1831)
CHIS⁵, COL³, GRO², HGO^{3,5}, JAL³, MOR^{2,3,5}, NAY², PUE^{3,5}
- 0927.** *Notiobia viridula* (Dejean, 1829)
CHIS^{2,3,7}, NAY², OAX², TAMPS³, VER²
- 0928.** *Pelmatellus balli* Goulet, 1974
CHIS^{2,5,35}
- 0929.** *Pelmatellus brachyptera* Goulet, 1974
CHIS^{2,3,7,35}
- 0930.** *Pelmatellus cyanescens* Bates, 1882
HGO^{2,3,5,35}, MEX³⁵, OAX³⁵, VER^{2,7,16,35}
- 0931[#].** *Pelmatellus infusca* Goulet, 1974
OAX^{2,35}
- 0932.** *Pelmatellus nitescens* Bates, 1882
CDMX³⁵, CHIS^{2,3,5,35}, DGO³⁵, GRO³⁵, HGO^{2,24}, MEX³⁵, MOR³⁵, OAX^{2,16}, PUE³, SLP^{2,35}, VER^{2,3,4,16,35,52}
- 0933[#].** *Pelmatellus nubicola* Goulet, 1974
CHIS^{2,35}
- 0934.** *Pelmatellus obtusa* Bates 1882
CDMX⁵, CHIS², DGO^{2,5}, GRO², HGO^{2,3}, JAL^{3,6}, MEX^{2,5}, MICH^{2,3,5,6}, OAX², PUE², TLAX², VER^{2,3,52}
- 0935[#].** *Pelmatellus rotundicollis* Goulet, 1974
NL³⁵
- 0936.** *Pelmatellus stenolophoides stenolophoides* Bates, 1882
GRO^{3,35}, JAL³⁵, MEX³⁵, MICH³⁵, MOR³⁵, OAX^{16,35}, SON⁵⁴, VER^{16,35}
- 0937.** *Pogonodaptus mexicanus* (Bates, 1978)
CAMP^{2,4}, GRO^{2,3}, JAL², NAY^{2,5}, SIN⁵, SLP², SON^{2,54}, TAB², TAMPS^{2,3}, VER²
- 0938.** *Polpochila capitata* (Chaudoir, 1852)
AGS^{1,2}, BCN^{1,26}, CDMX^{1,4}, CHIH², DGO^{1,2}, GTO^{1,2}, HGO²⁴, JAL^{1,2}, MEX^{1,2}, OAX¹, YUC², ZAC²
- 0939.** *Polpochila erro* (LeConte, 1854)
CHIH², COAH², DGO^{1,2,3,4,6}, JAL^{2,3,6}, MOR^{2,3,4,5,6}, NAY^{3,5}, SIN^{1,2,3,5,6}, SON^{1,2,3,4,5,6,54}, YUC²
- 0940.** *Polpochila hendrichsi* Nègre, 1967
MICH¹, MOR⁴
- 0941.** *Polpochila rotundicollis* Bates, 1882
AGS¹, GRO¹, GTO^{2,4,16}, JAL^{1,2}, SIN¹, SLP², VER²
- 0942.** *Polpochila schaumii* Nègre, 1963
AGS¹, DGO¹, GTO¹, JAL^{1,2}, MEX¹, OAX^{1,2}
- 0943.** *Selenophorus aeneopiceus* Casey, 1884
CHIH²⁸, COAH², DGO², GTO², JAL², NL², OAX², SLP², SON^{2,54}, TAMPS², ZAC²
- 0944.** *Selenophorus aequinoctialis* Dejean, 1829
BCN²⁶, BCS^{2,3}, CHIS², CHIH^{3,6}, COAH², COL^{2,3}, DGO², GRO², JAL^{2,3}, MOR^{2,4}, NAY², NL², OAX², PUE², QRO², SIN², SLP^{2,3}, SON^{2,3,6,54}, TAMPS², VER^{2,3,4}
- 0945.** *Selenophorus agonoides* (Putzeys, 1878)
MOR⁴
- 0946.** *Selenophorus alternans* Dejean, 1829
SIN⁷
- 0947[#].** *Selenophorus assimilis* Putzeys, 1878
OAX⁵⁹; (type locality: “Oaxaca, [México]”, Putzeys 1878: 44)
- 0948[#].** *Selenophorus aureocupreus* Bates, 1891
MICH¹⁹; (type locality: “Huetamo, Michoacan, [México]”, Bates 1891: 241)
- 0949.** *Selenophorus balli* Messer and Raber, 2021
CAMP^{2,48}, CHIS^{2,48}, COL^{2,48}, GRO^{2,48}, JAL⁴⁸, MOR^{2,48}, NL^{2,48}, OAX⁴⁸, PUE², SIN^{2,48}, SLP^{2,48}, SON^{2,48,54}, TAMPS^{2,48}, VER^{2,48}

0950. *Selenophorus bradycelloides* Bates, 1891
CHIS², TAB^{2,19}
0951. *Selenophorus chaparralus* Purrington, 2000
NL⁴⁸
0952. *Selenophorus concinnus* Schaeffer, 1910
BCN², CHIS⁴⁸, JAL⁴⁸, OAX⁴⁸, SIN^{5,48}, SON^{2,3,6,48,54}
0953. *Selenophorus desalinesii* Ball and Shpeley
GTO⁴; **note**: most likely mis-identified or mis-labelled; type locality: “just north of Dessalines, Antibonite Department, Haiti”, Ball and Shpeley 1992: 102; known only from Haiti and Dominican Republic (Shpeley et al. 2017: 58)
0954. *Selenophorus dichromatus* Casey, 1914
COAH⁴⁸, DGO²
0955. *Selenophorus dispar* Bates, 1891
CHIS², COL², GRO^{2,3,19}, JAL², MOR^{2,19}, NAY², OAX², QRO², SIN², SLP², SON^{2,54}, TAMPS², VER², YUC²
- 0956*. *Selenophorus ellipticus* Dejean, 1829
Type locality: “l’Amerique septentrionale”, Dejean 1829: 108; “Canada, United States, México” (Bousquet 2012: 1138); “México” (Messer & Raber 2021: 25)
0957. *Selenophorus fabricii* Shpeley, Hunting and Ball, 2017
YUC (per. comm. P.W. Messer, 2025)
0958. *Selenophorus faldermanni* Putzeys, 1878
BCN^{2,5}; **note**: most likely mis-identified or mis-labelled; type locality: “Bresil”, Putzeys 1878: 64; “Brazil” (Blackwelder 1944: 49)
0959. *Selenophorus fatuus* LeConte, 1863
GRO³, HGO⁷, JAL³, NL⁷, SLP⁷, SON^{2,54}, TAMPS^{2,3}, VER^{2,3,7}, YUC³
- 0960#. *Selenophorus flavipes* Puteys, 1878
VER⁵⁹; type locality: “Mexique (Cordova)” [= Veracruz, México], Putzeys 1878: 39
- 0961*. *Selenophorus granarius* Dejean, 1829
Type locality: “l’Amerique septentrionale”, Dejean 1829: 110; “México” (Blackwelder 1944: 49; Messer and Raber 2021: 26)
0962. *Selenophorus intermedius* (Putzeys, 1878)
CHIS², COL², GRO², JAL², MEX², MICH², MOR², OAX², VER^{2,3}
0963. *Selenophorus laevicollis* Bates, 1884
GRO², OAX²
0964. *Selenophorus maritimus* Casey, 1914
OAX⁴⁸, SIN⁴⁸
0965. *Selenophorus mexicanus* (Puteys, 1878)
CHIS², TAB²
0966. *Selenophorus obscurus* Puteys, 1878
BCN¹; **note**: most likely mis-identified or mis-labelled; type locality: “Brésil”, Putzeys 1878: 59; “Brazil” (Blackwelder 1944: 50)
0967. *Selenophorus palliatus* (Fabricius, 1798)
BCN¹, CHIH², SLP², TAMPS³
0968. *Selenophorus parumpunctatus* Dejean, 1829
VER⁴; **note**: most likely mis-identified or mis-labelled; type locality [English translation]: “I have had this insect for a very long time; I believe it comes from America and probably the Antilles, but I am not certain.”, Dejean 1929: 105; “known from Florida, Georgia, South Carolina and the West Indies” (Bousquet 2012: 1145); “type area restricted to the island of Hispaniola” (Shpeley et al. 2017: 106); “Florida; West Indies” (Messer and Raber 2021: 40)
0969. *Selenophorus pedicularius* Dejean, 1829
BCN¹, DGO², COAH³, SON^{2,54}
0970. *Selenophorus planipennis* LeConte, 1848
COAH², ZAC²
0971. *Selenophorus pumilus* Messer and Raber, 2021
CHIS², COL², JAL⁴⁸, NAY^{5,48}, OAX^{2,48}, SIN^{3,48}, SON^{2,5,48,54}
- 0972*. *Selenophorus putzeysi* Csiki, 1932
Type locality: “Guatemala”, Csiki 1932a: 1200; “México” (Blackwelder 1944: 50; Erwin et al. 1977: 4.51)
0973. *Selenophorus pyritosus* Dejean, 1829
BCS², CHIS², COL², DGO², GRO^{2,3,7}, JAL^{2,3,6}, MEX², MICH², MOR^{2,7}, NAY², OAX², PUE⁴, SIN^{2,3}, SLP^{2,7}, SON^{2,6,54}, TAB², TAMPS², VER^{2,3}
0974. *Selenophorus rileyi* Messer and Raber, 2021
JAL⁴⁸, MICH⁴⁸, PUE⁴⁸, SIN^{2,48}, SON^{2,48,54}, TAB^{2,48}, VER^{2,48}
0975. *Selenophorus sallei* Putzeys, 1878
DGO², JAL², MEX², QRO², SLP², VER⁴
0976. *Selenophorus semirufus* Bates, 1882
BCN⁵, CHIH², GTO¹⁶, HGO^{2,7,24}, PUE⁴, SLP², SON^{2,54}
0977. *Selenophorus seriatoporus* Putzeys, 1878
CHIS², OAX², SLP^{2,7}, SON⁵⁴, TAB², TAMPS², VER^{2,3}, YUC²
0978. *Selenophorus sinuaticollis* Notman, 1922
SON^{2,54}, TAMPS^{2,3}
0979. *Selenophorus splendidus* Putzeys, 1878
CHIS², VER²
0980. *Selenophorus striatopunctatus* Putzeys, 1878
CHIS², COL², GRO², HGO²⁴, MICH², MOR^{2,5}, NL², OAX², PUE², SIN^{2,5}, SLP^{2,7}, SON^{2,5,54}, TAMPS^{2,3}, VER^{2,3}
0981. *Selenophorus suavis* Bates, 1884
CHIS², CHIH², COL², GRO², HGO², JAL², MICH², NAY², NL², OAX^{2,18}, SIN², SLP², SON², VER¹⁸
0982. *Selenophorus subsinuatus* Putzeys, 1878
CHIS², MOR⁴, SIN², TAMPS³, VER^{2,3}
0983. *Selenophorus tarsalis tarsalis* Putzeys, 1878
CHIS², TAB², VER⁴
0984. *Selenophorus tarsalis liosomus* Bates, 1891
QROO², YUC¹⁹
0985. *Selenophorus tubericauda* Bates, 1884
CDMX¹⁸, TAB⁵⁹, VER¹⁸
0986. *Selenophorus undatus* Messer and Raber, 2021
NL^{2,48}, OAX^{5,48}, TAMPS^{2,48}, VER²
- 0987#. *Selenophorus yucatanus* Putzeys, 1878
YUC⁵⁹; (type locality: “Yucatan”, [México], Putzeys 1878: 24)
0988. *Stenolophus cincticollis* LeConte, 1858
SIN⁵, SON^{3,6}
0989. *Stenolophus comma* (Fabricius, 1775)
CHIH², DGO², NAY², SON^{2,54}
0990. *Stenolophus conjunctus* Say, 1823
BCS⁵, CDMX⁴, CHIH², GTO⁴, HGO⁵, PUE², OAX^{2,5}, SLP², TAB⁵, TLAX², VER², ZAC²

0991. *Stenolophus dissimilis* Dejean 1829
COAH², NL⁵, QRO², SLP², TAMPS^{2,3}, VER³
0992. *Stenolophus flavipes* LeConte, 1858
AGS⁵, BCN^{3,5,6}, BCS⁵, MOR²
0993. *Stenolophus fuliginosus* Dejean, 1829
BCN⁵
0994. *Stenolophus limbalis* LeConte, 1857
BCN^{2,5}
0995. *Stenolophus lineola* (Fabricius, 1775)
CHIH^{3,6}, SON^{2,5,4}
0996. *Stenolophus maculatus* LeConte, 1869
BCN³
0997. *Stenolophus mexicanus* Bates, 1882
MOR³, VER^{4,16}
0998. *Stenolophus ochropepus* (Say, 1823)
BCN², CHIS², COAH², GRO², MEX², MICH², NL²,
OAX², SIN², SON^{2,3,5,4}, TAB², VER^{2,3,6}
0999. *Stenolophus rugicollis* (LeConte, 1869)
VER⁷
1000. *Stenolophus trichotichnoides* Ito, 2000
TAMPS³; **note**: most likely mis-identified or mis-labelled; type locality: “Mon Angget, Chiang Mai, Thailand”, Ito 2000: 124
- 1001*. *Stenomorphus californicus californicus* (Ménétriés, 1843)
Type locality: “California” (lectotype designation by Ball, Shpeley and Currie 1991: 961)
1002. *Stenomorphus californicus rufipes* LeConte, 1858
CHIH^{2,5}, COAH^{3,5}, DGO³, GTO⁴, HGO⁴, JAL^{4,5},
MEX³, MICH^{3,4,5}, MOR^{4,5}, NL², SIN⁴, SLP^{2,5}, SON^{5,5,4},
TAMPS^{2,3,4,5}, VER³
1003. *Stenomorphus convexior* Notman, 1922
JAL^{2,4,5}, NAY^{2,5}, SIN^{2,3,4,5}, SON^{2,5,4}
1004. *Stenomorphus penicillatus* Darlington, 1936
COL^{2,3,7}, GRO^{2,5}, JAL², MICH^{2,3,4,5}, MOR^{2,3,5}, OAX³,
SIN²
1005. *Stenomorphus sinaloae* Darlington, 1936
BCN¹, BCS⁵, GRO⁵, SIN^{2,3,5,7}, SON^{5,5,4}
1006. *Trichopselaphus erwinorum* Ball, 1978
SON^{2,5,4}
1007. *Trichopselaphus meyeri* Ball, 1978
CHIS¹, VER^{1,5}
1008. *Trichopselaphus minor* Bates, 1882
VER^{2,3,16}
1009. *Trichotichnus nitidulus* (Chaudoir, 1843)
COAH⁷

32. Sphodrini

1010. *Calathus ambigens ambigens* Bates, 1891
CHIH², DGO^{1,2,3,5,19}
1011. *Calathus ambigens serricola* Ball and Nègre, 1972
DGO^{1,2,3,5}
1012. *Calathus ambigens tarahumarus* Ball and Nègre, 1972
CHIH^{1,3,5}
- 1013*. *Calathus ambiguus* (Paykull, 1790)
“México”¹; no type locality/area by Paykull 1790: 130;
“?México” (Csiki 1931: 773; Blackwelder 1944: 41);
note: all indications are that this is a palaearctic species

[GBIF: <https://www.gbif.org/species/5755293>]

1014. *Calathus aztec* Ball and Nègre, 1972
CDMX^{2,3}, JAL^{1,2,3}, MEX^{1,2,3}, MICH^{1,2,3}, PUE^{1,2},
VER^{1,2,3,5}
1015. *Calathus bolivari* Nègre, 1970
MEX^{1,2,3,5}, MICH¹, PUE²
1016. *Calathus clauseni* Ball and Nègre, 1972
VER^{1,3}
1017. *Calathus durango* Ball and Nègre, 1972
DGO^{1,2,3,5}
1018. *Calathus erwini* Ball and Nègre, 1972
JAL^{1,2,3,5}, MICH^{1,2}
1019. *Calathus gregarius* (Say, 1823)
HGO^{2,4}, SON^{3,6}
1020. *Calathus leechi* Ball and Nègre, 1972
MEX^{2,3,5}, MICH^{2,3}
1021. *Calathus marmoreus* Ball and Nègre, 1972
HGO^{1,2,3,5}, QRO²
- 1022#. *Calathus mcclevei* Shpeley and Van Devender, 2021; Figure 2B
SON^{2,5,5,4}; (type locality: “México, Sonora, Municipality de Yecora”, Shpeley and Van Devender, 2022: 101)
1023. *Calathus mexicanus* Chaudoir, 1837
CDMX^{1,2}, HGO^{1,2,4}, MEX^{1,2}, MICH^{1,2}, MOR¹, OAX¹,
PUE^{1,2,4}, TLAX², VER^{1,2,3,5}
1024. *Calathus opaculus* LeConte, 1854
SON^{5,4}; (type locality: “Middle, Southern and Western States”, LeConte 1854b: 37)
1025. *Calathus ovipennis* Putzeys, 1873
OAX^{1,2,3,5}
1026. *Calathus potosi* Ball and Nègre, 1972
COAH^{1,2}, NL^{1,2,3,5}
1027. *Calathus rotgeri* Ball and Nègre, 1972
NL^{1,2}
1028. *Calathus ruficollis rufficollis* Dejean, 1828
BCN^{1,2,3,5}, SIN³, SON^{2,5,4}
1029. *Calathus ruficollis quadalupensis* Casey, 1897
BCN³ (Guadalupe Island)
- 1030#. *Calathus tigrinus* Shpeley and Van Devender, 2022
SON^{2,5,5,4}; (type locality: “México, Sonora Sierra El Tigre”, Shpeley and Van Devender, 2022: 102)
1031. *Calathus whiteheadi* Ball and Nègre, 1972
DGO^{1,2,3,5}
1032. *Laemostenus algerinus* Gory, 1833
DGO³; **note**: most likely mis-identified or mis-labelled; type locality: “Algeria”, Gory 1833: 232
1033. *Laemostenus complanatus* Dejean, 1828
BCN³, CDMX^{3,4}, GTO³, HGO³, MEX³, VER³
- 1034#. *Miquihuana rhadiniformis* Barr, 1982
TAMPS^{1,13}; (type locality: “Sotano de Riachuelo, Tamaulipas, México”, 1982: 188)
1035. *Synuchus semirufus* (Casey, 1913)
CHIH^{2,3}, SON^{2,5,4}

33. Platynini

1036. *Agonum anthracinum* Dejean, 1831; Figure 2C
CDMX^{1,4,5}, CHIH^{1,2,4,5}, COL^{1,2,5,4,5}, DGO^{1,5,4,5}, GTO^{1,2,4,5},
GRO^{1,2,4,5}, HGO^{1,4,5}, JAL^{1,2,5,4,5}, MEX^{1,2,3,5,4,5}, MICH^{1,2,3,5,4,5},
MOR^{1,2,5,4,5}, NAY^{1,5,4,5}, OAX^{1,2,5,4,5}, PUE⁴, QRO^{1,2,4,5},

- SIN^{1,2,5,45}, SON^{1,2,54,45,54}, VER^{1,45}
- 1037[#]. *Agonum colonicum* Casey, 1920
CHIH^{1,59}; (type locality: “México (Colonia García, Sierra Madre Mts., Chihuahua)”, Casey, 1920: 94)
- 1038[#]. *Agonum curtipenne* Casey, 1920
MOR⁵⁹; (type locality: “México (Tres Marías, Morelos)”, Casey 1920: 27)
1039. *Agonum cyanopis* (Bates, 1882)
CDMX^{1,16,45}, CHIH^{1,2,45}, DGO^{1,2,4,5,45}, GRO^{1,45}, GTO^{1,2,16,45}, HGO^{1,2,45}, JAL^{1,5,45}, MEX^{1,2,45}, MICH⁴, MOR^{1,45}, NAY^{1,5,45}, NL², OAX^{1,2,45}, PUE^{1,4,5,16,45}, SLP^{1,45}, SON^{1,2,5,45,54}, TAMPS¹, VER^{1,45}, ZAC^{1,2,45}
1040. *Agonum cyclifer* (Bates, 1884)
CDMX¹⁸, DGO^{1,2,45}, NL^{1,2,45}, SLP^{1,45}, SON^{2,54}
1041. *Agonum decorum* (Say, 1823)
BCN^{1,5,26,45}, BCS^{5,7}, CDMX^{1,45}, CHIH^{1,3,45}, CHIS^{1,2,45}, DGO^{1,2,45}, GTO^{1,2,45}, JAL^{1,2,45}, MEX^{1,2,45}, MICH^{1,2,45}, MOR^{1,5,45}, NAY^{1,2,5,45}, NL^{1,2,45}, OAX^{1,45}, PUE^{1,2,7,45}, QRO^{1,2,45}, SIN^{1,5,45}, SLP^{1,2,45}, SON^{1,2,3,5,7,45,54}, TAB^{1,2,45}, TAMPS^{1,2,3,45}, VER^{1,2,45}, ZAC^{1,2,45}
1042. *Agonum districtum* Casey, 1920
MOR⁴
1043. *Agonum extensicolle* (Say, 1823)
AGS^{1,2,45}, BCS²⁶, CHIH^{1,2,3,5,45}, COAH³, DGO^{1,2,45}, HGO^{1,2,45}, JAL^{1,2,45}, OAX⁴, PUE⁴, SLP^{1,45}, SON^{1,2,3,5,54,45,54}, TAMPS⁴, ZAC^{1,2,5,45}
1044. *Agonum extimum* Liebherr, 1886
CHIH^{1,2,5,45}, COL¹, COAH^{2,3,4,5,6,45}, SON²
1045. *Agonum fossiger* Dejean, 1828
BCN⁵
- 1046[#]. *Agonum infidum* Casey, 1920
MOR⁵⁹; (type locality: “México (Tres Marías, Morelos)”, Casey 1920: 26)
1047. *Agonum leptodes* Bates, 1882
DGO¹⁶
- 1048[#]. *Agonum logicum* Casey, 1920
MOR⁵⁹; (type locality: “México (Tres Marías, Morelos)”, Casey 1920: 24)
- 1049[#]. *Agonum lymphaticum* Casey, 1920
CDMX⁵⁹; (type locality: “México (Federal District)”, Casey 1920: 44)
1050. *Agonum megilum* Bates, 1891
DGO¹⁹
- 1051[#]. *Agonum morelosense* Casey, 1920
MOR⁵⁹; (type locality: “México (Tres Marías, Morelos)”, Casey 1920: 25)
1052. *Agonum pacificum* Casey, 1920
BCN^{1,5,26,45}
1053. *Agonum parextimum* Liebherr, 1886
SIN^{1,3,5,45}, SON^{1,2,5,45,54}
1054. *Agonum patinale* Bates, 1882
CDMX^{1,45}, CHIS^{1,2,3,5,45}, COL⁵, GRO^{2,45}, GTO¹, JAL^{1,45}, MEX^{1,45}, MOR^{1,2,16,45}, NAY^{1,2,45}
1055. *Agonum placidum* (Say, 1823)
AGS^{1,5,45}, CDMX^{1,2,5,45}, CHIH^{1,2,5,45}, COAH^{2,45}, COL¹, DGO^{1,2,5,45}, HGO^{1,2,5,24,45}, JAL^{1,2,45}, MEX^{1,2,4,5,45}, MICH^{1,2,3,4,5,45}, MOR^{1,45}, NL^{1,2,45}, OAX^{1,2,45}, PUE^{1,2,3,45}, QRO^{1,2,45}, SLP^{1,45}, SON^{1,2,45,54}, TAMPS^{1,2,45}, TLAX^{1,2,3,45}, VER^{1,2,3,45}
1056. *Agonum propinquum* (Gemminger and Harold, 1868)
CDMX^{1,45}; (type locality: “Massachusetts”, LeConte 1846: 226; *Agonum propinquum* Gemminger and Harold 1868: 375 is a replacement name for *Agonum piceum* LeConte 1846: 226)
1057. *Agonum punctiforme* (Say, 1823)
CHIH^{1,45}, CHIS^{1,2,5,45}, GTO^{1,4,45}, HGO^{1,3,24,45}, MEX^{1,2}, MICH^{1,2,45}, NL^{1,2,45}, SLP^{1,2,45}, SON^{1,2,45,54}, TAMPS^{1,2,45}
1058. *Agonum scutifer* (Bates, 1878)
CDMX^{1,2,45}, CHIH^{1,2,45}, DGO^{1,2,45}, HGO^{1,5,24,45}, JAL^{1,2,4,5,45}, MEX^{1,2,5,45}, MICH^{1,3,4,45}, PUE^{1,2,45}, TAMPS¹, VER^{1,4,45}
1059. *Agonum suturale* Say, 1834
AGS^{1,45}, CHIH^{1,2,45}, DGO^{1,45}, MEX^{1,45}, PUE⁴, TAMPS¹, TLAX^{3,45}
1060. *Agonum texanum* LeConte, 1878
CHIH^{1,2,5,45}, CHIS^{1,2,45}, COAH^{2,5,45}, COL¹, DGO^{1,2,45}, SON^{1,2,3,5,6,45,54}, TAMPS^{1,2,45}, VER³
- 1061[#]. *Agonum umbripenne* Casey, 1920
MOR⁵⁹; (type locality “México (Cuernavaca), [Morelos]”, Casey 1920: 17)
1062. *Anchomenus capensis* Liebherr, 1991
BCN¹, BCS^{3,5}
1063. *Anchomenus funebris* (LeConte, 1854)
BCN^{1,5}
1064. *Dyscolus acuminatus* (Chevrolat, 1835)
DGO², GTO⁴, MOR⁵, NAY², NL², OAX^{1,2,3,5}, PUE³, SIN^{1,2}, SLP^{2,3}, SON^{2,54}, VER^{1,2,3,5}
1065. *Dyscolus acutulus* (Bates, 1891)
CHIS^{1,3,7}, HGO^{1,19}, PUE³, SLP¹, VER^{2,3,50,52}
1066. *Dyscolus agilis* (Chaudoir, 1878)
PUE⁵⁹, VER⁴
1067. *Dyscolus amplicollis* (Chaudoir, 1878)
PUE², VER^{1,2}
1068. *Dyscolus angulosus* (Chaudoir, 1878)
MEX^{1,2}, OAX²
1069. *Dyscolus aphaedrus* (Chaudoir, 1859)
CHIS^{1,2}, OAX^{1,2}, VER^{1,2,3,52}
1070. *Dyscolus atratus* (Chaudoir, 1859)
CHIS^{1,2}, GRO¹, OAX², PUE², VER^{2,3,52}
1071. *Dyscolus bacatellus* (Liebherr, 1992)
CHIS^{1,2,3}
1072. *Dyscolus bicolor* (Chaudoir, 1879)
CHIS^{1,2,5}, DGO⁵, GRO^{2,3}, JAL², MEX², MICH², MOR^{2,4}, NAY^{1,2}, OAX^{2,5}, PUE^{1,2}, SIN², TAMPS³
1073. *Dyscolus biovatus* (Chaudoir, 1878)
OAX^{1,2}, VER⁵²
1074. *Dyscolus brachyderus* (Chaudoir, 1878)
CHIS^{1,2,3}, VER^{1,2}
1075. *Dyscolus brullei* (Chaudoir, 1837)
CHIS^{1,2}, MICH⁴, VER^{1,2,4}
1076. *Dyscolus brunnellus* (Liebherr, 1992)
OAX^{1,2}, QRO^{1,2}, SLP^{1,2}
1077. *Dyscolus caeruleomarginatus* (von Mannerheim, 1837)
CHIS^{2,7}, OAX², PUE², VER^{2,4}
1078. *Dyscolus caeruleus* (Chaudoir, 1859)
CDMX^{4,5}, JAL², MEX^{2,5}, MICH^{2,3,4,5}, MOR^{4,5}, NAY², OAX^{2,3,5}, PUE⁴, QRO², SIN², SLP², TAMPS^{2,3}

- VER^{1,2,3,4,52}
1079. *Dyscolus caerulipennis* (Liebherr, 1992)
DGO^{1,2}, GRO², GTO¹, JAL^{1,2,3}, MEX², MICH^{1,2,4},
MOR^{1,3}, NAY^{1,2}, OAX^{1,2}, SIN^{1,2}, ZAC^{1,2}
1080. *Dyscolus cazieri* Liebherr and Will, 1996
SON³
1081. *Dyscolus championi* (Bates, 1882)
CHIS^{1,2}
1082. *Dyscolus chaudiroidi* (Coquerel, 1866)
CHIS^{2,3}, VER^{1,59}
1083. *Dyscolus columbinus* (Chaudoir, 1878)
CHIS², VER^{1,2,3,52}
1084. *Dyscolus concisus* (Bates, 1878)
CDMX⁴, CHIH², DGO², GRO², HGO², JAL²,
MEX^{1,2,3,5}, MICH², MOR², OAX², PUE², QRO², SIN²,
SLP², VER^{2,3}
- 1085[#]. *Dyscolus cordatus* (Chaudoir, 1859)
VER¹; (type locality: “Toxpam [= Tuxpan], and Cordova, Veracruz, México”, Chaudoir 1859: 337)
1086. *Dyscolus crypticulus* (Liebherr, 1992)
CHIS^{1,2}, OAX^{1,2}
1087. *Dyscolus cupripennis* (Laporte de Castelnau, 1835)
HGO^{1,24}, VER^{1,2,3,4,52}
1088. *Dyscolus cyanides* (Bates, 1882)
CHIS^{1,2}, MOR^{1,2}, OAX^{1,2}
1089. *Dyscolus cycloderus* (Chaudoir, 1859)
CHIS^{1,2,3,5}, OAX², SIN²
1090. *Dyscolus decorellus* (Liebherr, 1992)
CHIS^{1,2}, GRO^{1,4}, JAL¹, MICH^{1,2}, MOR^{1,2}, OAX^{1,2}
1091. *Dyscolus delicatulus* (Chaudoir, 1878)
VER^{3,50}
- 1092[#]. *Dyscolus deyrollei* (Chaudoir, 1878)
Type locality: “México”, Chaudoir, 1878: 336
1093. *Dyscolus dilutus* (Chaudoir, 1859)
OAX^{1,2}, VER^{1,2}
1094. *Dyscolus dominicensis* (Bates, 1882)
CHIS^{2,3,7}, HGO², OAX^{1,2,16}, PUE², VER^{1,2,3,4,16}
1095. *Dyscolus durangensis* (Bates, 1882); Figure 2D
DGO^{1,2,5,16}
- 1096*. *Dyscolus dyschromus* (Chaudoir, 1878)
“México”¹; (type locality: “Nouvelle Grenada [= Colombia and Panama]”, Chaudoir 1878:303)
1097. *Dyscolus ebeninus* (Chaudoir, 1878)
GRO¹, PUE², VER¹
- 1098[#]. *Dyscolus elliptolellus* (Liebherr, 1992)
OAX¹; (type locality: “5 km N Suchixtepec, Oaxaca, México”, Liebherr 1992: 30)
1099. *Dyscolus erythrocerus* (Chaudoir, 1859)
OAX^{1,2}, VER^{1,2}
1100. *Dyscolus falli* (Darlington, 1936)
MICH², SON^{2,54}
1101. *Dyscolus femoralis* (Chaudoir, 1878)
BCN³, BCS⁵, CDMX⁵, CHIH⁴, CHIS², DGO^{1,2,5}, JAL¹,
MOR^{1,2}, MEX¹, OAX^{1,2}
1102. *Dyscolus fragilis* (Chaudoir, 1878)
CHIS¹, VER⁵²
1103. *Dyscolus fratellus* (Chaudoir, 1878)
CAMP^{1,2}, CHIS^{1,2,5}, GRO^{2,3}, JAL^{2,3}, MEX^{2,5}, MICH^{2,3},
NAY^{2,3}, OAX^{1,2,3}, SIN^{2,3}, TAMPS^{1,2}, VER^{1,2,3,52}
1104. *Dyscolus gracilis* (Chaudoir, 1859)
VER^{1,3,50}
1105. *Dyscolus gratus* (Bates, 1878)
PUE⁴
1106. *Dyscolus haptoderoides* (Bates, 1891)
GRO^{1,2,19}, HGO², OAX²
1107. *Dyscolus harpaloides* (Bates, 1891)
CHIS³, GRO^{1,2,3,19}, OAX²
1108. *Dyscolus imitativus* (Liebherr, 1992)
CHIS^{1,2}
1109. *Dyscolus iricolor* (Bates, 1882)
CHIS^{1,2,3,7}
1110. *Dyscolus laetiusculus* (Chaudoir, 1878)
CHIS², OAX¹
- 1111[#]. *Dyscolus limbicollis* (Chaudoir, 1878)
OAX^{1,59}; (type locality: “Oaxaca, Mexique”, Chaudoir 1878: 363)
1112. *Dyscolus longipes* (Chaudoir, 1878)
CAMP¹, COL², GRO^{1,2,3}, JAL^{1,2,5}, MOR¹, NAY^{1,2,5},
OAX^{2,3}, PUE⁵⁹
1113. *Dyscolus lucilius* (Bates, 1884)
OAX¹, VER^{1,2,18}
1114. *Dyscolus lugens* (Dejean, 1831)
CDMX², MEX^{1,2,3,5}, MICH², MOR^{1,2}, OAX²
1115. *Dyscolus lyrophorus* (Chaudoir, 1878)
VER^{1,2,52}
1116. *Dyscolus machetellus* (Liebherr, 1992)
CHIS^{2,3}, VER^{1,3}
1117. *Dyscolus macrourus* (Chaudoir, 1878)
CHIS^{1,2,3,7}
1118. *Dyscolus marginicollis* (Chaudoir, 1859)
HGO^{1,24}, VER^{1,59}
1119. *Dyscolus marginissimus* (Liebherr, 1992)
CAMP^{1,2}, CHIS^{1,2,3,4,7}, GRO^{1,2}, JAL^{2,3}, MEX¹, OAX^{1,2,3},
QRO¹, SLP^{1,2}, TAMPS^{1,2,3}, VER^{2,3}, YUC^{1,2,3}
- 1120[#]. *Dyscolus mateui* (Moret, 1991)
DGO^{1,59}; (type locality: “La Ciudad, Sierra de Durango, Durango, México”, Moret 1991: 67)
1121. *Dyscolus megalops* (Bates, 1882)
BCS⁵, CHIS^{2,5}, DGO^{2,5}, GTO^{1,16}, JAL^{2,5}, MOR^{2,3,5,6},
NAY⁵, OAX^{1,2,3,6}, SLP⁷, SON^{2,3,6,54}, VER²
1122. *Dyscolus melanocnemis* (Chaudoir, 1878)
CHIS^{1,2}, GRO¹, OAX^{1,2}, VER²
1123. *Dyscolus metallicus* (Chaudoir, 1859)
CHIS^{2,4}, VER^{1,4}
1124. *Dyscolus minimus* (Bates, 1884)
HGO¹⁸
1125. *Dyscolus minusculus* (Liebherr, 1992)
VER^{1,2,3}
1126. *Dyscolus moestus* (Dejean, 1831)
CDMX^{2,5}, CHIH², HGO^{3,5}, JAL², MEX^{2,3,4}, MICH^{2,4,5},
MOR^{1,2,3}, OAX^{2,5}, PUE^{2,5}, QRO⁵ (Tres Marias Islands), SIN², TAB⁵, TLAX², VER³
1127. *Dyscolus monachus* (Dejean, 1831)
CDMX⁵, GRO^{2,5}, GTO⁴, JAL^{2,3,4,5}, MEX^{2,4,5}, MICH^{2,5},
MOR^{1,2,5}, NAY², PUE^{1,4}, QRO²
1128. *Dyscolus niger* (Chaudoir, 1859)
CHIS^{2,7}, VER^{1,2}

1129. *Dyscolus nitidulus* Liebherr, 1992
CHIS⁷
1130. *Dyscolus nitidus* (Chaudoir, 1837)
CHIS³, MEX⁵, NL⁴
1131. *Dyscolus nugax* (Bates, 1878)
CDMX^{1,2}, GRO², MEX^{1,2,5}, MICH², MOR², OAX²,
PUE^{2,3,4}, TLAX²
1132. *Dyscolus nyctimus* (Bates, 1884)
MEX^{5,18}, MICH⁵, MOR⁵
1133. *Dyscolus obscurus* (Chaudoir, 1859)
CHIS^{1,2,3,5}, DGO², JAL², MEX², MICH², NAY², OAX⁵,
QRO², TAMPS³, VER^{1,2,52}, ZAC²
1134. *Dyscolus olivaceus* (Chaudoir, 1878)
CHIS¹, GRO^{2,3}, JAL², MICH², OAX², VER^{1,2}
1135. *Dyscolus omaseoides* (Bates, 1891)
GRO^{1,2,19}
1136. *Dyscolus orbicollis* (Chaudoir, 1859)
CHIS^{1,2,3,7}, OAX^{1,2}, VER^{1,2}
1137. *Dyscolus ovatus* (Bates, 1884)
CHIH^{1,2,3,18}, SON^{1,2,3,5,54}
1138. *Dyscolus pallidipes* (Chaudoir, 1850)
CHIS^{1,2}, OAX², SLP⁷, VER^{1,2,3,52}
1139. *Dyscolus pectoralis* (Chaudoir, 1878)
VER^{1,59}; (type locality: “Orizaba [Veracruz]
(Mexico)”, Chaudoir 1878: 353)
- 1140[#]. *Dyscolus phaeocnemis* (Chaudoir, 1878)
Type locality: “Mexico”, Chaudoir 1878: 358
1141. *Dyscolus phaeolomus* (Chaudoir, 1878) (**note**: GBIF
has the year as 1879)
CAMP^{1,2}, CHIS^{1,2}, OAX², SLP^{1,2}, TAMPS^{1,2}, VER^{1,2}
1142. *Dyscolus picicornis* (Chaudoir, 1878)
CHIS⁷, OAX¹, VER¹
1143. *Dyscolus platynellus* (Liebherr, 1992)
OAX^{1,2,3}
1144. *Dyscolus platysmoides* (Bates, 1891)
GRO^{1,19}
1145. *Dyscolus porrectus* (Chaudoir, 1878)
NL³, OAX^{2,3}
1146. *Dyscolus pristonichoides* (Chaudoir, 1878)
GRO², OAX²
1147. *Dyscolus procephalus* (Bates, 1878)
CHIS^{1,2}
1148. *Dyscolus purpuratus* (Reiche, 1842) (**note**: GBIF has
the year as 1843)
CHIS^{1,3,5,7}, VER²
1149. *Dyscolus purulensis* (Bates, 1882)
CHIS^{1,2}
1150. *Dyscolus quadrilaterus* (Bates, 1882)
MEX¹, VER^{1,2,3,16}
1151. *Dyscolus rectilineus* (Bates, 1891)
GRO^{1,2,19}, OAX²
1152. *Dyscolus reflexicollis* (Chaudoir, 1859)
CHIS³, VER^{1,52}
- 1153[#]. *Dyscolus reflexus* (Chaudoir, 1859)
VER^{1,59}; (type locality: “Cordova, Veracruz, México”,
Chaudoir 1878: 347)
1154. *Dyscolus robustulus* (Liebherr, 1992)
TAMPS^{1,2,3}
1155. *Dyscolus rotundatulus* (Liebherr, 1992)
JAL^{1,2,3}, MICH^{1,2,3}, MOR^{1,2}, NAY^{1,2}
- 1156[#]. *Dyscolus rubidus* (Chaudoir, 1878)
OAX^{1,59}; (type locality: “Mexico (Oaxaca)”, Chau-
doir 1878: 329)
1157. *Dyscolus ruficornis* (Chaudoir, 1859)
OAX², SIN³, VER^{1,2,3}
1158. *Dyscolus rufiventris* Van Dyke, 1926
CHIH³, SIN³, SON^{3,6,54}
1159. *Dyscolus rufulus* Bates, 1884
CHIS³, OAX⁷
1160. *Dyscolus scabricollis* (Bates, 1882)
CHIS^{2,3,7}, VER¹
1161. *Dyscolus segregatus* (Bates, 1891)
COL^{1,2,5}, GRO^{1,2,7}, JAL^{2,5}, MEX^{4,5}, MICH^{1,2,4,5,7},
MOR^{1,2,5,19}, OAX^{1,2,5,7}, PUE², SIN⁵, SON⁵
1162. *Dyscolus semiopacus* (Chaudoir, 1878)
CHIH², DGO², GRO², OAX^{1,2}, PUE²
1163. *Dyscolus severus* (Chaudoir, 1878)
CHIS^{1,2,3,7,11}
1164. *Dyscolus sexfoveolatus* (Chaudoir, 1878)
CHIS^{1,2}, VER^{3,50}
1165. *Dyscolus sphodroides* (Chaudoir, 1859)
GRO², MEX^{2,3}, OAX^{1,2}, VER¹⁶
1166. *Dyscolus spinifer* (Bates, 1882)
CHIS^{1,2,3}, VER^{2,52}
1167. *Dyscolus stenophthalmus* (Liebherr, 1992)
CHIS^{1,2,3,7}
1168. *Dyscolus steropoides* (Bates, 1891)
GRO^{1,2,19}
1169. *Dyscolus striatopunctatus* (Chaudoir, 1859)
CHIS², VER^{1,2}
1170. *Dyscolus subauratus* (Bates, 1882)
GRO², PUE¹, VER^{1,16}
1171. *Dyscolus subcyaneus* (Chaudoir, 1878)
OAX¹, PUE¹, VER¹
1172. *Dyscolus subviolaceus* (Chaudoir, 1842)
OAX^{1,2}, PUE², VER¹
1173. *Dyscolus tenuicornis* (Chaudoir, 1859)
CHIS², OAX², VER^{1,2}
1174. *Dyscolus teter* (Chaudoir, 1878)
CHIS^{1,2,3}
1175. *Dyscolus tinctipennis* (Bates, 1891)
DGO^{5,19}
1176. *Dyscolus transfuga* (Chaudoir, 1878)
OAX^{1,2}
1177. *Dyscolus transversicollis* (Chaudoir, 1859)
CHIS¹, OAX², VER^{1,2,52}
1178. *Dyscolus valens* (Bates, 1891)
CHIH², DGO^{1,2,19}, SIN², SON^{2,54}, ZAC²
- 1179[#]. *Dyscolus validus* (Chaudoir, 1859)
VER¹; (type locality: “Mexico”, Chaudoir 1859: 343)
1180. *Dyscolus variabilis* (Chaudoir, 1837)
CHIS^{2,3,5}, GRO², GTO¹, HGO², JAL^{1,2}, MEX^{2,4,5},
MICH⁴, OAX^{1,2}, PUE^{2,3}, QRO², TAMPS^{1,2}, VER^{2,3,4,52}
- 1181[#]. *Dyscolus violaceipennis* (Chaudoir, 1859)
VER^{1,59}; (type locality: “Veracruz, México”, Chaudoir
1859: 341)
- Note:** Erwin’s superscript ¹ for *Elliptoleus* refers to Li-

- ebherr, 1991.
1182. *Elliptoleus acutesculptus* Bates, 1882
CDMX¹⁶, CHIH^{1,2}, COAH¹, COL¹, DGO^{1,2,4}, NL², SLP^{1,2}, ZAC^{1,2}
1183. *Elliptoleus balli* Liebherr, 1991
JAL^{2,3}, MEX^{1,2}, MICH^{1,2}
- 1184#. *Elliptoleus corvus* Liebherr, 1991
MEX^{1,59}; (type locality: “México. Mexico [state], Crc. de Tlalmananco [= Tlalmanalco]”, Liebherr 1991: 97); Liebherr (1991: 97) also states that the locality label incorrectly reads “D.F.” [Distrito Federal]
1185. *Elliptoleus crepericornis* Bates, 1882
OAX^{1,2,3,16}
1186. *Elliptoleus curtulus* Bates, 1882
OAX^{1,2}
1187. *Elliptoleus luteipes* Csiki, 1931
VER¹
1188. *Elliptoleus olithopoides* Bates, 1891
CHIH², DGO^{1,2,5,19}, SIN¹
1189. *Elliptoleus tequilae* Liebherr, 1991
JAL^{1,2}
1190. *Elliptoleus vixstriatus* (Bates, 1878)
CDMX^{1,2}, MEX^{1,2,4,5,7}, MICH^{1,2,5}, MOR^{1,2,4}, PUE^{1,2}, TAMPS¹, TLAX², VER^{1,2}
1191. *Elliptoleus whiteheadi* Liebherr, 1991
GRO^{1,2}
1192. *Elliptoleus zapotecorum* Liebherr, 1991
OAX^{1,2,3}
1193. *Glyptolenoides purpuripennis* (Chaudoir, 1878)
CHIS², GRO², HGO²⁴, OAX², TLAX²⁷, VER^{2,3,52}
1194. *Glyptolenus ater* (Chaudoir, 1858)
TAMPS², VER^{1,2}
1195. *Glyptolenus latitarsis* Bates, 1884
CHIS^{1,2}, OAX²
1196. *Hemiplatynus chihuahuae* (Bates, 1884)
CHIH^{1,18}, SIN^{3,6}, SON^{2,54}
1197. *Mexisphodrus boneti boneti* (Bolívar y Pieltain and Hendrichs, 1964)
NL^{1,4,13}
- 1198#. *Mexisphodrus boneti monterreyensis* Barr, 1982
NL^{1,13}
- 1199#. *Mexisphodrus boneti nortoni* Barr, 1982
NL^{1,13}
- 1200#. *Mexisphodrus boneti palmitensis* Barr
NL^{1,13,53}
- 1201#. *Mexisphodrus canuc* Barr, 1982
CHIS^{1,13}
- 1202#. *Mexisphodrus cuetzalan* Barr, 1982
PUE^{1,13}
1203. *Mexisphodrus gertschi gertschi* Hendrichs and Bolívar y Pieltain, 1966
HGO^{1,4,13}
- 1204#. *Mexisphodrus gertschi ahuacatlan* Barr, 1982
SLP^{1,13}
- 1205#. *Mexisphodrus gertschi camposantos* Barr, 1982
QRO^{1,13}
- 1206#. *Mexisphodrus gertschi sprousei* Barr, 1982
SLP^{1,13}
- 1207#. *Mexisphodrus profundus* (Barr, 1966)
TAMPS^{1,13}
- 1208#. *Mexisphodrus purgatus* Barr, 1982
TAMPS^{1,13}
- 1209#. *Mexisphodrus spiritus* Barr, 1982
SLP^{1,13}
- 1210#. *Mexisphodrus tlamayensis* Barr, 1966
SLP^{1,59}; (type locality: “Sótano de Tlamaya, near village of Tlamaya, municipality Xilitla, San Luis Potosí, México”, Barr 1966: 113)
1211. *Mexisphodrus urquijoi* Hendrichs and Bolívar y Pieltain, 1973
OAX^{1,4,13}
1212. *Mexisphodrus veraecrucis* Barr, 1965
VER^{1,3,13}
- 1213#. *Mexisphodrus zoquitlan* Barr, 1982
PUE^{1,13}
- Note:** Erwin’s superscript ¹ for *Onypterygia* species refers to Whitehead and Ball 1997.
1214. *Onypterygia aeneipennis* Chaudoir, 1878
GRO², MOR², OAX^{1,2,3,5}, SLP², VER²
- 1215#. *Onypterygia amecameca* Whitehead and Ball, 1997
MEX^{1,59}; (type locality: “Amecameca, state of Mexico, México”, Whitehead and Ball 1997: 317)
1216. *Onypterygia angustata* Chevrolat, 1835
CDMX¹, CHIS^{1,2,3,7}, COL¹, HGO^{1,24}, JAL^{1,2}, MICH^{1,2,5}, OAX^{1,2}, PUE^{1,2,4,5}, VER^{1,2}
- 1217#. *Onypterygia atoyac* Whitehead and Ball, 1997
GRO¹; (type locality: “73.8 km northeast Atoyac de Alvarez, Guerrero, México”, Whitehead and Ball 1997: 318)
1218. *Onypterygia batesi* Whitehead and Ball, 1997
HGO^{1,24}, JAL³, OAX^{1,2}, QRO^{1,4}, SLP⁵, TAMPS^{1,2,3}, VER¹
1219. *Onypterygia chrysur* Bates, 1882
CHIS¹; (type locality: (“Guatemala, Zapote, San Geronimo, Cerro Zunil”, Bates 1882: 7131; type locality restricted to: “San Geronimo, Baja Verapaz, Guatemala”, Whitehead and Ball, 1997: 357)
1220. *Onypterygia cupricauda* Casey, 1920
GRO^{1,2,5}
1221. *Onypterygia cyanea* Chaudoir, 1878
CDMX⁵, CHIH¹, COL^{1,3}, DGO¹, GRO^{1,5,3}, JAL¹, MEX^{1,5}, MICH¹, MOR^{1,5,3}, NAY¹, OAX^{1,5}, SIN¹, SON^{1,5,54}
1222. *Onypterygia exeuros* Whitehead and Ball, 1997
OAX^{1,4}
1223. *Onypterygia famini* Solier, 1835
CHIS^{1,2,5}, GRO², HGO^{1,2}, JAL², MOR², NAY², OAX^{1,2}, PUE^{1,5}, SLP^{1,2}, SON^{2,54}, VER^{1,3,4,52}
1224. *Onypterygia fulgens* Dejean, 1831; Figure 2E
CDMX^{1,5,7}, CHIH^{1,4}, CHIS^{1,2,3,4,5,6,7}, COL^{1,2,3,5}, DGO¹, GRO^{1,2,3,4,5,7}, GTO³, HGO^{1,2,3,4,5}, JAL^{1,2,3,4,5}, MEX^{1,4,5}, MICH^{2,3,4,7}, MOR^{1,2,3,4,5,6,7}, NAY^{1,2,3,4,5,7}, OAX^{1,2,4,5,7}, PUE^{1,4,7}, QRO³, SIN^{1,2,5,7}, SLP^{1,2}, SON^{1,5,54}, VER^{1,2,4,5,6,7}
1225. *Onypterygia hoepfneri* Dejean, 1831
CDMX⁵, CHIS^{1,2,5}, COL¹, GRO^{1,2,5}, JAL^{1,2,5}, MEX^{1,2}, MICH^{1,2}, MOR^{1,2}, NAY¹, OAX^{1,2}, SIN¹, VER⁵

1226. *Onypterygia iris* Chaudoir, 1863
CHIS^{1,5}, GTO¹, VER^{1,2,3,4,5,7}
1227. *Onypterygia kathleenae* Whitehead and Ball, 1997
CHIS^{1,5}, OAX^{1,2}
1228. *Onypterygia longispinis* Bates, 1882
CHIS^{1,2,5}
- 1229[#]. *Onypterygia pacifica* Whitehead and Ball, 1997
OAX^{1,59}; (type locality: “3.2 km north of San José Pacífico, Oaxaca, México”, Whitehead and Ball 1997: 320)
1230. *Onypterygia pallidipes* Chaudoir, 1878
MOR^{1,2}, PUE^{1,2}
- 1231[#]. *Onypterygia perissostigma* Whitehead and Ball, 1997
OAX^{1,2}
- 1232[#]. *Onypterygia pseudangustata* Whitehead and Ball, 1997
PUE^{1,2}
1233. *Onypterygia pusilla* Chaudoir, 1878
CDMX¹, CHIS^{1,2}, HGO^{1,2}, OAX^{1,2,3}, VER^{1,2,4,5}
1234. *Onypterygia rawlini* Whitehead and Ball, 1997
COL¹, NAY^{1,2}
1235. *Onypterygia rubida* Bates, 1884
COL¹, MICH^{1,2,18}
1236. *Onypterygia sallei* Chaudoir, 1863
CHIH⁶, SON⁶, VER^{1,2,52}
1237. *Onypterygia shpeleyi* Whitehead and Ball, 1997
GRO^{1,2,3,4,5}
1238. *Onypterygia stenapteryx* Whitehead and Ball, 1997
CDMX⁵, MEX¹, MICH^{1,2}
- 1239[#]. *Onypterygia sriblingi* Whitehead and Ball, 1997
OAX^{1,59}; (type locality: “Vista Hermosa, Oaxaca, 4.8 km south of Valle Nacional, on Highway 175 (ca. 17°45'N, 96°21'W”, Whitehead and Ball 1997: 348)
1240. *Onypterygia tricolor* Dejean, 1831
CDMX^{5,7}, CHIH^{1,2}, CHIS^{1,2,4,5}, COL^{1,2,3}, GRO^{1,2,3,4,5,7}, HGO^{1,2,3,5,24}, JAL^{1,2,3,4,5,6,7}, MEX^{1,2,3,4,5}, MICH^{1,2,3,4,6,7}, MOR^{1,2,3,4,5,7}, NAY^{2,3,4,5}, OAX^{1,2,4,5}, PUE^{1,2,3,4}, SIN^{1,2,3,5}, SLP^{1,2,3,7}, SON^{1,2,3,54}, TAMPS^{1,3}, VER^{1,2,3,4,5,7,52}, ZAC³
- 1241[#]. *Onypterygia wappesi* Whitehead and Ball, 1997
GRO^{1,5}
1242. *Platynus aeneicauda* (Bates, 1891)
DGO², GRO^{1,19}, MEX^{1,2}, MOR^{1,2}, VER⁵²
- 1243*. *Platynus agilis* LeConte, 1863
“California” Erwin et al. 1977: 4.29; *Platynus agilis* (LeConte 1863: 6) is a replacement name for *Platynus fragilis* LeConte 1854b: 41 [nec Mannerheim 1853]; (type locality of *P. fragilis*: “found in the mountains near Santa Isabel, in the southern part of California”, LeConte 1854b: 41).
- 1244[#]. *Platynus arboreus* (Mateu, 1979)
JAL^{1,59}; (type locality: “12 km N Ciudad Guzman, Jalisco, México”, Mateu 1979: 183)
1245. *Platynus ballorum* Liebherr, 1992
PUE^{1,2}, VER³
- 1246[#]. *Platynus baroni* (Casey, 1920)
GRO^{1,59}; (type locality: “México (Guerrero)”, Casey 1920: 25)
1247. *Platynus brunneomarginatus* (von Mannerheim, 1843)
BCN^{1,3,5,6,26}, BCS⁵, SON^{2,54}
1248. *Platynus cavatus* (Bates, 1882)
CDMX⁵, GRO⁵, OAX^{1,16}, SLP², VER^{2,52}
1249. *Platynus cohni* Liebherr and Will, 1996
DGO³, SON^{3,6}
1250. *Platynus conicicollis* (Chaudoir, 1878)
OAX^{1,2,3}, VER⁵²
- 1251[#]. *Platynus consularis* (Casey, 1920)
GRO^{1,59}; (type locality: “México (Guerrero)”, Casey 1920: 33)
1252. *Platynus convexulus* (Casey, 1920)
GRO², OAX²; VER⁵⁰
- 1253[#]. *Platynus dilatipes* Liebherr, 1989
CHIS^{1,59}; (type locality: “24.1 km SW El Bosque, Chiapas”, Liebherr 1989: 204)
- 1254[#]. *Platynus districtus* (Casey, 1920)
MOR⁵⁹; (type locality: “México, (Tres Marias, Morelos)”, Casey 1920: 24)
1255. *Platynus franiai* Liebherr, 1992
GRO^{1,2,3}, OAX^{1,2}
1256. *Platynus hamatus* Liebherr, 1989
CHIS^{1,2}, OAX^{1,2}
1257. *Platynus inops* (Chaudoir, 1878)
CHIS^{1,2,3,5}, HGO^{2,24}, OAX^{1,2,52}
1258. *Platynus logicus* (Casey, 1920)
CHIS², JAL², MEX^{1,2}, MOR¹, OAX^{2,3}, PUE², TLAX²
1259. *Platynus lyratus* (Chaudoir, 1878); Figure 2F
BCS^{5,26}, CDMX⁵, CHIH^{2,5}, CHIS², DGO^{2,5}, GRO², HGO^{2,24}, JAL⁵, MEX^{2,3,5}, MICH^{2,4}, MOR^{2,5}, NAY^{2,5}, OAX^{1,2}, PUE^{2,4,5}, QRO^{1,2,5}, SLP², SIN^{1,2}, SON^{2,3,54}, VER², ZAC²
1260. *Platynus montezumae* (Bates, 1878)
CDMX⁵⁹, GTO⁴, MEX¹, VER^{3,4}
1261. *Platynus pygmaeus* Liebherr, 1992
CHIS^{1,2}, OAX^{1,2,3}
1262. *Platynus robustus* (Chaudoir, 1878)
CHIS^{1,2,3}, QRO², OAX²
1263. *Platynus rubrofemoratus* Liebherr, 1989
CHIS^{1,2,3}
1264. *Platynus stricticollis* (Bates, 1878)
CHIS^{3,7}, GRO², SIN⁷, SLP^{2,3}
- 1265*. *Platynus tenuicollis* LeConte, 1848
VER¹; (type locality: “Sanctae Marie, (Sault de Ste. Marie) [Ontario]”, LeConte 1848: 222); “Canada and United States” (Bousquet 2012: 1248)
1266. *Platynus tolucensis* (Straneo, 1957)
MEX^{1,2,3,4,5}
- 1267*. *Platynus tropicum* (Motschulsky, 1865)
OAX¹, VER¹; (type locality: “Nicaragua”, Motschulsky, 1865:325)
1268. *Platynus umbripennis* (Casey, 1920)
GRO^{1,2,4}, SIN²
- 1269[#]. *Rhadine araiyai* (Bolívar y Peltain, 1944)
NL^{1,13,53}
1270. *Rhadine bolivari* Barr, 1982
COAH^{13,53}, COL¹

- 1271[#]. *Rhadine chipinque* Barr, 1982
NL^{1,13}
- 1272[#]. *Rhadine elliotti* Barr, 1982
NL^{1,13}
1273. *Rhadine euprepes* Bates, 1882
CHIH¹, DGO^{1,2,4,5,16,13}, MEX¹, OAX¹
1274. *Rhadine hendrichsi* Barr, 1982
SLP¹³, TAMPS¹
1275. *Rhadine leptodes* Bates, 1982; Figure 2G
CHIH⁵, DGO^{1,2,4,5,13}
1276. *Rhadine longipes* Casey, 1913
JAL³
1277. *Rhadine medellini* Bolivar y Pieltain and Hendrichs, 1964
SLP^{1,4,13}
1278. *Rhadine pelaezi* Bolivar y Pieltain and Hendrichs, 1964
CHIH¹, NL^{4,53}
1279. *Rhadine perlevis* Casey, 1913
CHIH^{1,2,13,53}
- 1280[#]. *Rhadine reddelli* Barr, 1982
TAMPS¹³
1281. *Rhadine rotgeri* Bolivar y Pieltain and Hendrichs, 1964
COL¹, DGO⁵³, COAH^{4,13}
1282. *Sericoda bembidioides* Kirby, 1837
CDMX^{1,44}, CHIS^{1,44}, GTO^{1,44}, HGO^{1,24,44}, MEX⁴⁴, MICH^{1,44}, MOR^{1,44}, OAX^{1,44}, PUE^{1,44}, TLAX^{1,44}, VER^{1,3,44}
1283. *Sericoda bogemannii* (Gyllenhal, 1813)
CHIS⁴⁴, SLP³
1284. *Tanystoma diabolicum* Liebherr, 1989
BCN^{1,43}, BCS⁴³
1285. *Tanystoma maculicolle* (Dejean, 1828)
BCN^{1,3,43}, TAB⁴

34. Perigonini

1286. *Diploharpus mexicanus* (Chevrolat, 1841)
OAX¹⁶, VER^{2,3,4,16}
1287. *Diploharpus perpolitus* Bates, 1882
CHIS², OAX², VER²
1288. *Perigona cordicollis* Bates, 1882
CHIS², JAL²
1289. *Perigona laevigata* (Bates, 1872)
CHIS², MOR², NL², OAX², PUE², SLP², VER^{2,3}
1290. *Perigona nigriceps* (Dejean, 1831)
TAMPS³

35. Lachnophorini

1291. *Anchonoderus apicalis* Reiche, 1843
BCS^{3,5}
1292. *Anchonoderus binotatus* Reiche, 1843
VER³
1293. *Anchonoderus fulvipennis* Bates, 1891
SIN¹⁹, TAMPS³
1294. *Anchonoderus horni* Csiki, 1931
BCN^{1,2}, BCS⁵, NAY⁵, OAX⁵, SIN⁵, SON^{5,54}
1295. *Anchonoderus subaeneus* Reiche, 1843
VER⁴

1296. *Anchonoderus subtilis* Bates, 1871
NL⁴
1297. *Ega laetula* LeConte, 1851
GRO⁴, JAL⁶, MOR⁴, SON^{4,5,54}
1298. *Ega sallei* Chevrolat, 1839
CAMP³, CHIS⁷, NL^{4,7}, QROO², SLP⁷, SON³, TAMPS^{2,3}, VER²
1299. *Euphorticus pubescens* (Dejean, 1831)
CHIS³
1300. *Lachnophorus cuprellus* Bates, 1891
DGO¹⁹, SIN¹⁹
1301. *Lachnophorus elegantulus* von Mannerheim, 1843
CHIS⁷, DGO⁷, QRO³, SIN⁷, SON^{2,3,6,54}, TAB⁴
1302. *Lachnophorus pictipennis* Bates, 1871
CHIS⁷, VER⁴

36. Pentagoniciini

1303. *Pentagonica bicolor* (LeConte, 1863)
CHIS^{2,3,7}, COL², PUE², VER^{2,3,5}
1304. *Pentagonica bifasciata* Chaudoir, 1877
CHIS^{2,3}, OAX²
1305. *Pentagonica felix* Bell, 1987
DGO², SON^{2,54}, TAMPS², VER³
1306. *Pentagonica flavipes* LeConte, 1853
CHIS^{2,3,7}, SLP², VER^{2,3}
1307. *Pentagonica maculicornis* Bates, 1883
CHIS³
1308. *Pentagonica obscura* Chaudoir, 1877
SLP²
1309. *Pentagonica picticornis* Bates, 1883
BCS³, COAH^{2,3,5}, NL^{2,5}, SLP², TAMPS^{2,3}
1310. *Pentagonica trivittata* Dejean, 1831
CHIS², QRO², VER²

37. Catapiesiini

1311. *Catapiesis mexicana* (Chaudoir, 1854)
CHIS^{1,2}, HGO²⁴, TAMPS^{1,2,3}
1312. *Catapiesis sulcipennis* Bates, 1882
VER^{1,16}
- 1313*. *Homalomorpha castanea* Brullé 1835 (**note:** GBIF has the year as 1837)
Type locality: “Cayenne”, Brullé 1835: 46; “México”¹; (Erwin et al. 1977: 4.65)

38. Odacanthini

- 1314[#]. *Colliuris bivittis* (Chaudoir, 1872)
Type locality: “México”, Chaudoir 1872b: 402; “México” (Blackwelder 1944: 65; Erwin et al. 1977: 4.56)
1315. *Colliuris ellipticeps* Liebke, 1930
CAMP², CHIS²
- 1316[#]. *Colliuris laeviceps* Liebke, 1930
Type locality: “México”, Liebke 1930: 683; “México” (Blackwelder 1944: 66; Erwin et al. 1977: 4.56)
1317. *Colliuris lengi* (Schaeffer, 1910)
SON⁵⁴; (type locality: “Nogales, Arizona”, Schaeffer 1910: 395)
1318. *Colliuris lioptera* (Bates, 1891)
CAMP², CHIH², CHIS^{2,3}, COL⁵, GRO⁵, JAL^{2,3,6}, NAY^{2,5}, SIN⁵, SLP², SON^{2,3,5,6,54}, VER^{2,5,19}

- 1319*. *Colliuris ludoviciana* (Sallé, 1849)
Type locality: “Nouvelle-Orléans (Louisiane)”, Sallé 1849: 298
1320. *Colliuris marginestriata* (Putzeys, 1845)
CHIS^{2,5}, JAL², MICH^{2,3,4}, MOR⁵, NAY², OAX², SIN⁵, SLP², SON^{2,5,4}, VER^{2,5}, ZAC⁵
1321. *Colliuris pensylvanica* (Linnaeus, 1758)
AGS^{2,5}, BCN^{3,5,6}, BCS^{2,3,6,26}, CDMX⁴, CHIH^{2,3,5}, COAH^{3,5,6}, COL², DGO², GTO^{2,4}, GRO², JAL^{2,3,5}, MEX^{2,3,5}, MICH², MOR⁵, NL², OAX^{2,3}, QRO³, SIN^{2,3,5}, SLP⁴, SON^{2,3,5,6,5,4}, TAMPS^{2,3}, VER⁵, YUC², ZAC^{3,5}
1322. *Colliuris pilatei* (Chaudoir, 1848)
BCS⁵, CHIS², COL², GRO^{2,3}, JAL^{2,3,5}, MEX⁵, MICH², NAY⁵, OAX^{2,3,5}, PUE^{3,5}, SLP², SIN^{2,5,7}, SON^{2,3,5,6,5,4}, TAMPS^{2,3}, VER^{2,3,4}
1323. *Colliuris subdistincta* (Chaudoir, 1863); Figure 2H
CHIS^{2,3,5}, COL², GRO^{2,3}, JAL^{2,3,5,7}, MEX⁵, MICH⁷, MOR², NAY^{2,3,5,6}, OAX^{2,5}, PUE^{2,3}, SIN^{2,5}, SLP^{2,3,7}, SON^{3,6}, TAB², TAMPS³, VER^{2,3,5}
1324. *Colliuris sulcicollis* (Bates, 1891)
DGO¹⁹
1325. *Colliuris tetrastigma* (Chaudoir, 1863)
NAY⁵, QROO³, VER⁵
1326. *Colliuris tristigma* (Bates, 1883)
CHIS^{2,3}, OAX^{2,5}, TAB², VER²
1327. *Colliuris tubulifera* Bates, 1878
VER²
- 1328#. *Colliuris yucantana* Liebke, 1930
YUC⁵⁹; (type locality: “Temex, N. Yucatan [México]”, Liebke 1930: 700)
- 39. Ctenodactylini**
1329. *Leptotrachelus dorsalis* (Fabricius, 1801)
CHIS², NAY^{3,6}
1330. *Leptotrachelus mexicanus mexicanus* (Chaudoir, 1852)
VER^{3,4}
- 1331*. *Leptotrachelus mexicanus puncticollis* Bates, 1878
Type locality: “Nicaragua”, Bates 1878: 600; “Guatemala” (Blackwelder 1944: 68)
- 1332#. *Leptotrachelus mexicanus tepicus* Liebke, 1938
NAY⁵⁹; (type locality: “El Cora, Tepic [Nayarit, México]”, Liebke 1938a: 117)
- 1333*. *Leptotrachelus testaceus* Dejean, 1831
Type locality: “Nouvelle Grenada [= Colombia and Panama]”, Dejean 1831: 287; “México”¹, (Blackwelder 1944: 68; Erwin et al. 1977: 4.56)
- 40. Cyclosomini**
- 1334#. *Anaulacus maclevei* Ball and Shpeley, 2002
CHIH¹, JAL², SON^{1,2,3,5,4}; (type locality: “17 km. south-west of Moctezuma, State of Sonora, México”, Ball and Shpeley 2002: 321)
1335. *Anaulacus piceolus* (Chaudoir, 1876)
CHIS^{1,2}, OAX^{1,2}, TAMPS¹, VER^{1,3}
1336. *Anaulacus sericatus* Chaudoir, 1846
CAMP^{1,2}, CHIS^{1,2,3}, OAX¹, QRO¹, VER^{1,2}, YUC¹
- 1337#. *Anaulacus whiteheadi* Ball and Shpeley, 2002
CHIS^{1,59}; (type locality: “San Quintin, State of Chiapas, México”, Ball and Shpeley 2002: 309)
1338. *Tetragonoderus fasciatus* (Haldeman, 1843)
BCN^{1,3}, BCS^{3,6,26}, COL², JAL², SIN², SON^{2,5,4}, TAMPS², VER³
- 1339#. *Tetragonoderus intermixtus* Bates, 1883
PUE¹⁷; (type locality: “México, Tehuacán, [state of Puebla]”, Bates 1883: 172)
1340. *Tetragonoderus intersectus* (Germar, 1823)
BCS^{2,5,26}, CHIS², COAH², COL², GRO², JAL^{2,3,5,6}, MOR², NAY⁵, NL², OAX⁵, PUE², SIN^{2,5}, SON^{2,3,5,6}, TAMPS², VER^{2,3}
- 1341*. *Tetragonoderus latipennis* LeConte, 1874
Type locality: “Texas”, LeConte 1874: 44; “United States and México” (Bousquet, 2012: 1275)
1342. *Tetragonoderus mexicanus* (Chaudoir, 1876); Figure 2I
CHIS^{2,7}, COL², GRO², JAL², MOR², OAX², PUE^{2,4}, SLP², SON^{2,5,4}, TAMPS², VER^{2,3}
1343. *Tetragonoderus pallidus* G. Horn, 1869
BCN^{2,5}, BCS^{2,3,5,6}, SON^{3,6,5,4}
- 1344#. *Tetragonoderus poecilus* Bates, 1883
OAX⁵⁹; (type locality: “México, Oaxaca; Guatemala, San Geronimo”, Bates, 1883: 172, not restricted)
Note: *Tetragonoderus poecilus* Bates, species number 298 in Shpeley & Van Devender, 2022: 115, was recorded in error; the correct Sonoran species is *Tetragonoderus intersectus* Germar, and all data recorded as well as the map applies to *T. intersectus* Germar (see above).
1345. *Tetragonoderus simplex* Bates, 1883
CHIS^{2,5,7}, OAX²
1346. *Tetragonoderus sinuosus* Chaudoir, 1876
CHIS², COL², OAX², SLP^{2,7}, VER^{2,3}
- 41. Lebiini**
1347. *Agra cyanippe* Bates, 1891
MOR¹⁹
1348. *Agra dimidiata* Chevrolat, 1856
CHIS¹
1349. *Agra eurytelma* Bates, 1883
VER¹⁷
1350. *Agra fada* Chevrolat, 1856
VER¹⁷
- 1351#. *Agra foveella* Liebke, 1937
Type locality: “México”, Liebke 1937: 158
- 1352*. *Agra guatemalana* Csiki, 1932
“Guatemala” (Csiki 1932: 1513; Blackwelder 1944: 64; Erwin et al. 1977: 4.66); *Agra guatemalana* Csiki 1932: 1513 is a replacement name for *A. regularis*, type locality: “San Geronimo, Guatemala”, Bates 1883: 248
- 1353#. *Agra hovorei* Erwin, 2000
VER⁵⁹; (type locality: “México, Veracruz, Estacion Biologica Los Tutxlas”, Erwin 2000: 15)
- 1354#. *Agra ictina* Bates, 1883
VER¹⁷; (type locality: “México, Cordova, [state of Veracruz]”, Bates 1883: 252)
- 1355#. *Agra kayae* Erwin, 1984
CHIS⁵⁹; (type locality: “México, Chiapas, Arroyo Santa Maria, east slope of Sierra de Colmena”, Erwin

- 1984b: 39)
- 1356[#].** *Agra lutea* Liebke, 1938
Type locality: “México”, Liebke 1938a: 61
- 1357[#].** *Agra mexicana* Buquet, 1835
CHIS⁴
- 1358[#].** *Agra negrei* Straneo, 1965
VER⁵⁹; (type locality: “México: Environs of San André, V.C. [Veracruz]”, Straneo 1965: 476)
- 1359[#].** *Agra nigripes* Chaudoir, 1847
Type locality: “México”, Chaudoir 1847: 98
- 1360[#].** *Agra oblongopunctata* Chevrolat, 1835
VER^{1,59}; (type locality: “Veracruz”, Chevrolat 1835: no. 183)
- 1361[#].** *Agra obscuripes* Chaudoir, 1854
Type locality: “México”, Chaudoir 1854: 313
- 1362[#].** *Agra pacifica* Bates, 1891
GRO¹⁹
- 1363[#].** *Agra paloma* Erwin, 1984
OAX⁵⁹; (type locality: “México, Oaxaca, Palomeres”, Erwin 1984b: 39)
- 1364[#].** *Agra perkinsorum* Erwin, 1986
NAY⁵⁹; (type locality: “México, Nayarit, 17.6 km. southwest of Compostela”, Erwin 1986: 298)
- 1365[#].** *Agra rosettiae* Straneo, 1960
GRO⁵⁹; (type locality: “Messico [México], Acapulco [Guerrero]”, Straneo 1960: 428)
- 1366[#].** *Agra rotundangula* Straneo 1955
MEX⁵⁹; (type locality: “Messico [México], Bejucos, Temascaltepec, [Mexico (state)]”, Straneo 1955: 21)
- 1367.** *Agra rufoaena* Chevrolat, 1835
GRO⁴, SIN³, VER⁵⁹
- 1368[#].** *Agra rugosostriata* Chaudoir, 1854
CAMP⁵⁹; (type locality: “Campeche, [México]”, Chaudoir 1854: 318)
- 1369.** *Agra truquii* Chaudoir, 1866
SON⁵⁴; (type locality: “México”, Chaudoir 1866b: 97)
- 1370[#].** *Agra tuxtlas* Erwin, 2000
VER⁵⁹; (type locality: “México, Veracruz, Estacion Biologica Los Tuxtlas, Erwin, 2000: 15)
- 1371[#].** *Agra vate* Erwin, 1986
VER⁵⁹; (type locality: “México, Veracruz, Los Tuxtlas, La Playa Escondida”, 1986: 302)
- 1372[#].** *Agra virgata* Chevrolat, 1856
Type locality: “México”, Chevrolat, 1856: 352; “México” (Blackwelder, 1944: 65; Erwin et al., 1977: 4.66)
- 1373.** *Agra zapotal* Erwin, 2000
CHIS^{1,31}, VER^{1,31}; (type locality: “Guatemala, Alta Verapaz, San Cristobal Verapaz, Quixal”, Erwin 2000: 15)
- 1374.** *Apenes amplicollis* Bates, 1891
DGO¹⁹, MICH⁴, NAY^{2,5,19}, OAX², SIN⁵
- 1375[#].** *Apenes brevivittis* Chaudoir, 1875
YUC⁵⁹; (type locality: “Yucatan, [México]”, Chaudoir 1875: 26)
- 1376.** *Apenes circumcincta* Chaudoir, 1875
TAB²; type locality: “México”, Chaudoir 1875: 30; “México” (Blackwelder 1944: 62; Erwin et al. 1977: 4.58)
- 1377.** *Apenes hamigera* (Chaudoir, 1875)
CHIS^{2,3,7}, VER^{2,4}
- 1378.** *Apenes hilariola* Bates, 1891
COL¹⁹, PUE³, MOR¹⁹, SON³
- 1379*.** *Apenes limbata* G. Horn, 1895
BCN¹; (type locality: “San Luis Obispo, [California]”, G. Horn 1895: 232)
- 1380.** *Apenes lucidula* Dejean, 1831
CHIH², HGO⁵, JAL², MOR², NAY⁵, NL², SIN⁵, SLP², SON^{2,5,53}, TAMPS², VER^{2,5}
- 1381.** *Apenes lunulata* Chaudoir, 1875
CAMP^{2,4}, CHIS², MOR², NAY², NL², SIN^{2,5}, SLP², TAMPS², VER^{2,3,5}
- 1382.** *Apenes marginalis* Dejean, 1831
DGO⁵, MOR^{2,5}, OAX², SIN^{2,5}, SON^{2,53}
- 1383*.** *Apenes morio* (Dejean, 1825)
Type locality: “l’Amerique septentrionale”, Dejean, 1825: 219; “This species is known only from the island of Hispaniola” (Ball and Shpeley 2009: 119)
- 1384.** *Apenes nebulosa* LeConte, 1866
BCN^{1,3}, BCS²⁶, COL², DGO⁵, NAY², NL², OAX², SIN^{2,5}, SON^{2,3,5,53}, TAMPS², ZAC²
- 1385.** *Apenes obscura* Chaudoir, 1875
CHIS², COL², MOR², NAY², QRO²
- 1386.** *Apenes pallidipes* (Chevrolat, 1835)
SLP², VER⁵⁹, YUC²
- 1387*.** *Apenes parallela* (Dejean, 1825)
Type locality: “Cuba”, Dejean, 1825: 218; “This species is known only from the Bahamas, and the Greater Antillean islands of Cuba and Jamaica” (Ball and Shpeley 2009:116)
- 1388.** *Apenes parvula* (Chaudoir, 1875)
TAB², TAMPS², VER^{2,3,5}, YUC⁵⁹
- 1389.** *Apenes postica* Dejean, 1831
CHIS², JAL⁵, SIN⁵, VER⁴
- 1390.** *Apenes quadripennis* (Chaudoir, 1875)
CHIS⁴, VER²
- 1391.** *Apenes sallei* (Chaudoir, 1875)
TAMPS³
- 1392.** *Apenes seriata* (Motschulsky, 1864)
CAMP⁴, CHIS², YUC²
- 1393.** *Apenes sinuata* (Say, 1823)
COAH², TAMPS^{2,4}
- 1394.** *Apenes variegata* Dejean, 1825
CAMP², CHIS^{2,4,5}, JAL², MOR⁵, OAX², SLP², SON², TAB², VER^{2,4,5}, YUC²
- 1395.** *Apristus laticollis* LeConte, 1851
BCS^{21,26}
- 1396.** *Apristus mexicanus* Bates, 1883
CHIS³, JAL^{3,4}, OAX¹⁷, VER^{3,4}
- 1397.** *Apristus pugetanus* Casey, 1920
BCN⁵, BCS⁵, SON⁵⁴
- 1398.** *Apristus subcyaneus* G. Horn, 1894
BCS^{3,5}
- 1399[#].** *Aspasiola rutilans* Chaudoir, 1877
Type locality: “México”, Chaudoir 1877: 209; “México” (Blackwelder 1944: 57; Erwin et al. 1977: 4.63)
- 1400.** *Axinopalpus biplagiatus* (Dejean, 1825)
DGO⁵, SON^{5,54}

1401. *Axinopalpus fusciceps* LeConte, 1851
BCN², CHIS², COAH³, PUE², QRO², SON⁵⁴, TAMPS⁶
1402. *Axinopalpus jucundus* Bates, 1883
VER^{3,7}
1403. *Axinopalpus mexicanus* Bates, 1883
OAX¹⁷, VER⁵⁹
1404. *Calleida amethystina* Fabricius, 1787
CAMP², CHIS², OAX²
1405. *Calleida aurata* Motschulsky, 1864
GRO⁴
1406. *Calleida aurescens* Bates, 1883
CHIS², GRO⁴, MICH², OAX², VER⁴
1407. *Calleida basalis* Putzeys, 1845
CAMP², CHIS², OAX², SLP^{2,7}, TAB², TAMPS², VER^{2,4}
1408. *Calleida bella* Chaudoir, 1872
VER⁴
1409. *Calleida brunnea* Dejean, 1831
GRO², JAL², OAX², SON^{2,54}
1410. *Calleida bryanti* Liebke, 1939
Type locality: “Mexiko und Bahia”, Liebke, 1939: 99;
“México” (Blackwelder, 1944: 59; Erwin et al., 1977: 4.59)
1411. *Calleida chlorotaenia* Bates, 1891
COL², GRO¹⁹, SIN², SLP², SON²
1412. *Calleida circumcincta* Bates, 1883
CHIS², VER^{2,4,17}
1413. *Calleida cordicollis* Putzeys, 1845
BCN^{3,5,6}, BCS^{2,3,5}, CAMP², CHIS^{2,4,5}, COL², GRO^{2,4},
JAL^{2,3,5,6}, MEX², MICH², MOR^{2,3,5}, NAY^{2,5}, NL^{2,3},
OAX^{2,5}, PUE^{3,5}, SIN^{2,3,5}, SLP^{2,3}, SON^{2,3,5,6,54}, TAB²,
TAMPS^{2,5}, VER^{2,3,4,5}, YUC^{2,5}
1414. *Calleida cyanippe* Bates, 1883
GRO², HGO³, MICH², MOR², OAX², SON^{2,54}, VER^{2,3,17}
- 1415[#]. *Calleida dives* Chaudoir, 1852
Type locality: “México and Guatemala”, not restricted,
Chaudoir, 1852: 50; “México” (Blackwelder 1944: 59;
Erwin et al. 1977: 4.60)
1416. *Calleida fimbriata* Bates, 1883
MICH², VER¹⁷
- 1417[#]. *Calleida flava* Chevrolat, 1833
VER⁵⁹; (type locality: “Orizaba [= Orizaba, Veracruz,
México]”, Chevrolat 1833: “Errata et Addenda”, whe-
re he writes (English translation): “The name *Calleida*
brunnea will have to be converted to that of *Calleida*
flava; delete Dejean’s quote.”)
1418. *Calleida flohri* Bates, 1883
VER^{4,17}
1419. *Calleida georgii* Casale, 2008
COL^{2,22}, GRO²², JAL²², NAY^{2,22}, OAX²²
1420. *Calleida hoegei* Bates, 1883
CAMP², GRO², MICH², VER¹⁷
1421. *Calleida ignobilis* Bates, 1883
OAX¹⁷
1422. *Calleida jansoni* Bates, 1878
CHIS², GRO², MOR², PUE², ZAC³
1423. *Calleida kayae* Casale, 2008
CHIS^{2,22}, OAX^{2,22}
- 1424*. *Calleida levistriata* Chaudoir, 1872
Type locality: “México and Bolivia”, not restricted,
Chaudoir 1872a: 144; Chaudoir, English translation,
states “M. Chevrolat gave me a male, selling from
México, and M. Guerin-Meneville a female selling
from Bolivia; I believe that it is this last country which
is the real part of this insect”); “México”¹; “México”
(Blackwelder 1944: 59; Erwin et al. 1977: 4.60).
1425. *Calleida misella* Chaudoir, 1872
OAX¹⁷, VER¹⁷
1426. *Calleida planulata* LeConte, 1858
CAMP², OAX², VER^{2,3,50}, YUC²
1427. *Calleida platynoides* G. Horn, 1882
BCN⁵, BCS^{2,3,5}, CHIH⁵, MICH^{2,5}, NL², SON^{5,54}
1428. *Calleida punctata* LeConte, 1846
SON³, TAMPS³
1429. *Calleida punctulata* Chaudoir, 1848
BCS^{2,3,5}, CAMP², CHIS², GRO², MICH², MOR²,
NAY^{2,5}, OAX^{2,5}, PUE², QROO², SIN^{2,5}, SON^{2,5,54},
TAMPS^{2,5}, VER², YUC²
1430. *Calleida pupuripennis* Chaudoir, 1872
TAMPS², VER³
1431. *Calleida regina* Bates, 1883
CHIS²
1432. *Calleida rustica* Bates, 1883
VER¹⁷
1433. *Calleida semifacta* Bates, 1883
COL³, HGO³, VER¹⁷
1434. *Calleida sericinitens* Bates, 1883
CAMP², CHIS², COL², MICH², MOR², OAX², VER²
- 1435[#]. *Calleida skwarrae* Liebke, 1932
VER; (type locality: “Mirador, [state of Veracruz],
Mexiko”, Liebke 1932: 193)
1436. *Calleida sumptuosa* Bates, 1883
VER^{3,17,52}
1437. *Calleida tropicalis* Bates, 1883
CHIS², OAX², VER^{2,17}
1438. *Calleida truncata* Chevrolat, 1835
CAMP², CHIS², QROO², SLP², VER^{2,50}
1439. *Catascopus brasiliensis* Dejean, 1831
CHIS^{1,2,5}, VER^{1,2}
1440. *Catascopus obscuriviridis* Chevrolat, 1835
CHIS^{2,7}, JAL², MOR², NAY⁵, OAX², PUE^{2,5}, TAB²,
VER^{1,4,5}, YUC^{2,7}
1441. *Catascopus rufifemoraus* Chaudoir, 1837
MOR⁴; **note**: most likely mis-identified or mis-la-
belled; type locality: “Cap de Bonne Esperance”
[South Africa], Chaudoir 1837: 9
1442. *Catascopus validus* Chaudoir, 1854
CHIS¹, OAX¹, VER^{1,2}
Note: Erwin’s superscript ¹ for *Coptodera* species re-
fers to Shpeley and Ball 1993.
1443. *Coptodera acutipennis* (Buquet, 1834)
CHIS^{1,2}, OAX^{1,2}, VER¹
1444. *Coptodera aerata* Dejean, 1825
COL³
1445. *Coptodera aurata* Chevrolat, 1835
CHIS^{1,2}, JAL⁵, MOR⁴, OAX^{1,2,5}, PUE^{1,2}, VER^{1,2,3,4}
1446. *Coptodera brunnea* Shpeley and Ball, 1993
CHIS², SON^{2,54}

1447. *Coptodera cupreotincta* Bates, 1869
CHIS^{1,2}, OAX^{1,2}, VER^{1,2}
1448. *Coptodera elongata* Putzeys, 1845; Figure 2J
CHIS^{1,5}, OAX^{1,2}, PUE¹, VER^{1,2,5}
1449. *Coptodera festiva* Dejean, 1825
CAMP^{1,2}, CHIS^{1,2}, NAY^{1,2}, OAX^{1,2}, SLP^{1,7}, VER^{1,2,3}
1450. *Coptodera foveolata* Shpeley and Ball, 1993
VER²
1451. *Coptodera lineata* (Bates, 1883)
CHIS^{1,2}, JAL^{1,2}, OAX³, QRO¹, SLP^{1,5}, TAB¹, VER^{1,2,5,17}
- 1452#. *Coptodera nigroviridis* Shpeley and Ball, 1993
CHIS¹, HGO¹; (type locality: “Parque Montebello, state of Chiapas, México”, Shpeley and Ball, 1993: 84)
1453. *Coptodera nitidula* (Buquet, 1834)
CHIS^{1,2,3,7}, COL^{1,5}, MOR^{1,2}, NAY^{1,5}, OAX^{1,2}, PUE¹, QRO¹, SIN¹, SLP⁷, SON^{1,2,54}, VER¹
1454. *Coptodera picea* Dejean, 1826
CHIS^{1,2}, QRO¹, QROO^{2,3}, OAX¹, SLP^{1,5}, TAB¹, VER^{1,2}, YUC¹
1455. *Coptodera relucens* Bates, 1869
VER⁴; **note:** most likely mis-identified or mis-labelled; type locality: “Ega” [Amazonas, Brazil], Bates 1869: 73; known only from Brazil (Shpeley and Ball 1993: 77)
1456. *Coptodera sallei* Shpeley and Ball, 1993
QRO¹, SLP⁷, TAMPS¹, VER¹
1457. *Coptodera transversa* (Reiche, 1843)
CHIS¹, OAX¹, QRO¹, SLP^{1,7}, VER^{1,2,4}
1458. *Coptodera viridis* Shpeley and Ball, 1993
COL^{1,5}, JAL^{1,2,3}, NAY⁵ (Tres Marias Islands)
1459. *Coptodera xanthopleura* Bates, 1891
CHIS^{1,2,5}, GRO¹⁹, JAL^{1,2}, NAY^{1,2,5}, OAX^{1,2,19}, SIN¹
1460. *Cylindronotum aeneum* Putzeys, 1845
CHIS²
1461. *Cymindis apache* Hilchie and Ball, 2024
= *Cymindis* sp. son-3 (species #326, Shpeley & Van Devender, 2022: 122)
CHIH^{2,36}, SON^{2,36}
1462. *Cymindis arizonensis* Schaeffer, 1910
BCN⁵, SON^{2,54}
- 1463#. *Cymindis atrolucens* (Casey, 1913)
MOR⁵⁹; (type locality: “México (Tres Marias, Morelos)”, Casey 1913: 178)
1464. *Cymindis basipunctata* (Chaudoir, 1875)
CHIS², HGO^{1,2}, NL², OAX², QRO², SLP², VER^{1,2,7,52}
1465. *Cymindis chalcea* (Bates, 1883)
CHIS²
1466. *Cymindis chevrolati chevrolati* Dejean, 1836
CDMX^{2,4,5}, CHIH^{2,5}, COAH², DGO^{2,5}, GTO⁴, HGO^{2,5}, JAL^{2,5}, MEX^{2,5}, MICH^{2,4,5}, MOR^{2,5}, NAY⁵ (Tres Marias Islands), NL², PUE^{2,5}, QROO⁵, SLP², TLAX², VER², ZAC⁵
- 1467#. *Cymindis chevrolati amblygona* (Bates, 1878)
Type locality: “México”, Bates 1878: 606
- 1468#. *Cymindis chevrolati angulifera* (Bates, 1878)
Type locality: “México”, Bates 1878: 606
1469. *Cymindis crenatoverpa* Hilchie and Ball, 2024
GRO^{2,36}, JAL^{2,36}, MEX³⁶, MICH^{2,36}, NAY^{2,36}
- 1470#. *Cymindis cribrata* (Chaudoir, 1875)
VER⁵⁹; (type locality: “México (Etat de Vera Cruz)”, Chaudoir 1875: 5)
- 1471#. *Cymindis cuyuteca* Hilchie and Ball, 2024
JAL³⁶
1472. *Cymindis geminata* Hilchie and Ball, 2024
CHIS^{2,36}, GRO^{2,36}, MOR^{2,36}, OAX^{2,36}, PUE³⁶
- 1473#. *Cymindis huichilobos*, Hilchie and Ball
OAX^{2,36}
1474. *Cymindis interior* Lindroth, 1969
CHIH³
1475. *Cymindis laevior* Bates, 1891
HGO², JAL², MEX², MOR², OAX^{2,5,7}, PUE², VER²
1476. *Cymindis latiuscula* (Chaudoir, 1875)
CAMP³⁶, CHIS^{2,7,36}, GRO^{2,36}, JAL³⁶, OAX^{2,5,36}, PUE^{2,36}, SLP^{2,36}, TAMPS^{2,7,36}, VER^{2,5,36}, YUC^{2,36}
1477. *Cymindis platicollis* (Say, 1823)
NL²
1478. *Cymindis punctifera punctifera* LeConte, 1884
CHIH^{2,5,36}, COAH^{2,36}, DGO⁵, MOR⁵, NL^{2,5,36}, OAX⁵, SIN⁵, SON^{2,5,36,54}, TAMPS³⁶
1479. *Cymindis punctifera toltec* Hilchie and Ball, 2024
COL^{2,36}, DGO^{2,36}, GRO^{2,36}, HGO^{2,36}, JAL^{2,21}, MEX³⁶, MICH^{2,36}, MOR^{2,36}, NAY^{2,36}, OAX^{2,36}, PUE^{2,36}, SIN^{2,36}, VER³⁶, ZAC³⁶
1480. *Cymindis punctigera punctigera* LeConte, 1851
BCN^{2,5}, BCS⁵, COAH², DGO², JAL², MICH², NAY^{2,5}, NL^{2,5}, SON^{2,5,54}
1481. *Cymindis punctigera sulcipennis* Horn, 1881
BCN^{1,5,37}, BCS^{5,37}
1482. *Cymindis ruficornis* Bates, 1891
GRO^{2,5}, JAL⁵
1483. *Cymindis rugofrons* Hilchie and Ball, 2024
COL³⁶, JAL^{2,36}, MICH^{2,36}, NAY^{2,36}
- 1484#. *Cymindis semisulcata* G. Horn, 1881
BCS^{1,59}; (type locality: “Penins. Cal.”, Horn 1881: 40); (type locality: “La Paz, Baja California Sur, México”, designated by Hunting 2013: 48)
- 1485#. *Cymindis tonatiuh* Hilchie and Ball, 2024
OAX^{2,36}
1486. *Cymindis uniseriata* Bates, 1884
CHIH^{5,18}, DGO^{2,5}, GRO⁵, SON^{2,54}
1487. *Cymindis yaqui* Hilchie and Ball, 2024
= *Cymindis* sp. son-2 (species #325, Shpeley & Van Devender, 2022: 121)
SON^{2,3}
1488. *Cymindis zapotec* Hilchie and Ball, 2024
GRO^{8,36}, OAX³⁶
- 1489#. *Dromius chihuahuae* Casey, 1920
CHIH⁵⁹; (type locality: “México (Colonia Garcia, Sierra Madre Mts., Chihuahua)”, Casey 1920: 278)
1490. *Dromius flohri* Bates, 1883
CHIS², OAX², VER¹⁷
1491. *Dromius guatemalensis* Bates, 1883
OAX²
1492. *Dromius piceus* Dejean, 1831
BCN⁵, BCS⁵, CDMX⁵, COAH², DGO⁵, MICH², OAX², SON^{2,54}, TAMPS³, VER²

1493. *Eucheila boyeri* (Solier, 1835)
CAMP², CHIS², OAX²
1494. *Eucheila cordova* Ball and Shpeley, 1983
VER^{3,5}
1495. *Eucheila planipennis* (Bates, 1891)
CAMP², VER^{2,19,50}, YUC^{2,19}
Note: Erwin's superscript ¹ for *Euproctinus* species refers to Shpeley 1986.
1496. *Euproctinus abjectus* (Bates, 1883)
BCN¹, BCS^{2,5}, CHIS^{1,2,3}, COL^{1,2}, GRO^{1,2,3}, HGO^{1,5}, JAL^{1,2,3,5}, MEX², MICH^{1,2,3,5}, MOR^{1,2,5}, NAY¹, OAX^{1,2,3,5,17}, PUE^{1,5}, SIN^{1,2,3}, SLP¹, SON^{1,2,3,5,4}, TAMPS^{2,3}, VER^{1,2,3,5,17}
1497. *Euproctinus balli* Shpeley, 1986
CHIS^{1,2}, HGO¹, MICH^{1,2}, OAX^{1,2}, SON^{1,2,5,4}
1498. *Euproctinus ornatellus* Bates, 1883
CHIS¹, SLP^{1,2}, VER³, YUC¹
1499. *Euproctinus pallidus* Shpeley, 1986
VER^{1,2}
1500. *Euproctinus quadriplagiatus* (Reiche, 1842)
HGO^{1,2}, OAX^{1,2}, PUE¹, SLP¹, VER^{1,4}
1501. *Euproctinus sigillatus* (Bates, 1883)
CHIH^{2,7}, CHIS², GRO¹, JAL^{1,2}, MICH², MOR¹, NL¹, OAX^{1,2,17}, SIN^{1,2}, SON^{5,4}
1502. *Euproctinus subdeletus* (Bates, 1883)
CHIS^{1,2}, DGO¹, OAX¹, QRO¹, VER^{1,2,5,17}
1503. *Eurycoleus macularius* (Chevrolat, 1835)
CAMP^{1,2}, CHIS^{1,4,5}, TAB^{1,2}, VER^{1,2}
1504. *Eurycoleus octosignatus* Bates, 1883
OAX^{1,17}, SIN¹
1505. *Eurycoleus ornatus* Bates, 1883; Figure 2K
CAMP^{1,2}, CHIS^{1,2}, VER^{1,2,3,17}
1506. *Eurycoleus tredecimpunctatus* Chaudoir, 1869
GRO^{1,5}, OAX^{2,7}
1507. *Gallerucidia erotyloides* Bates, 1883
CHIS², NL², OAX^{2,17}, VER^{2,17}
- 1508*. *Hyboptera auxiliadora* Erwin, 2004
Type locality: "Texas, Hidalgo County, Mission, Bentzen State Park", Erwin 2004: 36
1509. *Infernophilus castaneus* (G. Horn, 1882)
BCS^{2,21,26}
1510. *Lebia abdita* Madge, 1967
BCN^{2,3}, BCS^{2,3}, SON^{2,3,6,5,4}
1511. *Lebia abdominalis* Chaudoir, 1843
CDMX⁵, CHIH⁵, CHIS⁵, MICH³, NAY⁵, OAX⁵, SIN⁵, SON², TAMPS³, VER^{3,5}
1512. *Lebia agnata* Chaudoir, 1870
HGO², JAL², MOR², OAX², SLP²
1513. *Lebia analis* Dejean, 1825
BCN^{3,6}, BCS^{2,3,5}, CAMP^{2,5}, CDMX⁵, CHIH^{2,3,6}, CHIS², COAH³, GRO^{2,5}, JAL², NAY^{2,5}, NL^{2,5}, OAX^{2,5}, SIN^{2,5}, SLP^{2,5}, SON^{2,3,5,6,5,4}, TAB², TAMPS², VER^{2,5}, YUC²
- 1514#. *Lebia anchorifera* (Chaudoir, 1871)
Type locality: "México", Chaudoir 1871: 55
- 1515#. *Lebia apicalis* Chevrolat, 1834
VER⁵⁹; type locality: "Orizaba [= Orizaba, Veracruz México]", Chevrolat 1834: no. 39
1516. *Lebia atriceps* LeConte, 1863
AGS³, CHIH⁵, COAH², DGO², GRO³, GTO², HGO^{3,5,7}, JAL^{2,3,5,6}, MICH², MOR^{2,4}, NAY⁵, NL³, PUE³, QRO³, SON^{2,3,5,6,5,4}, VER², ZAC^{2,3,6}
- 1517*. *Lebia atriventris* Say, 1823
No type locality/area by Say 1823: 13; "U.S. and Canada" (Bousquet, 2012: 1315); "México (Chaudoir, 1870: 142) is likely in error" (Bousquet, 2012: 1315)
1518. *Lebia azteca* (Reichardt, 1972)
CHIS²
1519. *Lebia balli* (Reichardt, 1972)
CHIH², CHIS^{2,7}, MICH², SON^{2,3,5,4}
- 1520#. *Lebia biannulata* Chaudoir, 1871
Type locality: "México", Chaudoir 1871:15
- 1521#. *Lebia bipunctata* Chevrolat, 1835
Type locality: "environs de México", Chevrolat 1835: no. 133
1522. *Lebia bilineata* Motschulsky, 1859
GTO⁴
1523. *Lebia bitaeniata* Chevrolat, 1834; Figure 2L
CHIS^{2,7}, COAH², GRO^{2,3}, JAL⁵, MOR^{2,5}, NAY⁵, OAX², SIN^{2,5}, SLP^{2,7}, TAB², TAMPS^{2,3,7}, VER^{2,3}, YUC³
1524. *Lebia bivittata* (Fabricius, 1798)
CHIH⁵, CHIS³, DGO^{2,3}, GTO², MEX⁵, NL², VER²
1525. *Lebia bivitticollis* Bates, 1883
CAMP^{2,3}, CHIS^{2,3,7}, OAX^{2,3}, VER^{2,3}, YUC^{2,3}
1526. *Lebia bumeliae* Schaeffer, 1910
OAX^{2,3}, TAMPS³
1527. *Lebia calliope* Bates, 1883
CHIS⁷, SLP⁷, TAMPS³, VER^{2,4,17}
1528. *Lebia calomicra* Bates, 1891
CHIS^{2,7}, DGO², GRO¹⁹, MOR², OAX², TAMPS²
1529. *Lebia centromaculata* Putzeys, 1846
CHIS², MOR², NAY², OAX², SON²
1530. *Lebia chalybe* Bates, 1883
AGS², CHIH², DGO², GRO², JAL², MICH², MOR², NAY², PUE^{2,17}, SIN^{2,5}, SON^{2,5,4}
1531. *Lebia charilla* Bates, 1883
VER¹⁷
1532. *Lebia charina* Bates, 1878
CHIS^{2,3}
- 1533#. *Lebia chloroptera* Chaudoir, 1835
Type locality: "Mexique", Chaudoir 1835: 437
1534. *Lebia chlorotica* Dejean, 1831
CHIS², GRO^{2,3}, MICH^{2,3}, MOR², OAX², VER^{2,7}
1535. *Lebia clio* Bates, 1883
CHIS^{2,3,7}, GRO², QRO³, SLP², SON², TAB², VER^{2,17}
1536. *Lebia cordelia* Bates, 1883
VER¹⁷
1537. *Lebia costulata* (Bates, 1883)
MOR², VER^{2,17}
1538. *Lebia croceicollis* Bates, 1883
CHIS⁴, VER¹⁷
1539. *Lebia cyane* (Bates, 1883)
CDMX¹⁷, CHIS⁷, GRO⁴, GTO¹⁷, MOR⁴, VER⁴
1540. *Lebia cymindoides* Bates, 1883
= *Lebia scapula* G. Horn 1885 **NEW SYNONYMY**; comparison of external morphology as well as armature of everted endophallus of male genitalia confirms that these two species are conspecific.
AGS⁵, CHIH^{2,5}, CHIS², DGO², GTO^{4,17}, HGO², JAL²,

- MOR^{2,5}, OAX^{2,5,17}, PUE¹⁷, SON^{2,3,6,54}, VER^{2,17}, ZAC²
 1541. *Lebia deceptrix* Madge, 1967
 SON^{2,3,6,54}
 1542. *Lebia dugesi* Bates, 1883
 GTO^{4,17}, MICH⁴, MOR⁴
 1543. *Lebia duillia* Bates, 1883
 CHIH², COL², GRO², JAL², MOR², SLP², SON^{2,54},
 VER^{2,17}
 1544. *Lebia eburata* (Bates, 1883)
 CAMP^{3,7}, VER^{2,3,7}
 1545*. *Lebia esurialis* Casey, 1920
 Type locality: “Brownsville, Texas”, Casey 1920: 257
 1546. *Lebia exarata* (Kirsch, 1873)
 CDMX⁴; **note**: most likely mis-identified or mis-labeled; type locality: none stated, “Peru [inferred from title]”, Kirsch 1873: 127
 1547. *Lebia flammea* Bates, 1883
 GTO¹⁷, PUE¹⁷
 1548#. *Lebia flohri* Bates, 1883
 Type locality: “México”, Bates 1883: 227; “México”¹⁷
 1549. *Lebia fuscata* Dejean, 1825
 CHIS³, MEX³, TLAX³
 1550. *Lebia goniessa* Bates, 1883
 HGO², MOR², QROO², VER²
 1551. *Lebia grammica* Perty, 1830
 VER³
 1552. *Lebia grandis* Hentz, 1830
 BCS^{3,5}, MOR⁵; “Baja California and U.S.A.” Blackwelder 1944: 54
 1553. *Lebia guatemalena* Bates, 1883
 CHIS^{2,7}, GRO², SIN², SLP⁷
 1554. *Lebia guttula* LeConte, 1851
 BCN⁵, BCS⁵, DGO⁵, SON^{5,54}
 1555. *Lebia hiliaris* (Chaudoir, 1871)
 TAMPS³, VER³
 1556. *Lebia histrionica* Bates, 1883
 AGS³, CHIH², CHIS^{2,7}, COL², DGO², GRO^{2,3}, GTO³,
 JAL^{3,6}, MEX⁵, MICH^{2,3}, NAY^{2,5}, NL², OAX⁵, QRO³,
 QROO³, SIN², SON^{2,3,54}, TAB^{2,5}, TAMPS², VER²,
 YUC²
 1557. *Lebia hoegei* (Bates, 1883)
 NAY⁴, VER^{4,17}
 1558. *Lebia humeroguttata* (Chaudoir, 1871)
 CHIS^{3,7}
 1559#. *Lebia incohaerens* Chaudoir, 1870
 YUC⁵⁹; (type locality: “Teapa (Yucatan), [México]”, Chaudoir 1870: 244)
 1560#. *Lebia incommoda* Chaudoir, 1870
 CAMP⁵⁹; (type locality: “Campeche, [México]”, Chaudoir 1870: 213)
 1561#. *Lebia intermedia* (Chaudoir, 1871)
 YUC⁵⁹; (type locality: “Yucatan, [México]”, Chaudoir, 1871: 55)
 1562#. *Lebia interrupta* Chaudoir, 1870 **NEW STATUS**
 Type locality: “México”, Chaudoir 1870: 211; “México” (Blackwelder 1944: 56; Erwin et al. 1977: 4.63)
 = *Lebia tschitscherini* Lutshnik, 1922 **NEW SYNONOMY**
 Dejean 1825: 261-262 in his description of *Lebia*

cruxminor Linnaeus writes (English translation): “It is found under stones and on trees and plants, in almost all of Europe. It is quite rare from around Paris. Mr. Gebler also sent it to me from Siberia. He did pass to me at the same time, under the name *interrupta*, an individual which does not seem to differ in any way.” Years later Chaudoir (1870:210) published the name *Lebia interrupta*. Lutshnik realized that this appeared to be a case of homonymy, and published *Lebia tschitscherini* in 1922: 70 as a replacement name for *Lebia interrupta* Chaudoir (1870: 210) [nec Dejean, 1825]. Csiki (1933: 1320) then listed the name *Lebia interrupta* Dejean as a variety of *Lebia cruxminor*. However, *Lebia interrupta* Dejean (1825:262) is not a valid binomen, and as such should be struck down. *Lebia interrupta* Chaudoir 1870 is given new status as a valid species, and *Lebia tschitscherini* Lutshnik (1922: 70) becomes a synonym of *Lebia interrupta* Chaudoir 1870.

- 1563#. *Lebia latifascia* Chaudoir, 1870
 TAB⁵⁹; (type locality: “Tabasco, México”, Chaudoir 1870: 238)
 1564. *Lebia lecontei* Madge, 1967
 NL², QRO³
 1565*. *Lebia marginicollis* Dejean, 1825
 Type locality: “Georgia”, Dejean 1825: 271
 1566. *Lebia melanocephala* (Chaudoir, 1870)
 CAMP⁴, GRO⁴, GTO⁴, MICH⁴, VER⁴
 1567. *Lebia melanthera* Bates, 1883
 VER¹⁷
 1568. *Lebia mesostigma* Bates, 1883; Figure 2M
 CHIS², MICH², OAX¹⁷, YUC^{3,7}
 1569. *Lebia microtes* Bates, 1883
 CHIS^{3,7}, TAMPS³
 1570. *Lebia miranda* (G. Horn, 1872)
 BCS⁵, MOR⁵, SIN⁵, SON^{2,3,5,6,54}
 1571. *Lebia neanthe* Bates, 1883
 CHIS², OAX², PUE², TAMPS², VER², YUC²
 1572#. *Lebia nigriceps* Chaudoir, 1870
 Type locality: “México”, Chaudoir 1870: 242
 1573. *Lebia ocelligera* Bates, 1883
 HGO²⁴, SIN³, SLP², VER^{2,4,17}
 1574. *Lebia picicollis* Chaudoir, 1871
 CHIS², MICH², NAY², OAX², VER^{2,3,52}
 1575. *Lebia pimalis* (Casey, 1920)
 HGO^{3,5}, SON^{2,3,5,54}
 1576*. *Lebia pleuritica* LeConte, 1848
 Type locality: “Habitat ad Lacum Superiorem [Lake Superior]”, LeConte 1848: 193
 1577. *Lebia pleurodera* Chaudoir, 1871
 CHIH², CHIS², OAX², SIN², TAB²
 1578. *Lebia pulchella* Dejean, 1826
 DGO², HGO²⁴, NL²
 1579. *Lebia pumila* Dejean, 1831
 COL³, GRO³, SLP³
 1580. *Lebia quadriannulata* (Bates, 1878)
 TAB³
 1581. *Lebia quadricolor* Chevrolat, 1834
 CHIS^{2,7}, HGO², OAX², SLP^{2,7}, VER^{2,7}

1582. *Lebia quadrinotata* (Chevrolat, 1835)
CHIH², CHIS^{2,3,7}, COL⁴, GRO², JAL^{2,3}, MICH^{2,3},
MOR^{2,4,7}, NAY², PUE^{3,7}, SIN⁷, SON^{2,54}, TAMPS²,
VER^{3,4}, ZAC³
1583. *Lebia quadriplagiata* (Chaudoir, 1871)
VER^{3,52}
1584. *Lebia retusa* Bates, 1883
MOR³, SLP²
1585. *Lebia rhodope* Bates, 1883
CHIS², MOR², VER^{2,17}
1586. *Lebia rhombifera* Louwerens, 1953
MICH⁴; **note**: most likely mis-identified or mis-la-
belled; type locality: “East Sumba, Melolo” [Indone-
sia], Louwerens 1953: 314
1587. *Lebia rufilia* Bates, 1883
VER¹⁷
1588. *Lebia rufolimbata* (Chaudoir, 1870)
CHIS², MEX², MICH², MOR², VER^{2,3}
1589. *Lebia rufopyga* Chaudoir, 1870
MEX², OAX², PUE², QRO², VER²
- 1590*. *Lebia rufosutura* Motschulsky, 1864
Type locality: “Nicaragua”, Motschulsky 1864: 225;
“México” (Blackwelder 1944: 55; Erwin et al. 1977:
4.63)
1591. *Lebia rugatifrons* (Chaudoir, 1871)
CHIS², COL², MOR², NL², OAX², TAMPS², VER²
1592. *Lebia scalpta* Bates, 1883
MOR², SLP², SON^{2,3,6,54}, VER¹⁷, YUC¹⁷
1593. *Lebia scitula* Chaudoir, 1870
CAMP², CHIS², MOR², TAMPS², VER², YUC²
1594. *Lebia sellata* Dejean, 1825
CHIS²; type locality: “Cayenne”, Dejean 1825: 260;
“Nicaragua, Costa Rica” (Blackwelder 1944: 56);
“México” (Erwin et al. 1977: 4.61)
1595. *Lebia smithiella* Bates, 1891
GRO¹⁹, JAL¹⁹
1596. *Lebia solea* Hentz, 1839
JAL³
1597. *Lebia soror* Chaudoir, 1871
VER²
1598. *Lebia subdola* Madge, 1967
BCN², SON^{2,54}
1599. *Lebia subgrandis* Madge, 1967
CHIH², SIN⁵, SON^{2,3,5,6,54}
1600. *Lebia subrugosa* Chaudoir, 1871
MEX², MICH², SON^{2,54}, VER⁴
- 1601#. *Lebia tenenbaumi* Liebke, 1938
VER⁵⁹; (type locality: “Orizaba, [state of Veracruz],
México”, Liebke 1938b: 286)
1602. *Lebia tolteca* Bates, 1883
VER¹⁷
1603. *Lebia translucens* (Bates, 1883)
VER¹⁷
1604. *Lebia tricolor* Say, 1823
CHIS³, COL³, GRO³, JAL³, MOR³, OAX³, PUE³,
QRO³, TAMPS³, VER³
1605. *Lebia tuckeri* Casey, 1920
CDMX⁵, CHIS², GRO², MEX², MICH², MOR², OAX²,
SIN², SON^{2,3,6,54}, VER²
1606. *Lebia urania* (Bates, 1883)
CHIS², COL², JAL², VER⁵⁰
1607. *Lebia variegata* Dejean, 1831
CHIS², GRO², NAY², TAMPS², VER²
1608. *Lebia vicina* (Chaudoir, 1871)
GTO⁴
1609. *Lebia viridis* Say, 1823)
BCN⁵, CHIH^{2,3}, CHIS², COAH^{2,3,4}, DGO², MEX⁴,
MICH^{3,5,6}, MOR⁵, NL^{2,3}, OAX², PUE², QRO², SLP²,
SON^{2,3,6,54}, TAMPS^{2,3}
1610. *Lebia vittata* (Fabricius, 1777)
OAX⁵
1611. *Lebia xanthogaster* (Bates, 1883)
CAMP², CHIS², MOR², OAX², PUE², SLP², TAMPS^{2,3},
VER^{2,3,17}
1612. *Lebia xanthogona* Bates, 1883
VER¹⁷
- 1613#. *Lebia yucatana* Chaudoir, 1870
YUC⁵⁹; (type locality: “Yucatan, [México]”, Chaudoir
1870: 239)
1614. *Lebia zeta* Bates, 1883
HGO³, VER¹⁷
1615. *Lebia bicolor* Chaudoir, 1869
CHIS^{1,5,46}, VER^{1,46}
- 1616#. *Microlestes balli* Mateu, 1974
JAL⁴⁶; (type locality: “Tecalitlai, Jalisco, [México]”,
Mateu 1974a: 270)
- 1617*. *Microlestes brevilobus brevilobus* Lindroth, 1969
Type locality: “Long Point, Ont. [Ontario, Canada]”,
Lindroth, 1969: 1054; “Canada and United States”
(Bousquet 2012: 1304)
1618. *Microlestes brevilobus mexicanus* Mateu 1974
JAL^{2,46}, MICH^{2,46}
1619. *Microlestes halffteri* Mateu, 1974
DGO^{2,46}, HGO²⁵, SLP^{2,46}
- 1620#. *Microlestes lucidus subdeserticus* Mateu, 1974
DGO^{21,46}; (type locality: “Mapimi, Durango, [Méxi-
co]”, Mateu 1974a: 264; [paratypes from Queretaro]);
QRO^{2,21,46}
- 1621*. *Microlestes minutulus* (Goeze, 1777)
No type locality/area by Goeze, 1777: 665; “México,
Guatemala” (Blackwelder 1944: 58)
- 1622#. *Microlestes tehuantepec* Mateu, 1974
CHIS^{2,46}, OAX⁴⁶, TAB⁴⁶, VER⁴⁶
1623. *Negrea aldrete* Mateu, 1982
NL⁵⁹, VER³
1624. *Negrea chiapanensis* Mateu, 1975
CAMP², CHIS^{2,7}, QROO², SLP⁷, VER^{2,5}
1625. *Negrea mexicana* Mateu, 1975
CHIS⁷, TAMPS², VER^{2,5}
1626. *Nemotarsus limbicollis* Bates, 18831
CHIS², GRO², MOR², VER¹⁷
1627. *Nemotarsus rhombifer* Bates, 1883
CHIS², VER¹⁷
1628. *Onota angulicollis* Reiche, 1842
CAMP⁴, VER⁴
1629. *Philophuga brachinoides* Bates, 1883
OAX², VER¹⁷

1630. *Philophuga caerulea* Casey, 1913; Figure 2N
AGS², HGO²⁴, JAL³, MEX², SLP³, SON^{2,3,54}
1631. *Philophuga viridicollis* (LeConte, 1846)
CDMX³, MOR³, NL³, QRO³
1632. *Philophuga viridis* Dejean, 1831
CHIH⁵, JAL⁵, MEX⁵, NAY⁵, SON^{5,54}
1633. *Phloeoxena batesi* Ball, 1975
VER⁵⁰; (type locality: “S. Geronimo, Guatemala”, Ball 1975: 199)
1634. *Phloeoxena concolor* Ball, 1975
GRO⁴, MOR², NAY²
- 1635[#]. *Phloeoxena davidsoni* Shpeley and Ball, 2000
GRO⁵⁹; (type locality: “Fifteen kilometers northwest of El Paraiso, state of Guerrero, México”, Shpeley and Ball 2000: 83)
1636. *Phloeoxena geniculata* Chaudoir, 1869
CHIS⁷, COL⁵, JAL^{2,5}, MICH⁴, MOR^{2,5}, NAY^{5,33}
1637. *Phloeoxena henryi* Shpeley and Ball, 2000
CHIS²
1638. *Phloeoxena herculeano* Ball, 1975
CHIS^{2,3,5}
1639. *Phloeoxena limbicollis* Bates, 1884
OAX²
1640. *Phloeoxena newtoni* Ball, 1975
NL³, TAMPS²
1641. *Phloeoxena nigricollis* Ball, 1975
CHIS^{2,3,7}, OAX², VER³
- 1642[#]. *Phloeoxena nitida* Shpeley and Ball, 2000
OAX⁵⁹; (type locality: “51.5 kilometers south of Valle Nacional, State of Oaxaca, México”, Shpeley and Ball 2000: 110)
1643. *Phloeoxena obscura* Shpeley and Ball, 2000
VER^{2,3}
1644. *Phloeoxena picta picta* Chaudoir, 1869
CHIS^{3,7}, VER^{2,3,5,7,52}
- 1645[#]. *Phloeoxena picta apicalis* Ball, 1975
OAX²; (type locality: “México, Oaxaca, 16.9 mi. s. Valle Nacional”, Ball 1975: 143)
1646. *Phloeoxena picta franiae* Ball, 1975
CHIS²; (type locality: “Guatemala, Dpto. Huehuetenango, Cuchumatanes Mts., 5.6 mi. N. Santa Cruz Barillas”, Ball 1975: 143)
1647. *Phloeoxena picta unicolor* Chaudoir, 1869
CHIS²
1648. *Phloeoxena signata* (Dejean, 1825)
CAMP², CHIS^{2,5,7}, JAL², OAX², SIN², SLP^{2,7}, TAMPS⁵, VER²
- 1649[#]. *Phloeoxena totontepec* Shpeley and Ball, 2000
OAX²
1650. *Phloeoxena undata* Chaudoir, 1869
HGO⁷, SLP^{3,7}, TAMPS^{2,3,4}, VER^{2,3}
1651. *Plochionus amandus vittatus* (LeConte, 1844)
CHIS², VER²
1652. *Plochionus circumseptus* (Bates, 1883)
CHIS^{2,7}
1653. *Plochionus formosus* Bates, 1883
OAX², VER¹⁷
1654. *Plochionus incultus* (Bates, 1883)
OAX²
1655. *Plochionus pallens* (Fabricius, 1775)
VER⁴
1656. *Plochionus timidus* Haldeman, 1843
BCN⁵, BCS^{2,5}, SON^{2,5,54}
1657. *Pseudotoglossa marginella* Bates, 1883
CHIS³, TAMPS³
1658. *Pseudotoglossa rufitarsis* (Chaudoir, 1877)
CHIS², NL², SLP^{2,7}, TAMPS²
1659. *Pseudotoglossa terminalis* (Chaudoir, 1872)
SLP², YUC²
1660. *Stenognathus chadoiri* Ball, 1975
CHIS^{2,7}, OAX²
1661. *Stenognathus quadricollis* Chaudoir, 1869
CHIS⁷
- 1662*. *Technophilus croceicollis* Ménétériés, 1843
Type locality: “Californie”, Ménétériés, 1843: 53; “México” (Erwin et al. 1977: 4.61)
1663. *Xenodromius brachinoides* Mateu, 1976
HGO²⁵, MEX², ZAC²
1664. *Xenodromius curtulus* Mateu, 1976
MEX²
1665. *Xenodromius flohri* Bates, 1891
VER^{3,19}
1666. *Xenodromius microphthalmus* Mateu, 1976
PUE⁴
- 1667[#]. *Xenodromius reyesi* Mateu, 1976
VER⁵⁹; (type locality: “Perote, Veracruz, [México]”, Mateu 1976: 144)
- 1668[#]. *Xenodromius testaceous* Mateu, 1976
PUE²

42. Zuphiini

1669. *Pseudaptinus elegans* (Chaudoir, 1863)
CHIS², TAB²
1670. *Pseudaptinus hoegei* (Bates, 1883)
COAH^{2,3}, JAL³, SIN³, VER¹⁷
1671. *Pseudaptinus horni* (Chaudoir, 1872)
BCS⁵, CHIS², COAH⁵, COL², HGO^{3,5}, JAL², MOR⁴, NAY⁵, NL², SIN², TAMPS², VER²
1672. *Pseudaptinus lecontei* (Dejean, 1831)
JAL², SLP²
- 1673[#]. *Pseudaptinus magicus* Liebke, 1934
Type locality: “México”, Liebke 1934: 377; (Liebke 1934: 377 writes: “México without further information”)
1674. *Pseudaptinus nobilis* Liebke, 1934
MOR²; (type locality: “México”, Liebke 1934: 381; (Liebke 1934: 381 writes: “México without further information”)
1675. *Pseudaptinus rufulus* LeConte, 1851
CAMP², CHIS², COAH^{2,3}, GRO², MOR², NL², PUE², SLP², TAB², VER²
1676. *Pseudaptinus simplex* Liebke, 1934
COL², PUE², SIN², SON⁵⁴
1677. *Pseudaptinus tenuicollis* (LeConte, 1851)
CHIS², COL², GRO², JAL², MEX⁵, MICH², OAX², SIN^{3,5}, SON⁶, TAMPS², VER^{2,5}
1678. *Zuphioides americanum* Dejean, 1831
NAY⁵, SIN⁵

1679. *Zuphioides exquisitum* Liebke, 1933
GRO^{2,5}, MOR⁵
1680. *Zuphioides flohri* Liebke, 1933
MICH², NAY²
1681. *Zuphioides longicollis* LeConte, 1879
BCN⁵, COAH², SIN^{2,5}
1682. *Zuphioides magnum* Schaeffer, 1910
CHIS², NL², OAX², SLP², TAB², VER²
1683. *Zuphioides mexicanum* Chaudoir, 1863
CHIS², NAY², NL², QROO², TAB², TAMPS², VER²
1684. *Zuphioides punctipenne* Bates, 1891
CDMX⁵, COL², GRO², MOR^{5,19}

43. Galeritini

1685. *Ancystroglossus gracilis* Chaudoir, 1863
VER⁴
1686. *Ancystroglossus ovalipennis* Reichardt, 1967
JAL², NAY⁵, SIN⁵⁹
1687. *Galerita aequinoctialis* Chaudoir, 1852
CAMP², CHIS^{1,2,5}, COL², DGO¹, GRO^{1,2,5}, HGO²⁴,
JAL², NAY^{2,5}, NL², OAX², PUE⁴, QROO², SIN^{1,2,5},
SLP^{1,2,5,7}, TAMPS^{1,2,3}, VER^{1,2,3,5}, YUC¹
1688. *Galerita atripes* LeConte, 1858
CHIH², SON^{1,2,54}
1689. *Galerita azteca* Reichardt, 1967
GRO^{1,2,3}, OAX²
1690. *Galerita balli* Reichardt, 1976
OAX^{1,2}
1691. *Galerita bicolor* Drury, 1773
MOR⁴, NAY⁴
1692. *Galerita boucardii* Chaudoir, 1869
COAH⁵, COL², DGO^{1,2,5}, GRO², GTO⁴, JAL^{1,2,5},
MEX^{1,2,5}, MICH^{1,2,5}, MOR^{1,5}, NAY^{1,2,5}, OAX¹, SIN^{2,5}
1693. *Galerita forreri* Bates, 1883; Figure 2O
CHIH^{2,5}, COL^{1,5}, GRO^{1,5}, JAL^{3,4}, NAY^{1,5}, SIN^{1,2,3,5,17},
SON^{1,2,3,54}
1694. *Galerita janus* (Fabricius, 1792)
CHIH¹, MICH⁴, NAY³, SLP³
1695. *Galerita laevithorax* Reichardt, 1967
CHIS^{2,3,4,5}, VER²
1696. *Galerita lecontei lecontei* Dejean, 1831
BCS^{3,5}, CHIH¹, DGO¹, JAL², MICH³, SIN^{2,5}, SON^{2,54},
VER³
1697. *Galerita lecontei bicoloripes* (Reichardt, 1967)
GTO^{1,2}, JAL¹, MEX⁵, MICH^{1,59}, NAY^{1,2}
1698. *Galerita lecontei veracruzis* Ball and Nimmo, 1983
TAMPS^{1,2}, VER^{1,2,5}
1699. *Galerita mexicana* Chaudoir, 1872; Figure 2P
AGS³, CHIH², CHIS^{2,3,5}, COAH¹, COL^{2,5}, DGO^{1,2,6,7},
GRO^{1,2,3}, HGO^{3,24}, JAL^{2,3,4,5,6}, MICH^{1,2,3,4,5}, MOR^{1,2,3,4,5},
NAY^{1,2,3,5,6}, NL¹, OAX^{1,2,3,5,6}, PUE^{2,5}, QRO^{2,3}, SIN^{1,3,5},
SLP^{1,2,3,5}, SON^{2,3,5,6,54}, TAMPS^{2,3}, VER^{2,4,5}
1700. *Galerita nigra* Chevrolat, 1835
PUE¹, QRO², SIN¹, SLP^{1,2}, TAB⁵, TAMPS^{1,2}, VER^{1,2,4,5}
1701. *Galerita reichardti* Ball and Nimmo, 1983
CHIS², VER^{1,2,3,5}
1702. *Galerita ruficollis* Dejean, 1825
CAMP^{1,2}, CHIS^{1,2,3,5}, COL^{2,5}, GRO^{1,2,3,5,7}, JAL^{2,3,4,5,6},
MICH², MOR^{1,2,3,5,6}, NAY^{1,2,5}, OAX^{1,2,5}, PUE²,

SIN^{1,2,3,5,6}, SLP^{2,5,7}, SON^{1,2,3,5,6,54}, TAMPS², VER^{1,2,3,4,5,6},
YUC^{1,3}

1703. *Galerita simplex* Chaudoir, 1852
NAY², SIN², TAB², VER^{1,2,3}
1704. *Galerita strandi* Liebke, 1939
CHIS³, MICH^{3,4}
1705. *Galerita sulcipennis* Reichardt, 1967
CHIS^{1,2,3,5,7}, OAX², VER²

44. Helluonini

1706. *Helluobrochus cribratus* (Reiche, 1843)
OAX², CHIS⁵
1707. *Helluomorphoides balli* Reichardt, 1974
OAX²
1708. *Helluomorphoides ferrugineus* (LeConte, 1853)
AGS³, CHIH^{3,5,6}, COAH², COL⁴, GRO⁴, JAL², NAY⁵,
NL², SIN⁵, SON^{2,3,5,6,54}, TAMPS^{2,3,5,6}
1709. *Helluomorphoides latitarsis* Casey, 1913
CHIH⁵, COAH⁵, DGO⁵, HGO²⁷, JAL^{3,6}, SIN⁵,
SON^{2,3,5,6,54}
1710. *Helluomorphoides mexicanus* (Chaudoir, 1872); Figure 2Q
CHIS^{2,5}, COL², GRO², MICH², PUE², TAMPS²
1711. *Helluomorphoides papago* (Casey, 1913)
CHIH^{2,5}, DGO²
1712. *Helluomorphoides texanus* (LeConte, 1853)
COAH², TAMPS²

45. Pseudomorphini

- 1713#. *Pseudomorpha arrowi* Notman, 1926
DGO⁵⁹; (type locality: "Ciudad, Durango, México",
Notman 1926: 20)
1714. *Pseudomorpha angustata* G. Horn, 1883
SON⁵
- 1715*. *Pseudomorpha behrensi* G. Horn, 1870
Type locality: "west of the Sierras", G. Horn 1870: 76;
"México" (Blackwelder 1944: 72; Erwin et al. 1977:
4.66)
1716. *Pseudomorpha huachinera* Amundson and Erwin,
2013; Figure 2R
CHIH¹, SON^{1,5,54}
1717. *Pseudomorpha peninsularis* Van Dyke, 1953
BCN^{1,5}, BCS⁵
1718. *Pseudomorpha pilatei* Chaudoir, 1862
NL¹, YUC⁵⁹; (type locality: "Yucatan, [México]",
Chaudoir, 1862: 490)
1719. *Tuxtlamorpha tuxtla* (Liebherr and Will, 1997)
VER^{1,4}

DISCUSSION

This checklist recognizes 1,634 described carabid species in 45 tribes represented by 179 genera which are recorded from Mexico, based on over 67,100 digital records plus published records. There are 1,029 species (bold font checklist species number) which were described from type material collected only in Mexico. Of that number, 204 species (number sign (#) follows checklist species number) are currently known only from the type specimen or type specimens. There are 65 species (indicated with an aster-

Table 4. Number of carabid tribes, genera and described species for each of the states in Mexico.

State	Tribes	Genera	Species	State	Tribes	Genera	Species
Aguascalientes	15	24	53	Morelos	30	71	244
Baja Cal. Norte	25	54	159	Nayarit	27	62	210
Baja Cal. Sur	23	52	120	Nuevo León	27	60	177
Campeche	19	38	91	Oaxaca	36	102	485
Chiapas	32	111	470	Puebla	33	65	238
Chihuahua	29	61	253	Querétaro	24	50	127
Ciudad de México	21	46	134	Quintana Roo	18	25	32
Coahuila	28	55	148	San Luis Potosí	35	79	277
Colima	22	51	142	Sinaloa	31	74	237
Durango	28	63	243	Sonora	34	89	340
Guanajuato	18	36	111	Tabasco	21	44	120
Guerrero	32	76	282	Tamaulipas	32	81	252
Hidalgo	29	59	177	Tlaxcala	13	21	48
Jalisco	32	76	299	Veracruz	38	130	609
México (state)	26	55	216	Yucatán	18	34	85
Michoacán	25	59	222	Zacatecas	14	29	71

isk (*) after the checklist species number), some of which have been included in previous Mexico lists by Blackwelder (1944) and Erwin et al. (1977) for which there are no confirmed Mexico records. Perhaps one day specimens of some of these species will be collected in Mexico. Lastly, in this checklist, there are 19 species recorded from Mexico which have their identification or locality data questioned, and therefore, are not included in any of the species counts. Each of these 19 species are indicated with the following: “note: most likely mis-identified or mis-labelled”.

Even though the size of the data set is rather large with over 69,300 digital records as well as published records, 617 (37.8%) of the 1,634 described species are represented by single state records. There are 29 described species which have a type locality of only “Mexico”, and no recorded specimens in this checklist, other than type material. They are included in this count, as the type material would have most likely come from a single state which was not recorded at the time of collection. An additional 174 described species also have a type locality of only “Mexico”, but there are specimen records from one or more states for those species in this checklist.

The rather high percentage of single state records may be due to lack of collecting effort in those states, lack of suitable habitat in those states, lack of dispersal capability, usually indicated by brachyptery, or other factors we are unaware of. Many of the cave dwelling trechines are restricted to a single cave or cave system within a given state. On most of the collecting trips in Mexico, the colleagues and I were interested in higher elevation forests in which to collect, and usually just drove through the lowlands to the next collecting site without stopping. Perhaps, some of these species may truly be rare. Gidaspow (1963) stated that *Calosoma scrutator* was rare in Mexico. After 62 years since her publication, this species is currently found in 21 states in Mexico, and is recorded from as far south as Nicaragua (Bruschi, 2008). This is a good example of

the dispersal capability by a large macropterous carabid. Only 144 species were each collected in 10 or more different states. Of those 144 species, only 22 species are more widespread, with records in 20 or more states. *Cicindelidia ocellata ocellata* appears to have the widest distribution in Mexico, with records from 27 states (currently not recorded from BCN, BCS, MEX, QROO and TLAX).

Of the 179 genera recorded from Mexico, some genera are very diverse with many species, whereas other genera have fewer species. Some of the genera with large numbers of species are the following, with number of species in parentheses: *Dyscolus* (117), *Lebia* (101), *Cicindelidia* (74), *Bembidion* (59), *Calosoma* (53), *Chlaenius* (46), *Sele-nophorus* (42), *Brachinus* (35), *Calleida* (33), *Pasimachus* (28) and *Platynus* (24). At the other end of the spectrum, 108 genera each have 5 or fewer species, too many to list here. And of those 108 genera, 41 genera are each represented by a single species.

Burgos-Solorio and Trejo-Loyo 2001 recorded 62 species of Carabidae plus one species of Rhysodidae (here treated as a member of the family Carabidae) for a total of 63 species in Morelos. Only 38 of the species listed were determined to the species level; the remaining 25 taxa were determined to the genus level only, so they are morphospecies. Zaragoza-Caballero et al. 2019 recorded 148 species of Carabidae plus one species of Rhysodidae (here treated as a member of the family Carabidae) for a total of 149 species in Morelos. However, 26 of those 149 are morphospecies, which brings the total described species in Morelos in that paper to 123 species. Interestingly, the number of taxa recorded for Morelos in this checklist is 244 described species (morphospecies are not included in this checklist) substantially higher than the number published by Zaragoza-Caballero et al. 2019. Cerón-Gómez et al. (2022) recorded 134 species from the state of Hidalgo, five of which were morphospecies, so the number of described species for Hidalgo in that paper would be 129 species. This checklist

records 177 described species for Hidalgo. Cerón-Gómez et al. (2025a) recorded 152 species (132 species plus 20 sub-species) from the Baja California Peninsula which is made up of two Mexican states. Cerón-Gómez et al. (2022: 2; 2024: 1; 2025b: 45) had the potential species richness for Mexico at 2,012 species. Table 4 is a summary of the number of tribes, genera and described species recorded from each of the states in Mexico based mainly on digital records but also published records. The states with the highest diversity, expressed by numbers of described species, are Veracruz (609 species), Oaxaca (485 species), and Chiapas (470 species). This is the same conclusion as stated for the order Coleoptera by Zaragoza-Caballero et al. (2019: 16).

CONCLUDING REMARKS

At this point in time, the carabid fauna of Mexico consists of 45 tribes, 179 genera and 1,634 described species. The genera *Miopotachys*, *Thalassotrechus*, *Homopterus*, *Piesmus*, *Oodes*, *Homalomorpha* and *Hyboptera*, though included in this checklist as potentially occurring in Mexico, are not included in the preceding totals as there are no confirmed records of specimens collected in Mexico. This checklist has 13 more genera and 367 more described species than the number recorded in the year 2000. If the 440 morphospecies believed to represent new species in the University of Alberta Strickland Museum were added to the current count of 1,634 described species, that could increase the potential species richness of carabids in Mexico to 2,074 species. I do not claim that this checklist is definitive for the Mexican carabid fauna. More taxonomic studies in the form of revisions of genera are needed to eventually identify the large numbers of undetermined morphospecies. And, as other studies are undertaken, similar to those of Zaragoza-Caballero et al. (2019), Cerón-Gómez et al. (2022) and Cerón-Gómez et al. (2025a) for the other states or regions in Mexico, I predict that additional carabid species will be added to this list in the future.

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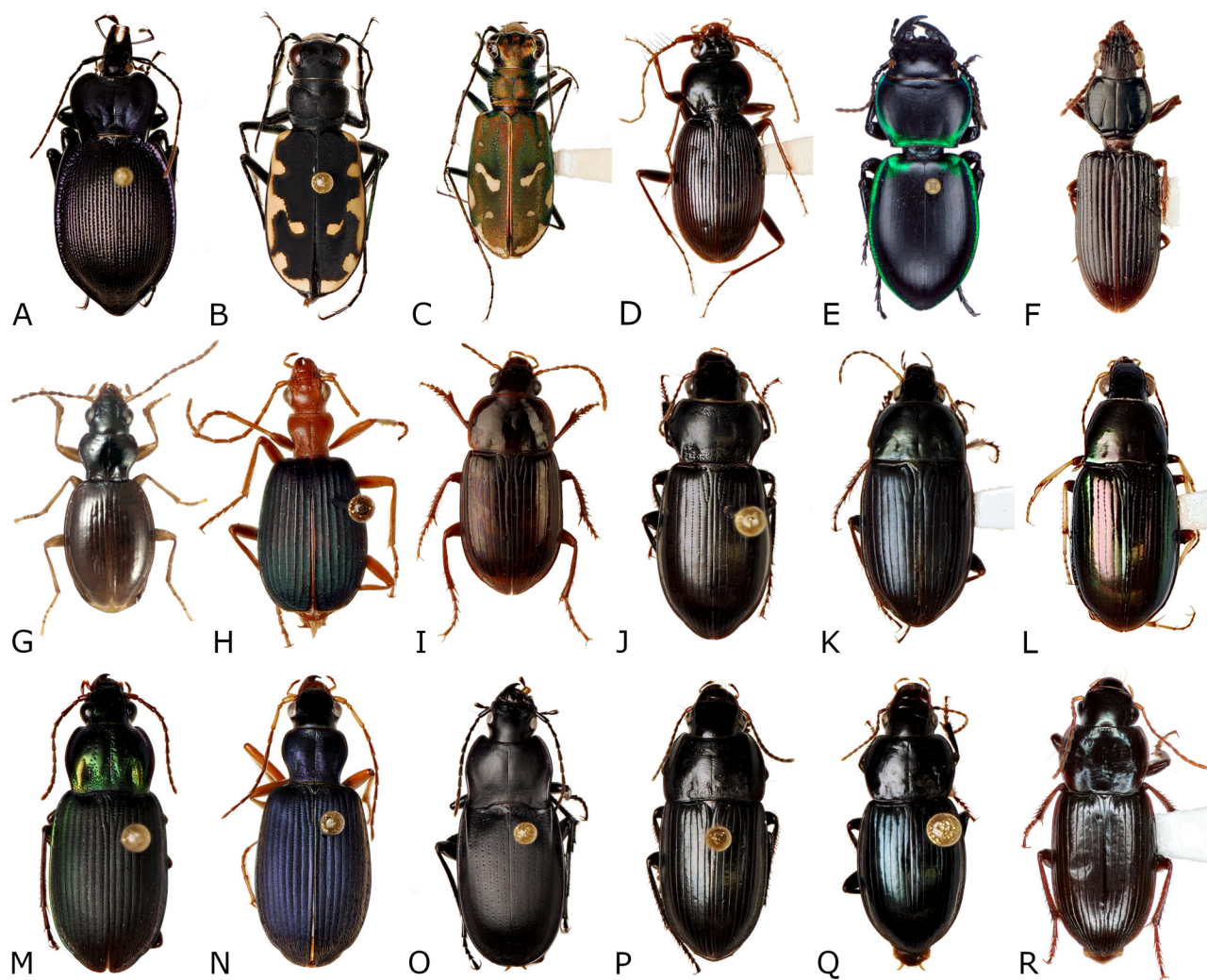


Figure 1. Digital images by author (except where indicated) of selected Mexican carabidae species, followed by state in which the specimen was collected, the overall body length and Strickland Museum accession number (only for UASM specimens). **A.** *Scaphinotus mexicanus* Bates (Chihuahua; 18.5 mm; UASM368357); **B.** *Cicindelidia nebuligera* (Bates) (Durango; 10.0 mm; UASM347606); **C.** *Cicindelidia obsoleta santaclarae* (Bates) (Sonora; 17 mm; UASM372601); **D.** *Loricera aptena* Ball and Erwin (Durango; 8.5 mm; UASM347749); **E.** *Pasimachus viridans* LeConte (Sonora; 27.0 mm; UASM438009); **F.** *Schizogenius longipennis* Putzeys (Sonora; 6.0 mm; UASM161815); **G.** *Bembidion elescarabajo* Maddison (Oaxaca; 3.5 mm; paratype; image courtesy of David R. Maddison); **H.** *Brachinus oaxacensis* Erwin (Oaxaca; 10.5 mm; UASM369003; allotype); **I.** *Amara chihuahuae* (Casey) (Nuevo Leon; 6.5 mm; UASM303636); **J.** *Amara durangensis* Van Dyke (Durango; 12.5 mm; UASM347993); **K.** *Amara sera* Say (Chihuahua; 8.4 mm; UASM422668); **L.** *Stenocrepis tibialis* (Chevrolat) (Tamaulipas; 7.5 mm; UASM324873); **M.** *Chlaenius forreri* Bates (Sonora; 14.5 mm; UASM386284); **N.** *Chlaenius leucoscelis leucoscelis* Chevrolat (Sonora; 15.0 mm; UASM388287); **O.** *Dicaelus suffusus* (Casey) (Sonora; 15.0 mm; UASM370579); **P.** *Anisodactylus anthracinus* (Dejean) (Oaxaca; 12.5 mm; UASM321109); **Q.** *Aztecarpalus trochotrichis* Ball (Oaxaca; 11.0 mm; UASM422669); **R.** *Discoderus shpeleyi* Messer and Raber (Oaxaca; 7.5 mm; UASM315346; holotype; image courtesy of Peter W. Messer and Brian T. Raber).

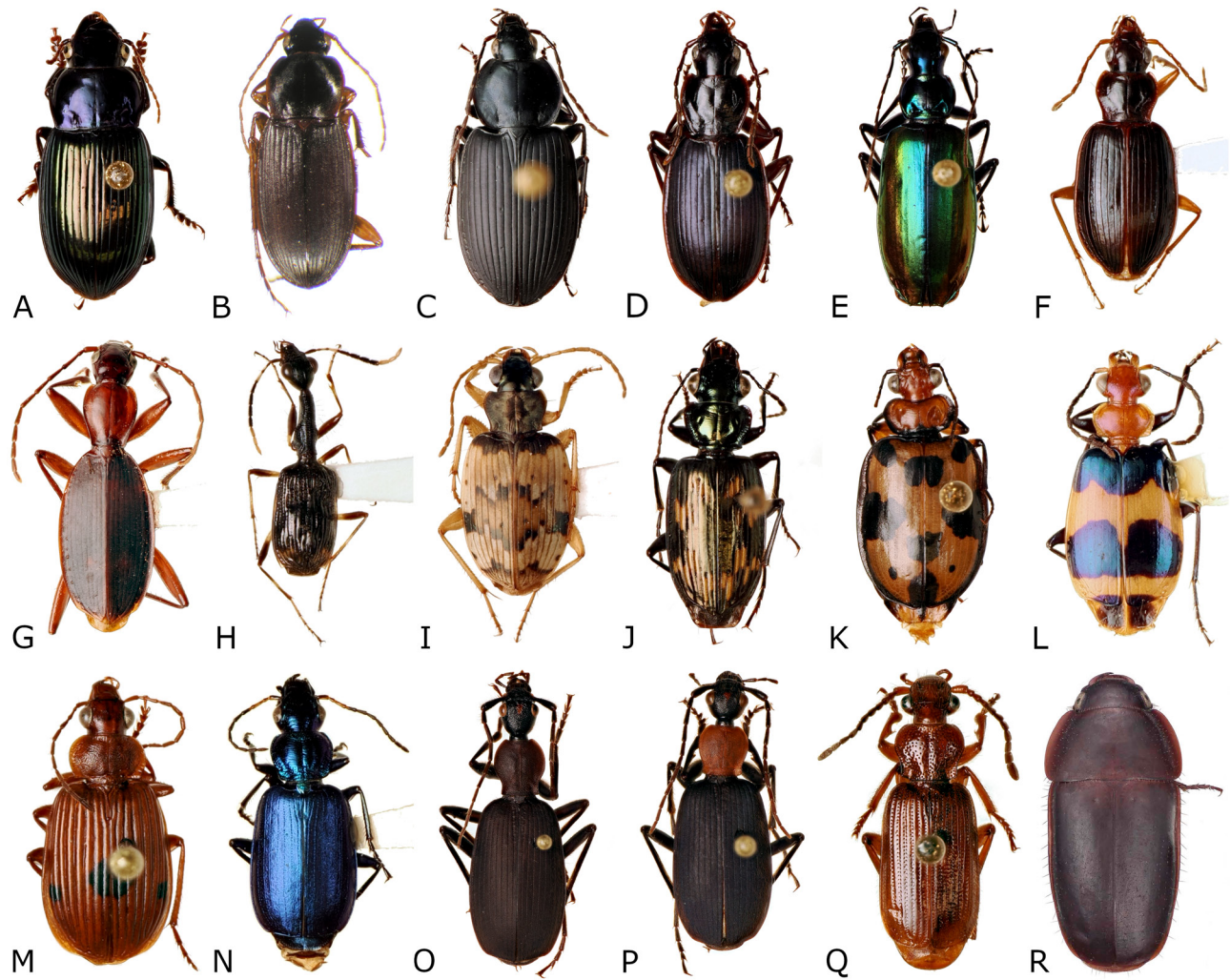


Figure 2. Digital images by author (except where indicated) of selected Mexican carabidae species, followed by state in which the specimen was collected, the overall body length and Strickland Museum accession number (for UASM specimens). **A.** *Notiobia lamprota* (Bates) (Veracruz; 13.0 mm; UASM319038); **B.** *Calathus maclevei* Shpeley and Van Devender (Sonora; 9.5 mm; holotype); **C.** *Agonum anthracinum* Dejean (Sinaloa; 12.5 mm; UASM274099); **D.** *Dyscolus durangensis* (Bates) (Durango; 13.5 mm; UASM347846); **E.** *Onypterygia fulgens* Dejean (Veracruz; 15.5 mm; UASM259759); **F.** *Platynus lyratus* (Chaudoir) (Sonora; 9.5 mm; UASM370343); **G.** *Rhadine leptodes* Bates (Durango; 8.0 mm; UASM347921); **H.** *Colliuris subdistincta* (Chaudoir) (Guerrero; 8.0 mm; UASM326407); **I.** *Tetragonoderus mexicanus* (Chaudoir) (Oaxaca; 5.0 mm; UASM422670); **J.** *Coptodera elongata* Putzeys (Veracruz; 12.0 mm; UASM340004); **K.** *Eurycoleus ornatus* Bates (Veracruz; 10.5 mm; UASM335976); **L.** *Lebia bitaeniata* Chevrolat (Chiapas; 5.5 mm; UASM327670); **M.** *Lebia mesostigma* Bates (Chiapas; 9.0 mm; UASM328585); **N.** *Philophuga caerulea* (Casey) (Puebla; 8.0 mm; UASM347361); **O.** *Galerita forreri* Bates (Sinaloa; 22.5 mm; UASM363545); **P.** *Galerita mexicana* Chaudoir (Sonora; 16.5 mm; UASM348226); **Q.** *Helluomorphoides mexicanus* (Chaudoir) (Tamaulipas; 11.5 mm; UASM362876); **R.** *Pseudomorpha huachinera* Amundson and Erwin (Sonora; 6.0 mm; image from Erwin and Amundson, 2013).

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<i>Chlaenius</i>	24	<i>Halocoryza</i>	14	<i>Pachyteles</i>	19
<i>Cicindela</i>	9	Harpalini	26	Panagaeini	24
<i>Cicindelidia</i>	10	<i>Harpalus</i>	27	<i>Panagaeus</i>	24
Cicindelini	9	<i>Helluobrochus</i>	45	<i>Paratrechus</i>	16
Clinidiini	7	<i>Helluomorphoides</i>	45	<i>Parvindela</i>	13
<i>Clinidium</i>	7	Helluonini	45	Pasimachini	13
<i>Clivina</i>	14	<i>Hemiplatynus</i>	34	<i>Pasimachus</i>	13
Clivinini	14	<i>Homalomorpha</i>	36	Paussini	19

Peleciini	24	<i>Polychaetus</i>	24	<i>Stenolophus</i>	29
<i>Pelmatellus</i>	28	<i>Pseudabarys</i>	22	<i>Stenomorphus</i>	30
<i>Pentagonica</i>	36	<i>Pseudaptinus</i>	44	<i>Stomis</i>	23
Pentagonicini	36	<i>Pseudomorpha</i>	45	<i>Synuchus</i>	30
<i>Percolaus</i>	22	Pseudomorphini	45	<i>Tachys</i>	19
<i>Pericompsus</i>	18	<i>Pseudotoglossa</i>	44	<i>Tachyta</i>	19
<i>Perigona</i>	36	Pterostichini	21	<i>Tanystoma</i>	36
Perigonini	36	<i>Pterostichus</i>	23	<i>Technophilus</i>	44
<i>Pheropsophus</i>	20	<i>Rawlinsius</i>	16	<i>Tetracha</i>	9
<i>Philophuga</i>	43	<i>Rhadine</i>	35	<i>Tetragonoderus</i>	37
<i>Phloeoxena</i>	44	<i>Scaphinotus</i>	7	<i>Thalassotrechus</i>	19
<i>Physea</i>	19	<i>Scarites</i>	13	Trechini	16
<i>Piesmus</i>	22	Scaritini	13	<i>Trechus</i>	16
Platynini	30	<i>Schizogenius</i>	14	<i>Trichopselaphus</i>	30
<i>Platynus</i>	35	<i>Selenophorus</i>	28	<i>Trichotichnus</i>	30
<i>Plochionus</i>	44	<i>Semiardistomis</i>	15	<i>Tuxtlamorpha</i>	45
<i>Poecilus</i>	22	<i>Sericoda</i>	36	<i>Xenodromius</i>	44
Pogonini	19	Sphodrini	30	Zabrini	23
<i>Pogonodaptus</i>	28	<i>Stenocrepis</i>	24	Zuphiini	44
<i>Polpochila</i>	28	<i>Stenognathus</i>	44	<i>Zuphioides</i>	44

***Pyropygodes huautlae* Zaragoza Caballero, 2000: deposición de tipos**

***Pyropygodes huautlae* Zaragoza Caballero, 2000: types deposition**

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ABSTRACT

Type deposition for *Pyropygodes huautlae* Zaragoza Caballero, 2000 is provided. Holotype and Allotype are in the National Collection of Insectss, Universidad Nacional Autónoma de México.

Key words: Type deposition, National Collection of Insects, Institute of Biology.

RESUMEN

Se indica donde se encuentran depositados los ejemplares Holotipo y Alotipo de *Pyropygodes huautlae* Zaragoza Caballero, 2000. Ambos se encuentran en la Colección Nacional de Insectos, Universidad Nacional Autónoma de México.

Palabras clave: Deposito de ejemplares tipo, Colección Nacional de Insectos, Instituto de Biología.

Con base en Código Internacional de Nomenclatura Zoológica vigente (ICZN 1999), el Art. 16.4.2: en la publicación original de todo nombre nuevo, debe acompañarse “de una declaración de la intención de, si el holotipo o los sintipos son ejemplares existentes, depositarlos (o haberlos depositado) en una colección y de una declaración indicando el nombre y la ubicación de la colección”. En este sentido, *Pyropygodes huautlae* fue descrito como género y especie nuevos en Dugesiana 7(1): 19-22 (Zaragoza-Caballero 2000), pero se omitió la indicación de la institución en donde se encuentra depositado el material tipo. Para solucionar dicha omisión, se precisa qué el Holotipo y Alotipo, están depositados en la Colección Nacional de Insectos (CNIN) del Instituto de Biología de la Universidad Nacio-

nal Autónoma de México, Circuito exterior s/n, Ciudad Universitaria, Universidad Nacional Autónoma de México, Coyoacán, 04510, Ciudad de México. Esta información se suma a la publicada por Zaragoza Caballero (2000) con la finalidad de hacer disponible los nombres propuestos en dicho trabajo.

LITERATURA CITADA

ICZN 1999. *International Code of Zoological Nomenclature*. Fourth Edition. International Commission on Zoological Nomenclature, London.

Zaragoza Caballero, S. 2000. Cantharoidea (Coleoptera) de México. Un nuevo género y una nueva especie de Lampyridae del estado de Morelos, México. *Dugesiana* 7(1): 19-22.

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First record of the genus *Didimioza* Quate and Brown in Mexico, with the description of *Didimioza noveloi* sp. nov. (Diptera: Psychodidae)

Primer registro del género *Didimioza* Quate and Brown en México, con la descripción de *Didimioza noveloi* sp. nov. (Diptera: Psychodidae)

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ABSTRACT

A new species of the Neotropical moth fly genus *Didimioza* Quate and Brown is described based on male characteristics. *Didimioza noveloi* sp. nov. is the fourth known species of this genus and represents the first record of the genus in Mexico. It is distinguished from other species of the genus by the angular curvature of the aedeagus and sigmoidal form of the paramere.

Key words: Moth fly; new species, Maruinini.

RESUMEN

Una especie nueva de moscas palomilla del género *Didimioza* Quate y Brown es descrita con base en las características del macho. *Didimioza noveloi* sp. nov. es la cuarta especie que se conoce del género y la primera que se registra en México. Se distingue de otras especies del género por la curvatura angular del edeago y la forma sigmoidal del parámetro.

Palabras clave: Moscas palomilla; nueva especie, Maruinini.

Quate (1999) described *Desmioza symphyllia* based on a male specimen collected in Barro Colorado Island, Panama. Later Quate and Brown (2004) determined that this species, along with two others collected in Venezuela and Peru, presented characteristics sufficiently different to be included in a new genus, *Didimioza* Quate and Brown, 2004.

Didimioza is recognized principally by the eyes separated, with interocular suture and eye bridge with four facet rows; antenna longer than wing width, with 13 flagellomeres of which the basal is enlarged, 2-11 pyriform with poorly defined internodes, flagellomere 12 without internode, flagellomere 13 with apiculus as long as base, ascoids paired in flagellomeres 1-12, unidigitate, longer than the flagellomere bearing them, wing with only small faint dark spots at vein tips and forks, radial and medial forks near the middle of the wing, and R_5 ending beyond wing apex (Quate and Brown 2004).

Before this work, three species were known: *Didimioza symphyllia* (Quate, 1999) known from Panama (type locality) and Costa Rica, *D. venezuelica* Quate and Brown, 2004, from Venezuela, and *D. chachapoya* Quate and Brown, 2004, from Peru.

In this work, a new species of *Didimioza* is described based on the male morphological characters from a specimen collected in southern Mexico.

MATERIAL AND METHODS

Collection site and field methods. Specimens were obtained through collections for the project: "Estratificación de tres enfermedades prioritarias en áreas de riesgo con

base en sus insectos vectores en el estado de Veracruz, México", granted to Sergio Ibáñez-Bernal by CONACYT FOMIX-Veracruz-Salud, 68317 (INECOL 1333), and additionally supported by the Scientific Collaborative Agreement between Servicios de Salud de Veracruz (SESVER) and the Instituto de Ecología, A.C. (INECOL).

The specimen was collected in an "acahual" (vegetation that covers fallow or crop fields after mowing or harvesting, particularly that formed by herbaceous plants or shrubs) near the locality of Medellín of the municipality Santiago Tuxtla, in the southern part of the state of Veracruz, Mexico (18°24'59"N 95°22'03"W, altitude 69 m).

The climate is warm and humid, with a temperature range of 24-26°C and rain most of the year. The vegetation in the Los Tuxtlas region has as dominant type the evergreen tropical forest as original vegetation, but some areas are transformed for agricultural use and for livestock breeding (INEGI 2010). This locality belongs to the Veracruz physiographical province (Morrone 2005; Durán-Luz and Ibáñez-Bernal 2025).

The specimen was collected using a Miniature CDC-UV light trap (Model 912, John W. Hock Company, Gainesville, FL.). The specimens were preserved in 70% ethanol.

Laboratory procedure. The specimen was cleared, dissected, and permanently mounted on slides following the procedure outlined by Ibáñez-Bernal (2024), with Euparal as permanent mounting medium (Bioquip Products, Inc., Rancho Dominguez, CA, USA). Specimens were examined using a Nikon Eclipse 50i phase contrast microscope. Measurements were obtained using an ocular micrometer

and are given in millimeters. Drawings were rendered with the aid of a Nikon Y-IDT drawing tube, artwork completed by mixed media drawing technique using Chinese ink and charcoal, and then digitally processed with Corel Photo Paint X3 (Version 13). The specimen is deposited in Colección de Insectos del Instituto de Ecología, A. C., Xalapa, Veracruz, Mexico (IEXA).

Terminology. The general morphological terminology of Cumming & Wood (2017) and Kvitte & Wagner (2017) is used, with the terms hypopod and gonocoxal condyle as proposed by Kvitte (2014). The term “frons” refers to the sclerite situated below the eyes at level of the antennal insertions, whereas “vertex” corresponds to the upper portion of the head above the eye bridge, and “face” the sclerite situated below the tentorial pits and above the oral margin that usually presents a patch of setae alveoli.

Measurements. Head width was taken at the widest part, whereas the length was measured from the vertex to the lower margin of the head. Wing length is measured from the beginning of the basal node of costa to the wing tip. Proportion of the palpus segments are given considering the length of palpus segment I as a unit (1.0).

TAXONOMY TREATMENT

Didimioza noveloi Ibáñez-Bernal, sp. nov.

(Figures 1-10)

<http://zoobank.org/BF20C716-01CA-499B-AC8A-2A0DF78F4E68>

Diagnosis. *Didimioza noveloi* sp. nov. differs from other species of the genus by the long and angularly curved aedeagus and the elongated and sigmoidal paramere.

Male description. Head (Figs. 1-2) oval in frontal view, vertex barely outstanding with surface covered with setae alveoli, except for a narrow longitudinal line in the middle. Eyes separated by 1.0 facet diameter, with an inverted Y-shaped interocular suture, eye-bridge with four rows of facets, and supraorbital macrosetae alveoli confined to the lateral and posterior portion of head. Frontal patch of alveoli complete, nearly quadrangular with upper margin slightly bilobed and with an upper extension of two rows of alveoli that reaches the level of the first inferior facet. Face with a quadrangular, longer than wide, patch of setae alveoli (Fig. 1). Occiput with about four supraorbital macrosetae, with a patch of alveoli reaching the posterior inferior angle of eye, separated from the margin of the oval foramen magnum (Fig. 2). Antenna longer than wing width; scape short, 2.3 times the length of the spherical pedicel (Fig. 4); flagellum with 13 flagellomeres, internodes poorly defined, those of basal flagellomeres short but progressively elongating (Fig. 1), flagellomere 12 without internode, flagellomere 13 with apiculus as long as its basal portion (Fig. 3). Ascoids much longer than the flagellomere of origin, paired on flagellomeres 1-12, unidigitate and strongly convoluted (Figs. 1, 3, 4). Palpus about 0.75 the length of the antenna, reaching the level of the flagellomere 8; proportion of palpal segments: 1.00: 1.37: 1.50: 2.30, last segment corrugated, fleshy (Fig. 1). Labium with an inverted Y-shaped sclerite and with six small spines at each side on internal margin and patches of long setae externally (Fig. 5).

Thorax with no allurement organs; anepisternum sparsely covered with setae alveoli, laterotergite with a patch of alveoli at center. Mid coxa with an anterior patch of small sensilla. Pretarsal claws at apex. Wing with small faint brown spots at R and M forks and at vein tips; costa without second costal node, Sc ending at level of the origin of R_5 , Rs not pectinate, R_{2+3} weakly joined to R_4 , R_3 with spur, radial and medial forks basal but near wing center, medial fork basal to radial fork, M_2 weakly joined to M_1 ; R_5 ending beyond wing apex (Fig. 6).

Male terminalia (Figs. 7-10). Epandrium nude, about as long as wide, the posterior margin concave and with a large single foramen; subepandrial sclerites as divergent laminar rods with irregular margins (Fig. 7). Hypopod longer than epandrium, tapering towards apex (Fig. 7), with one spatulate tenaculum at apex (Fig. 8). Epiproct triangular, hypoproct arrowhead shaped, more than twice the length of epiproct (Fig. 7). Hypandrium is a thin bar connecting the gonocoxites. Aedeagus strongly asymmetrical, basally broad with complex curvatures, apex long and lanceolate, bent near the middle almost at a right angle so the tip is laterally directed; ejaculatory apodeme expanded anteriorly, longer than aedeagus and with a large hole. Paramere broad at base with a pyramidal short protuberance and a long sigmoidal branch which is longer and somewhat narrower than the aedeagus, with the apex directed laterally in the opposite direction to the aedeagus (Fig. 9). Anterior gonocoxal condyles oval, greatly expanded, ending in a point anteriorly; posterior gonocoxal condyles rounded, nude, and in contact each other (Fig. 10). Gonocoxite short, with a patch of delicate sensilla on the inner area. Gonostylus longer than gonocoxite, simple, tapering towards apex (Figs. 9-10).

Measurements (in millimeters). Head height: 0.49; head width: 0.54; palpus length: 0.62; antenna length: 1.22; wing length: 2.16; Wing width: 0.82; epandrium length at middle: 0.10; hypopod length: 0.20; tenaculum length: 0.06; gonocoxite length: 0.10; gonostylus length: 0.14; aedeagus length: 0.16; paramere length: 0.18.

Female. Unknown.

Type material. Holotype male. Mexico, Veracruz, municipality Santiago Tuxtla, Medellín, 7-iv-2010, acahual, CDC light trap, Jurisdicción Sanitaria 10, colls.

Etymology. This species is named in honor of the prominent Mexican entomologist Rodolfo Novelo Gutiérrez, a specialist in the study of Odonata and other aquatic insect orders, in recognition to his work. Rodolfo Novelo has been a teacher, colleague and friend for more than 45 years.

Comments. Genus *Didimioza* has been placed in the tribe Maruinini Enderlein, 1937 (= Arisemini Vaillant, 1982; Setomimini Vaillant, 1982) (Quate and Brown 2004; Kvitte 2018) although its monophyly is questionable. The gonocoxal condyles of *Didimioza* are expanded as lobes, in contact with each other, but do not form a median keel that links to the ejaculatory apodeme, whereas the parameral sheath is untraceable (see Durán-Luz and Ibáñez-Bernal 2025), characteristics that are important for recognizing the tribe.

Didimioza noveloi sp. nov. is the fourth known species of this genus and the first recorded in Mexico. *Didimioza venezuelica* Quate and Brown differs because it has spines in the posterior margin of the posterior gonocoxal condyles, and the aedeagus and paramere straight.

Didimioza symphyllia and *D. chachapoya*, are similar to *D. noveloi* sp. nov., as the three species possess the posterior condyles destitute of cluster of spines in the posterior margin, but *D. chachapoya* has the gonocoxites as long as wide, with the aedeagus and paramere only slightly curved only at their distal fifth, whereas *D. symphyllia* has the ejaculatory apodeme nearly quadrate, being as long as 2.0 times its maximum width, and the aedeagus and paramere nearly straight.

It is important to mention that specimens of *D. symphyllia* from the Canal zone, Panama, and those of Guanacaste, Costa Rica, have the aedeagus nearly as long as paramere, whereas specimens from Cartago and Punta Arenas, Costa Rica, have the aedeagus much shorter than the paramere (Quate and Brown 2004), a difference that probably will be significant for separating both populations into species through future studies.

However, *D. noveloi* sp. nov. is distinguished from the other three species by the aedeagus strongly asymmetrical, basally broad with complex curvatures and apex long and lanceolate bent near the middle almost at a right angle so the tip is laterally directed, and by the paramere broad at base with a pyramidal short protuberance and a long sigmoidal branch which is longer and somewhat narrower than the aedeagus, with the apex directed laterally in the opposite direction to the aedeagus.

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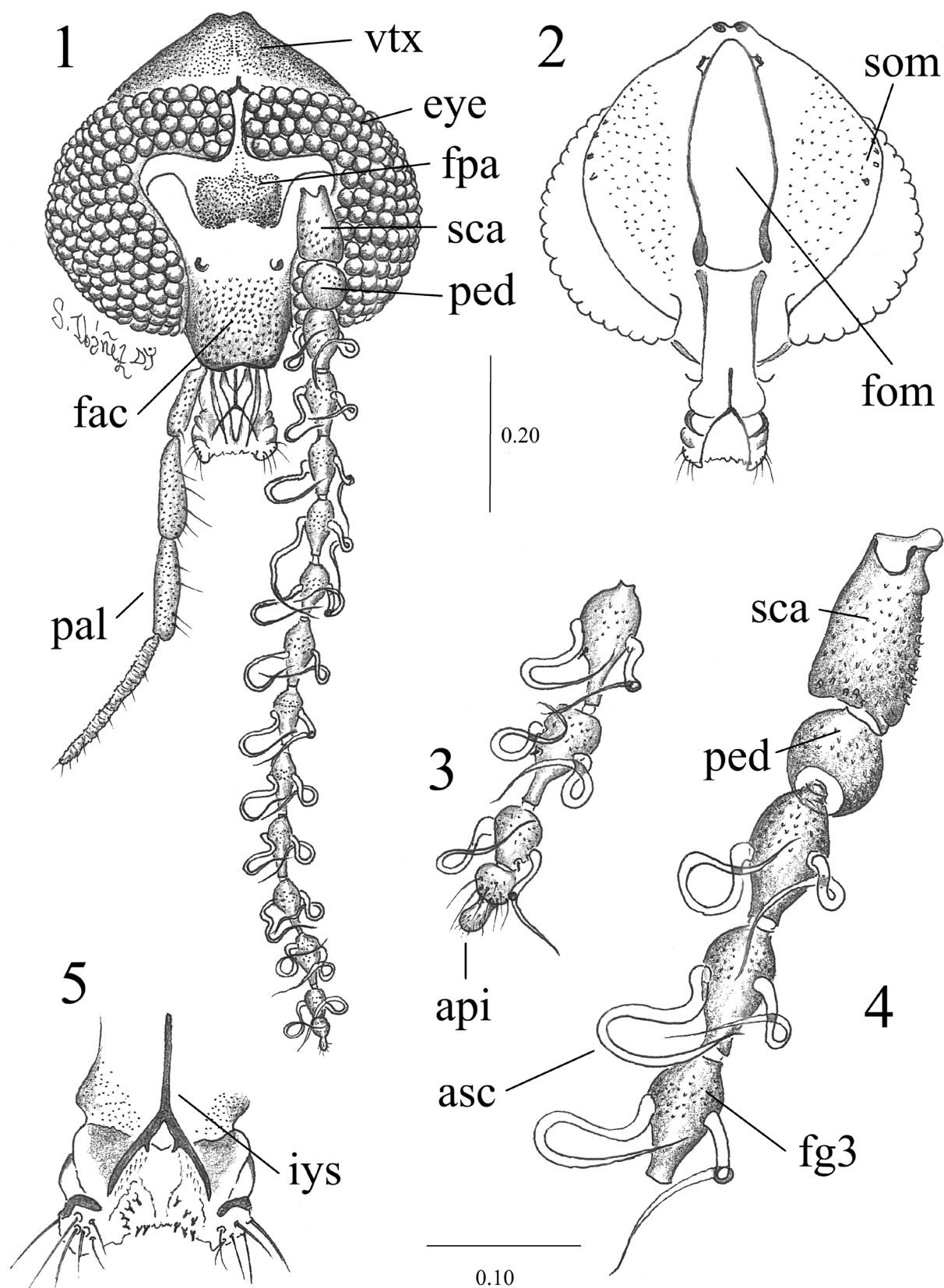
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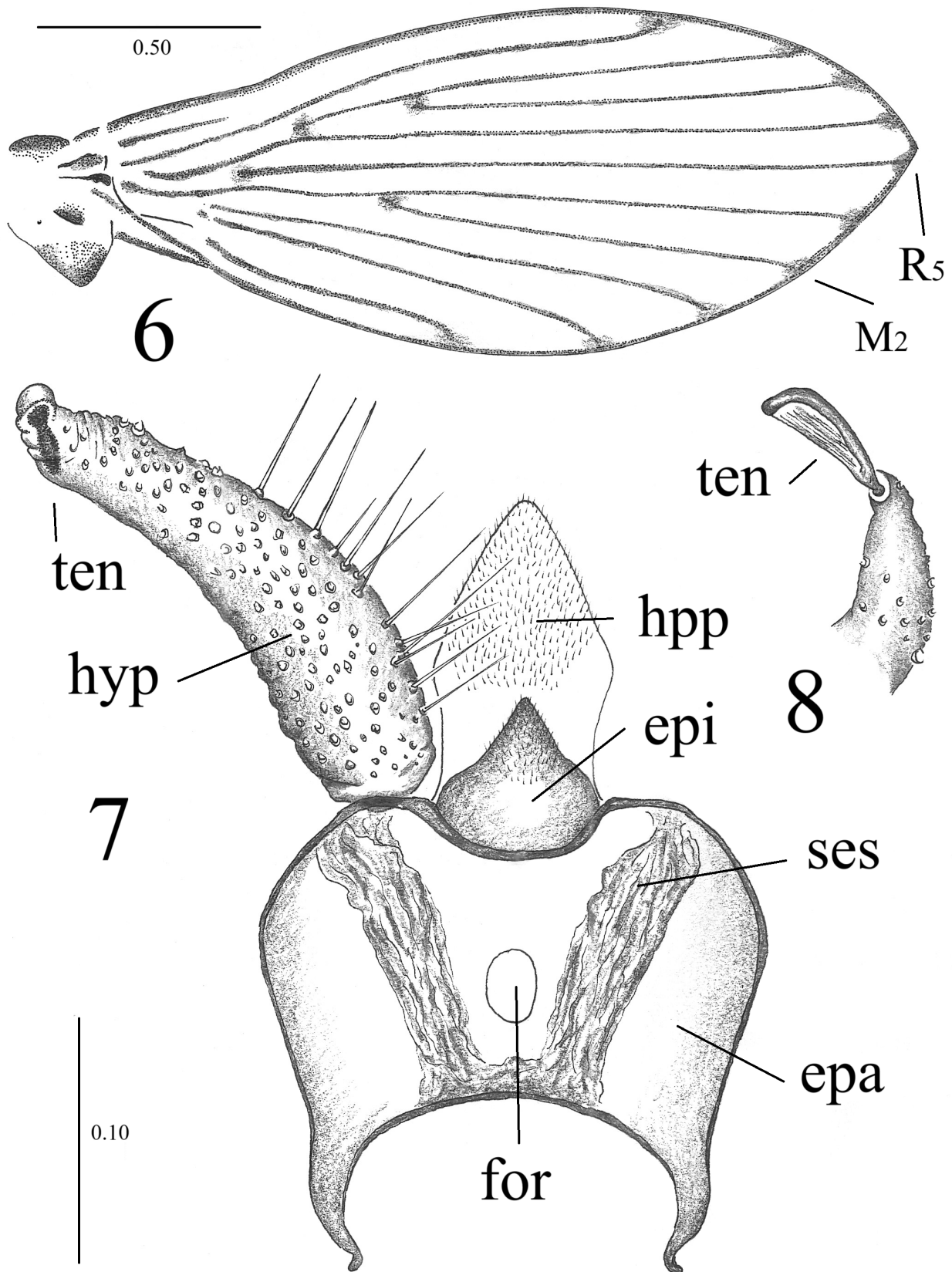
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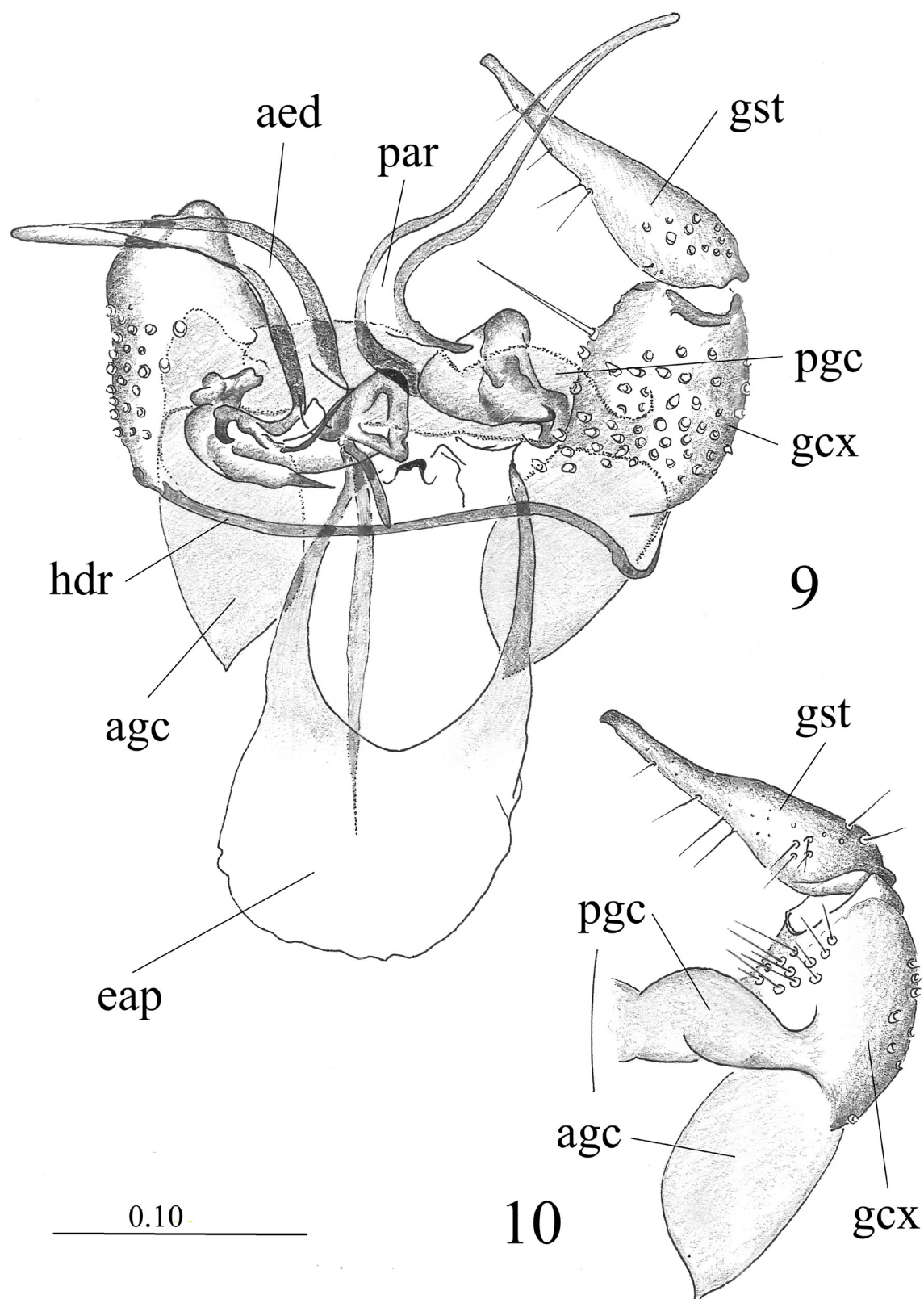
Aceptado: 12 de noviembre 2025



Figures 1- 5. *Didimioza noveloi* sp. nov., male. 1) Head, frontal view; left palpus and right antenna not drawn. 2) Head, posterior view. 3) Apical section of antenna. 4) Basal section of antenna. 5) Labium. Abbreviations: **api**: apiculus; **asc**: ascoid; **eye**: eye; **fac**: face; **fg3**: flagellomere 3; **fom**: foramen magnum; **fpa**: frontal patch of alveoli; **iys**: inverted Y-shaped sclerite; **pal**: palpus; **ped**: pedicel; **sca**: scape; **som**: supraorbital macrosetae **vtx**: vertex. Scales in millimeters.



Figures 6- 8. *Didimioza noveloi* sp. nov., male. 6) Wing. 7) Epandrium, hypopod, hypoproct, and epiproct. 8) Apex of hypopod showing the tenaculum in lateral view. Abbreviations: *epa*: epandrium; *epi*: epiproct; *for*: foramen; *hpp*: hypoproct; *hyp*: hypopod; M_2 : medial two vein; R_5 : radial five vein; *ses*: subepandrial sclerite; *ten*: tenaculum. Scales in millimeters.



Figures 9-10. *Didimioza noveloi* sp. nov., male. 9) Ventral view of terminalia. 10) partial view of terminalia showing the anterior and posterior gonocoxal apodemes. Abbreviations: **aed**: aedeagus; **agc**: anterior gonocoxal condyle; **eap**: ejaculatory apodeme; **gcx**: gonocoxite; **gst**: gonostyle; **hdr**: hypandrium; **par**: paramere; **pgc**: posterior gonocoxal condyle. Scale in millimeters.

Scolytinae y Platypodinae (Coleoptera: Curculionidae) asociados al ecosistema mangle en Tamaulipas, México

Scolytinae and Platypodinae (Coleoptera: Curculionidae) Associated with the Mangrove Ecosystem in Tamaulipas, Mexico

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RESUMEN

El presente estudio tuvo como objetivo registrar las especies de coleópteros de las subfamilias Scolytinae y Platypodinae asociadas al ecosistema de mangle en Tamaulipas, México. Las colectas se realizaron con trampas tipo botella durante el 2020 - 2021 en los ecosistemas de mangle de la “Laguna del Carpintero” en Tampico y del ejido “Lomas del Real” en Altamira, Tamaulipas. Se recolectaron 221 individuos, pertenecientes a ocho géneros y 14 especies. Para Scolytinae se registraron 13 especies de siete géneros. De la subfamilia Platypodinae, se registró a la especie *Teloplatypus ustulatus*. Las especies con mayor abundancia fueron *Microcorthylus minimus* e *Hypothenemus obscurus* con 54 y 52 individuos, lo cual representó el 47.96% del total recolectado. Las especies *Coccotrypes cyperi*, *Corthylus papulans*, *Hypothenemus eruditus*, *Hypothenemus interstitialis*, *Hypothenemus javanus*, *Hypothenemus obscurus* y *Xyleborus bispinatus* son nuevos registros para Tamaulipas. Este estudio aportó conocimiento sobre la composición de Scolytinae en los manglares de Tamaulipas.

Palabras clave: escarabajos ambrosiales, *Avicennia germinans*, *Conocarpus erectus*, *Laguncularia racemosa*, *Rhizophora mangle*.

ABSTRACT

The objective of this study is to record beetle species from the Scolytinae and Platypodinae subfamilies associated with mangrove trees in Tamaulipas, Mexico. Collections were carried out with bottle traps during 2020-2021 in the mangrove ecosystems of “Laguna del Carpintero” in Tampico and “Ejido Lomas del Real” in Altamira, Tamaulipas. A total of 221 individuals were collected, belonging to eight genera and 14 species. Thirteen species from seven genera were recorded for Scolytinae. From the Platypodinae subfamily, the species *Teloplatypus ustulatus* was recorded. The most abundant species were *Microcorthylus minimus* and *Hypothenemus obscurus*, with 54 and 52 individuals, representing 47.96% of the total collected. The species *Coccotrypes cyperi*, *Corthylus papulans*, *Hypothenemus eruditus*, *Hypothenemus interstitialis*, *Hypothenemus javanus*, *Hypothenemus obscurus*, and *Xyleborus bispinatus* are new records for Tamaulipas. This study contributed knowledge about the composition of Scolytinae in the mangroves of Tamaulipas.

Keywords: ambrosia beetles, *Avicennia germinans*, *Conocarpus erectus*, *Laguncularia racemosa*, *Rhizophora mangle*.

Los bosques de mangle están compuestos por especies de plantas únicas que forman la interfaz entre los ecosistemas marinos, terrestres, estuarinos y costeros en las regiones tropicales y subtropicales (Polidoro *et al.* 2010). Proporcionan múltiples servicios y funciones de valor para la sociedad, la flora y la fauna silvestre (Agráz-Hernández *et al.* 2006). Constituyen zonas de refugio y alimentación de fauna amenazada y en peligro de extinción, y de especies endémicas y migratorias (Osborne 2000, Agráz-Hernández *et al.* 2006), por lo anterior tienen un papel importante en el mantenimiento de la biodiversidad (Feller *et al.* 2010). Dentro de la diversidad de organismos asociados con los manglares encontramos a insectos barrenadores de las subfamilias Scolytinae y Platypodinae (Coleoptera: Curculionidae), estos escarabajos pasan la mayor parte de su vida al interior de los tejidos de las plantas, se asocian con la mayoría de los principales grupos de plantas terrestres en todas las regiones del mundo y colonizan casi todas las partes de

las plantas (Raffa *et al.* 2015, Falcon-Brindis *et al.* 2018). Los escarabajos de estas subfamilias desempeñan diversas e importantes funciones en los ecosistemas, contribuyen a la biodiversidad, participan en el reciclaje de nutrientes y la heterogeneidad (Burgos y Equihua 2007, Wood 2007, Raffa *et al.* 2015). La mayoría de las especies coexisten con sus plantas hospedantes donde se reproducen en ramas o troncos dañados, caídos o severamente debilitados por otros factores, así como en frutos y semillas ya sea antes o después de caer al suelo (Wood 2007, Atkinson 2017). Algunas especies afectan a plantas y productos vegetales valiosos, por lo que se consideran plagas forestales y agrícolas importantes, ya que ocasionan pérdidas socioeconómicas sustanciales (Grégoire *et al.* 2015, Raffa *et al.* 2015). A nivel mundial se conocen más de 6,000 especies de Scolytinae, la mayoría de las cuales se reproducen predominante o completamente en angiospermas, el 86% de los géneros se distribuyen en regiones tropicales o subtropicales (Kirkendall *et al.* 2015,

Atkinson 2017, Fernández-García *et al.* 2023). En México, se han registrado 954 especies de Scolytinae agrupadas en 91 géneros y 56 especies de Platypodinae agrupadas en 11 géneros (Atkinson 2025). Para el estado de Tamaulipas se tienen registradas un total de 65 especies de Scolytinae (Lázaro-Dzul *et al.* 2024, Atkinson 2025) y cuatro especies de Platypodinae, *Euplatypus parallelus* (Fabricius), *Euplatypus segnis* (Chapuis), *Megaplatypus discicollis* (Chapuis) y *Teloplatypus ustulatus* (Chapuis) (Atkinson 2025). Los trabajos relacionados con especies de Scolytinae y Platypodinae en México, se han enfocado principalmente en especies asociadas a bosques, selvas y otros agroecosistemas (Atkinson *et al.* 1986a, b, Atkinson y Equihua-Martínez 1986a, b, Noguera-Martínez y Atkinson 1990, Pérez-De La Cruz *et al.* 2009, 2016, Lázaro-Dzul *et al.* 2020). Para el caso del ecosistema de mangle en México, sobresalen los trabajos realizados por Gerónimo-Torres *et al.* (2015, 2021), Martínez-Zacarias *et al.* (2017) y Lázaro-Dzul *et al.* (2021, 2024) en donde se enlistan especies de Scolytinae y Platypodinae asociadas con este ecosistema. El objetivo de este trabajo es registrar las especies de coleópteros de las subfamilias Scolytinae y Platypodinae asociadas al ecosistema de mangle en Tamaulipas, México.

MATERIALES Y MÉTODOS

El presente estudio se llevó a cabo en dos sitios ubicados en el estado de Tamaulipas, México: el ejido “Lomas del Real” (22°33'04.8"N, 97°53'21.5"O; altitud de 10 msnm), en el municipio de Altamira, y la “Laguna del Carpintero” (22°14'19"N, 97°52'44"O; altitud de 3 msnm), localizada en el municipio de Tampico (Figura 1). Ambos sitios forman parte de un ecosistema de manglar caracterizado por la presencia de cuatro especies dominantes: *Avicennia germinans* (L.) (mangle negro), *Laguncularia racemosa* (L.) C. F. Gaertn. (mangle blanco), *Rhizophora mangle* (L.) (mangle rojo) y *Conocarpus erectus* (L.) (mangle botoncillo).

La vegetación adyacente a la “Laguna del Carpintero” se compone de especies hidrófilas como el mangle, vegetación secundaria de selva baja caducifolia y pastizales inducidos artificialmente en las áreas ganadas al cuerpo de agua, que sustentan grupos faunísticos de vertebrados (Rosas *et al.* 2016). El clima en este sitio es cálido subhúmedo, con lluvias en verano (INEGI 2009), y la temperatura media anual es de 25.1°C (Rosas *et al.* 2016). La vegetación asociada al manglar en el ejido “Lomas del Real”, se compone de selva baja caducifolia, vegetación halófila y vegetación de dunas costeras (Fierro-Cabo *et al.* 2022). El clima predominante en este sitio es cálido subhúmedo, con lluvias en verano, la temperatura media anual es mayor de 22°C (INEGI 2010, Fierro-Cabo *et al.* 2022).

Los muestreos se realizaron durante tres periodos: marzo-abril de 2020, octubre-diciembre de 2020 y octubre-diciembre de 2021. La captura de insectos se llevó a cabo mediante trampas de intercepción de vuelo, elaboradas a partir de botellas de PET de dos litros con una abertura lateral de 11 × 10 cm, siguiendo un diseño similar al propues-

to por Barrera *et al.* (2008). Como dispositivo liberador se emplearon frascos de polipropileno de 50 ml, los cuales se cargaron con 40 ml de solución atrayente por trampa. Se utilizaron etanol al 96 % y metanol al 99 % como atrayentes, y se incluyó un tratamiento testigo (agua+jabón). En cada sitio de muestreo se colocaron 12 trampas distribuidas en un transecto lineal, instaladas a 1.5 m de altura sobre los árboles y con una separación de 20 m entre cada trampa. Las trampas se revisaron una vez por semana y los insectos capturados se conservaron en alcohol al 70%. Estos fueron identificados utilizando las claves taxonómicas de Wood (2007), Burgos-Solorio y Hernández (2020) y Pérez Silva *et al.* (2021) y por comparación con ejemplares de esta subfamilia depositados en la Colección de Insectos del Colegio de Postgraduados, Campus Montecillo, México (CEAM). La corroboración de especies la realizó el Dr. T. H. Atkinson, de la Universidad de Texas, EE. UU. Los insectos se encuentran depositados en el Laboratorio de Entomología del Instituto Tecnológico de Ciudad Victoria, Tamaulipas (LEITCV).

Las fotografías de los insectos se tomaron en vista lateral utilizando una cámara réflex Canon EOS Rebel T5 acoplada a una base de microscopio óptico Carl Zeiss. Las imágenes fueron tomadas en capas a través del Software Entangle (Tether Tools, <https://www.tethertools.com/tethering-software/entangle/>), las capas se combinaron con Combine ZP (<https://combinezp.software.informer.com/>) y editadas con Gimp 2.8 (<https://gimp.software/2018/09/10/download-gimp-2-8-0/>).

Se determinó la abundancia total de individuos recolectados, así como la riqueza específica por localidad (Tampico y Altamira). Para cada especie se registró el número absoluto de individuos, su proporción relativa respecto al total recolectado (%) y su presencia o ausencia en cada ecosistema.

Con el fin de evaluar la efectividad de los atrayentes utilizados (etanol, metanol y testigo), se contabilizó el número de individuos, géneros y especies capturados en cada tipo de atrayente, por sitio de muestreo. Los resultados se expresaron en frecuencias absolutas y relativas (%), lo que permitió comparar la eficiencia de los atrayentes en ambas localidades.

Asimismo, las especies identificadas fueron clasificadas según sus hábitos alimenticios (fleófago, mielófago, xilomicetófago, fleófago-mielófago y espermatófago), con base en información reportada en literatura especializada. Finalmente, se generaron mapas de distribución de las especies registradas por primera vez en Tamaulipas, a partir de la información obtenida en este estudio y la disponible en la base de datos del Dr. T. H. Atkinson, con el propósito de ampliar el conocimiento sobre la distribución geográfica de dichas especies en el territorio mexicano.

RESULTADOS

En los 32 muestreos realizados se recolectaron un total de 221 individuos, pertenecientes a ocho géneros y 14

especies. Del total de especies registradas, el 64.2 % se encuentra reportado como asociado al ecosistema de mangle, mientras que el 35.7 % no presenta registros previos de asociación con este tipo de ecosistema (Cuadro 1).

En el manglar de la “Laguna del Carpintero” se identificaron 12 especies, con una abundancia total de 156 individuos, lo que representa el 70.58 % del total recolectado (Figuras 2 y 3). Las especies de Scolytinae más abundantes fueron *Microcorthylus minimus* Schedl con 54 individuos, *Hypothenemus obscurus* (F.) con 52 individuos y *Hypothenemus brunneus* (Hopkins) con 24 individuos, que en conjunto representaron el 58.82% de la colecta total; mientras que, de las especies *Coccotrypes cyperi* (Beeson), *Cryptocarenus seriatus* Eggers, *Pityophthorus* sp., y *Xyleborus bispinatus* Eichhoff, únicamente se recolectó un ejemplar. En este sitio también se recolectó un individuo de *Teloplatypus ustulatus* (Chapuis) de la subfamilia Platypodinae.

En el manglar del ejido “Lomas del Real” en Altamira se registraron seis especies y una abundancia de 65 individuos (29.41%) (Figuras 2 y 3). Las especies más abundantes para esta zona fueron *Hypothenemus eruditus* Westwood y *H. obscurus*, ambas con 18 individuos e *Hypothenemus crudiae* (Panzer) con 14 individuos que en conjunto representaron el 22.62% de la colecta total; la es-

pecie con menor abundancia fue *H. brunneus* (Hopkins) con dos ejemplares. El género con la mayor riqueza de especies fue *Hypothenemus* con seis especies en el manglar de la “Laguna del Carpintero” y cinco especies en el ejido “Lomas del Real”.

En el manglar de la “Laguna del Carpintero” en Tampico, en las trampas con etanol se registraron 127 individuos pertenecientes a cinco géneros y diez especies, que en conjunto representaron 81.4%. De los cuales cuatro géneros y nueve especies están incluidos en Scolytinae y un género y una especie está incluida en Platypodinae (*T. ustulatus*). En este mismo sitio en las trampas con metanol se registraron 29 individuos pertenecientes a tres géneros y cuatro especies, que en conjunto representaron 18.5% (Cuadro 2, Figura 4). En el manglar del ejido “Lomas del Real” en Altamira, en las trampas con etanol se registraron 32 individuos pertenecientes a dos géneros y seis especies. En este sitio en las trampas con metanol se registraron 29 individuos pertenecientes a dos géneros y cinco especies. En el tratamiento testigo se registraron cuatro individuos pertenecientes a un género y tres especies (Figura 4).

Se registraron siete especies de Scolytinae como nuevos registros para Tamaulipas: *Coccotrypes cyperi* Beeson: “Laguna del Carpintero”, 22-XI-2021, 22°14'19"N

Cuadro 1. Especies de Scolytinae y Platypodinae asociadas al ecosistema de mangle en Tampico y Altamira, Tamaulipas, México.

Especies	Número de individuos			
	Tampico	Altamira	HA	Total
Scolytinae				
<i>Coccotrypes cyperi</i> (Beeson) ²	1	0	es	1
<i>Corthylus minutissimus</i> Schedl ¹	18	0	xm	18
<i>Corthylus papulans</i> Eichhoff ^{1,2}	2	0	xm	2
<i>Cryptocarenus seriatus</i> Eggers ¹	1	4	m	5
<i>Hypothenemus brunneus</i> (Hopkins) ¹	24	2	f-m	26
<i>Hypothenemus crudiae</i> (Panzer) ¹	0	14	f-m	14
<i>Hypothenemus eruditus</i> Westwood ^{1,2}	15	18	f-m	33
<i>Hypothenemus interstitialis</i> (Hopkins) ^{1,2}	0	9	f-m	9
<i>Hypothenemus javanus</i> (Eggers) ^{1,2}	4	0	m	4
<i>Hypothenemus obscurus</i> (F.) ²	34	18	f-m	52
<i>Microcorthylus minimus</i> Schedl ¹	54	0	xm	54
<i>Pityophthorus</i> sp.	1	0	f	1
<i>Xyleborus bispinatus</i> Eichhoff ²	1	0	xm	1
Platypodinae				
<i>Teloplatypus ustulatus</i> (Chapuis)	1	0	xm	1
Total de individuos	156	65		221
Total de especies	12	6		14

¹Especies reportadas previamente como asociadas al ecosistema de mangle. ²Nuevos registros para Tamaulipas. HA= hábito alimenticio siguiendo la terminología de Wood (1982): f= fleófago; es= espermatófago; m= mielófago; xm= xilomicetófago.

Cuadro 2. Especies de Scolytinae y Platypodinae recolectadas por tipo de atrayente en el ecosistema de mangle en Tampico y Altamira, Tamaulipas, México.

Species	Tampico			Altamira		
	Atrayentes					
Scolytinae	E	M	T	E	M	T
<i>Coccotrypes cyperi</i> (Beeson)	1	0	0	0	0	0
<i>Corthylus minutissimus</i> Schdel	18	0	0	0	0	0
<i>Corthylus papulans</i> Eichhoff	2	0	0	0	0	0
<i>Cryptocarenus seriatus</i> Eggers	1	0	0	2	1	1
<i>Hypothenemus brunneus</i> (Hopkins)	14	10	0	2	0	0
<i>Hypothenemus crudiae</i> (Panzer)	0	0	0	7	6	1
<i>Hypothenemus eruditus</i> Westwood	15	0	0	10	6	2
<i>Hypothenemus interstitialis</i> (Hopkins)	0	0	0	4	5	0
<i>Hypothenemus javanus</i> (Eggers)	4	0	0	0	0	0
<i>Hypothenemus obscurus</i> (F.)	17	17	0	7	11	0
<i>Microcorthylus minimus</i> Schedl	54	0	0	0	0	0
<i>Pityophthorus</i> sp.	0	1	0	0	0	0
<i>Xyleborus bispinatus</i> Eichhoff	0	1	0	0	0	0
Platypodinae						
<i>Teloplatypus stulatus</i> (Chapuis)	1	0	0	0	0	0
Total	127	29	0	32	29	4

E: Etanol; M: Metanol; T: Testigo.

97°52'44"O, 3msnm, trampa con etanol; *Corthylus papulans* Eichhoff: "Laguna del Carpintero", 26-IV-2020, 22°14'19"N 97°52'44"O, 3msnm, trampa con etanol; *Hypothenemus eruditus* Westwood: "Laguna del Carpintero", 14-III-2021, 22°14'19"N 97°52'44"O, 3msnm, trampa con etanol; "Ejido Lomas del Real", 11-X-2020, 22°33'4.8"N, 97°53'21.5"O, 10 msnm, trampa con etanol; *Hypothenemus javanus* Eggers: "Laguna del Carpintero", 14-XII-2021, 22°14'19"N 97°52'44"O, 3msnm, trampa con etanol; *Hypothenemus interstitialis* Hopkins: "Ejido Lomas del Real", 01-XI-2020, 22°33'4.8"N, 97°53'21.5"O, 10 msnm, trampa con etanol; *Hypothenemus obscurus* F.: "Laguna del Carpintero", 14-III-2021, 22°14'19"N 97°52'44"O, 3msnm, trampa con etanol y *Xyleborus bispinatus* Eichhoff: "Laguna del Carpintero", 07-XI-2021, 22°14'19"N 97°52'44"O, 3msnm, trampa con metanol (Figura 5).

Con relación a los hábitos alimenticios para las especies detectadas destacaron los hábitos fleófago-mielófago (60.63%), xilomicetófago (34.38%), mielófago (4.07%), fleófago (0.45%) y espermátófago (0.45%).

DISCUSIÓN

Las diferencias en la riqueza y abundancia de especies en ambos sitios de estudio pueden atribuirse a la diversidad

vegetal circundante, disponibilidad de alimento, competidores, temperatura y humedad, dado que, dichos factores pueden influir en la distribución y abundancia de las especies de Scolytinae y Platypodinae en un sitio determinado (Rudinsky 1962, Wood 1982, Pérez-De la Cruz *et al.* 2009). En el caso del ecosistema de mangle en Tampico, además de la presencia de *A. germinans*, *L. racemosa*, *R. mangle* y *C. erectus*, la vegetación circundante se compone de especies de las familias Malvaceae, Fabaceae, Aracaceae, Bignoniaceae, Magnoliopsida, Oleaceae, entre otras ((De la Cruz y Tello 2012); mientras que, en Altamira el sitio se compone casi en su totalidad de especies de mangle, y pocas especies características de dunas costeras (Fierro-Cabo *et al.* 2022), lo anterior contribuye a explicar la mayor abundancia y riqueza de especies presente en Tampico, derivado de la mayor riqueza de especies vegetales en este último sitio; con respecto a las temperaturas predominantes, en el manglar de Tampico la temperatura promedio anual es de 25°C (Rosas *et al.* 2016) y en el manglar de Altamira de 22°C (INEGI 2010, Fierro-Cabo *et al.* 2022), si bien las diferencias en la temperatura promedio son mínimas es posible que el clima ligeramente más cálido en Tampico haya favorecido la mayor abundancia de Scolytinae.

Por otra parte, la riqueza de especies del género *Hypothenemus* que se encontró en este estudio coincide con lo

reportado en manglares de Tabasco (Gerónimo-Torres *et al.* 2015, 2021), y también con lo documentado para selvas y agroecosistemas en los estados de Tabasco y Michoacán (Pérez-De La Cruz *et al.* 2009, 2015, 2016, Falcon-Brindis *et al.* 2018, Lázaro-Dzul *et al.* 2020). La mayor riqueza de *Hypothenemus* en ambos sitios de estudio puede atribuirse a que, en áreas tropicales y subtropicales, es uno de los géneros con mayor número de especies dentro de la subfamilia Scolytinae (Wood 1986). Además, muchas de las especies en este género son altamente polífagas y poseen una alta tasa reproductiva (Wood y Bright 1992, Pérez-De La Cruz *et al.* 2009). Como ejemplo, se puede indicar que de *H. obscurus*, que fue una de las especies más abundantes, se han registrado alrededor de 30 plantas hospedantes de 13 familias botánicas distintas (Atkinson 2025). Se reproduce en tallos, frutos, semillas y en algunas ocasiones en ramas secas de sus hospedantes (Beardsley 1990, Wood 2007, Vega *et al.* 2015).

Para el caso de *M. minimus*, otra de las especies abundantes, se reporta como una especie polífaga (Atkinson 2025). Esta especie ha sido reportada previamente asociada al ecosistema de mangle en Tabasco (Gerónimo-Torres *et al.* 2015, 2021) y en Tamaulipas (Lázaro-Dzul *et al.* 2021). En este estudio la mayor abundancia de *M. minimus* se observó en el ecosistema de mangle ubicado en Tampico, en este sitio, además de los árboles de mangle, existe una mayor riqueza de especies vegetales, resaltando árboles de la familia Arecaceae (palmeras), Bignoniaceae (palo de rosa), Magnoliopsida (sauce llorón), Fabaceae (guamúchil, frambayán, lluvia de oro, guaje) y Oleaceae (fresno) (De la Cruz y Tello 2012), lo cual pudo favorecer la mayor disponibilidad de hospedantes y recursos alimenticios para la especie. Cabe mencionar que la mayor abundancia de *M. minimus* ocurrió en los meses de marzo y abril, periodo en el cual la precipitación fue prácticamente nula, estos resultados coinciden con lo documentado por Dorval y Peres Filho (2001) quienes reportaron que la abundancia de algunas especies de Scolytinae se incrementa en el periodo de sequía y se reduce durante el periodo de lluvia.

De igual manera las especies *C. seriatus*, *H. brunneus* y *H. crudiae*, se han reportado asociadas a manglares en Tabasco (Gerónimo-Torres *et al.* 2015, 2021) lo cual coincide con lo encontrado en este estudio. Además, no omitimos mencionar que estas especies son ampliamente polífagas (Atkinson y Riley 2013).

Con relación a las especies que representaron nuevos registros de distribución para Tamaulipas, cabe mencionar que todas ellas ya se han citado presentes en México (Figura 6). La especie *C. cyperi*, se ha reportado en Chiapas (Equihua-Martínez 1992), Tabasco (Pérez-De La Cruz *et al.* 2009, 2016, Álvarez-Ramón *et al.* 2025) y Veracruz (Atkinson 2025). Se reproduce en semillas grandes de frutos caídos (Wood 2007). Se reporta asociada a plantas de 21 familias distintas (Atkinson 2025).

De *Corthylus papulans* se tienen reportes en Campeche (Estrada-Valencia y Atkinson 1988), Chiapas, Guerrero

(Atkinson 2025), Jalisco (Atkinson y Equihua-Martínez 1986a), Michoacán (Lázaro-Dzul *et al.* 2020), Morelos (Atkinson *et al.* 1986a, b), Oaxaca (Wood y Bright 1992), Nayarit (Wood 2007), Tabasco (Pérez-De La Cruz *et al.* 2009, 2016, Gerónimo-Torres *et al.* 2015, 2021) y Veracruz (Noguera-Martínez y Atkinson 1990). Es una especie de hábito ambrosial o xilomicetófaga, polífaga, generalmente se desarrolla en material de hospedantes derribados o caídos (Atkinson *et al.* 1986a, b). Está asociada a plantas de 17 familias distintas (Atkinson 2025). Los resultados encontrados coinciden con lo citado por Gerónimo-Torres *et al.* (2015, 2021), quienes también reportan a esta especie asociada al ecosistema de mangle compuesto por *Avicennia germinans*, *Rhizophora mangle* y *Laguncularia racemosa*.

Hypothenemus eruditus, se ha reportado en Baja California Sur, Guerrero, Nuevo León, Oaxaca, Yucatán (Atkinson 2025), Campeche, Nayarit (Estrada-Valencia y Atkinson 1988), Chiapas (Wood 1982, Atkinson y Equihua-Martínez 1985), Colima, Puebla, San Luis Potosí (Wood 1982), Jalisco (Atkinson y Equihua-Martínez 1986a), Michoacán (Lázaro-Dzul *et al.* 2020), Morelos (Atkinson *et al.* 1986a), Quintana Roo (Equihua-Martínez 1988), Sinaloa (Kirkendall 1984), Tabasco (Pérez-De La Cruz *et al.* 2009, 2016, Gerónimo-Torres *et al.* 2015, 2021) y Veracruz (Noguera-Martínez y Atkinson 1990). Esta especie adapta sus hábitos al material hospedante disponible, se puede reproducir en corteza, médula, madera, frutos, semillas, en tallos de plantas herbáceas, e incluso en productos manufacturados (Browne 1961, Wood 1982, Atkinson *et al.* 1986a). Es una especie ampliamente polífaga, asociada a plantas de 57 familias distintas (Atkinson 2025). En mangle se reporta asociada al ecosistema de mangle compuesto por *A. germinans*, *R. mangle* y *L. racemosa* (Gerónimo-Torres *et al.* 2015, 2021).

Hypothenemus interstitialis, se ha reportado en Campeche (Estrada-Valencia y Atkinson 1988), Chiapas, Nayarit (Wood 1982), Guerrero, Oaxaca, Quintana Roo, Yucatán (Atkinson 2025), Jalisco (Atkinson y Equihua-Martínez 1986a), Tabasco (Pérez-De La Cruz *et al.* 2009, 2016, Gerónimo-Torres *et al.* 2015, 2021) y Veracruz (Noguera-Martínez y Atkinson 1990). Se reproduce en ramas pequeñas, médula y xilema de sus hospedantes (Wood 2007). Está asociada a plantas de 33 familias distintas (Atkinson 2025). También se reporta asociada al ecosistema de mangle compuesto por *A. germinans*, *R. mangle* y *racemosa* (Gerónimo-Torres *et al.* 2015, 2021).

Hypothenemus javanus, se ha reportado en Jalisco y Veracruz (Wood 1982, Atkinson y Equihua-Martínez 1985). Se reproduce en tallos de lianas y ramas pequeñas de árboles y arbustos (Wood 2007). Está asociada a plantas de 17 familias distintas, en mangle se reporta asociada con árboles de *Conocarpus erectus* y *R. mangle* (Atkinson 2025).

H. obscurus, se ha reportado en Campeche, Chiapas, Colima, Guerrero, Jalisco, Nayarit, Oaxaca, Sinaloa, Veracruz, Yucatán (Atkinson 2025), Michoacán (Lázaro-Dzul *et al.* 2020) y Sonora (Atkinson 2019). Dentro del género *Hypo-*

thenemus, es la segunda especie más dañina desde el punto de vista económico después de la broca del café, *H. hampei* (Vega *et al.* 2015).

Xyleborus bispinatus, se reporta en Campeche (Atkinson *et al.* 2013), Michoacán (García-Guevara *et al.* 2018), Chiapas, Puebla, Oaxaca, Querétaro, Quintana Roo (Pérez-Silva *et al.* 2015, 2021), Jalisco, Sinaloa, Tabasco y Veracruz (Atkinson 2025). Es una especie asociada a bosques tropicales del Golfo de México (Pérez-Silva *et al.* 2021). Es de hábito ambrosial, se reporta asociada con plantas de las familias Euphorbiaceae, Fagaceae, Lauraceae, Lecythidaceae, Leguminosae, Meliaceae, Urticaceae (Atkinson 2025). Del género *Pityophthorus* se colectó un solo ejemplar, si bien la mayoría de las especies dentro de este género se reproducen en coníferas, muchas especies neotropicales se reproducen en árboles de hoja ancha, arbustos y enredaderas (Atkinson 1993), lo cual explicaría la presencia de este ejemplar, ya que en la zona de manglar de Tampico, además de las especies de mangle, la vegetación aledaña se compone de árboles de hoja ancha como palo de rosa (Bignoniaceae), sauce llorón (Magnoliopsida), guamúchil, framboyán, lluvia de oro, guaje (Fabaceae) y fresno (Oleaceae) (De la Cruz y Tello 2012).

Teloplatus ustulatus, es un escarabajo barrenador ambrosial de la subfamilia Platypodinae, generalmente infesta troncos de árboles derribados en proceso de fermentación (Atkinson 2017, Burgos-Solorio y Hernández 2020). Esta especie se registra asociada con plantas de la familia Fabaceae (Leguminosae) de los géneros *Acacia*, *Enterolobium*, *Erythrina*, *Inga*, *Lonchocarpus*, *Lysiloma*, *Pithecellobium* y con plantas de la familia Urticaceae del género *Cecropia* (Burgos-Solorio y Hernández 2020, Atkinson 2025), sin embargo, no se ha reportado asociada con el ecosistema de mangle. En este estudio, *T. ustulatus*, se recolectó en una trampa con etanol, al respecto se ha documentado que, los platipódinos al igual que los escolitinos guían su vuelo hacia áreas de liberación de oleorresinas volátiles y alcoholes, como por ejemplo el alcohol etílico, que es liberado por microorganismos en el tejido leñoso en descomposición (Cade *et al.* 1970) y plantas estresadas (Kimmerer y Kozlowski 1982), por lo cual, este compuesto funciona como atrayente para una gran variedad de escarabajos ambrosiales y descortezadores (Byers 1992, Ghandi *et al.* 2010, Kelsey *et al.* 2013, Atkinson 2017).

Con relación a los hábitos de alimentación de las especies, en este estudio se observó que el mayor porcentaje correspondió a las especies fleófago-mielófago (60.63%) y xilomicetófago (34.38%), las especies mielófagas que se reproducen en médulas de ramitas encuentran el material más seco y aparentemente sin defensas activas de las plantas, lo que facilita la colonización, en este estudio seis especies del género *Hypothenemus* y la especie *C. seriatus* compartieron este hábito de alimentación; por otra parte, las especies xilomicetófagas o ambrosiales, de manera general requieren mayor humedad ambiental para sus hongos ectosimbiontes de los cuales se alimentan, por lo cual son más

diversas en ecosistemas con mayor precipitación (Atkinson y Equihua 1986a, 1986b, Noguera-Martínez y Atkinson 1990, Atkinson 2019). En cambio, las especies fleófagas son altamente restringidas, la mayoría de ellas se desarrollan en plantas con resinas o látex, lo cual requiere adaptaciones particulares para superar estas defensas (Atkinson 2019).

Es importante mencionar que, con las trampas de etanol se recolectó el mayor número de especies de Scolytinae, lo cual concuerda con lo señalado en otros estudios, donde también se ha comprobado la eficiencia de este volátil para la atracción de especies de Scolytinae en diferentes ecosistemas en México (Pérez-De La Cruz *et al.* 2009, 2016, Lázaro-Dzul *et al.* 2020, Gerónimo-Torres *et al.* 2015). Sin embargo, también existen otros métodos, como son las trampas de luz, plantas trampa y la recolecta directa, los cuales han demostrado ser complementarios para ampliar el conocimiento de estos insectos en una región determinada (Pérez-De la Cruz *et al.* 2009, Gerónimo-Torres *et al.* 2021).

Con este estudio se aportó conocimiento acerca de las especies de la subfamilia Scolytinae y Platypodinae asociadas al ecosistema de mangle en Tamaulipas y se incrementó el registro de especies de 63 a 72 Scolytinae para el estado. Los mapas de distribución de las especies que representaron nuevos registros para Tamaulipas aportan una visión más integral de la ubicación geográfica de las especies en México. Se sugiere la realización de otros estudios para conocer la fluctuación poblacional de las especies más abundantes para ampliar el conocimiento de las interacciones que tienen con los árboles de mangle.

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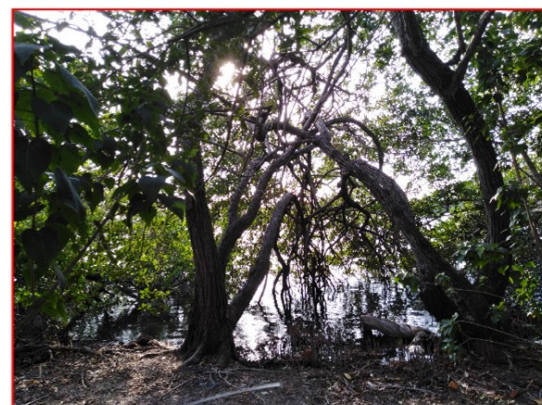
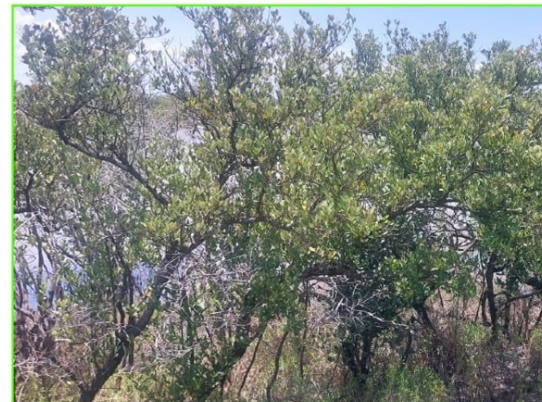
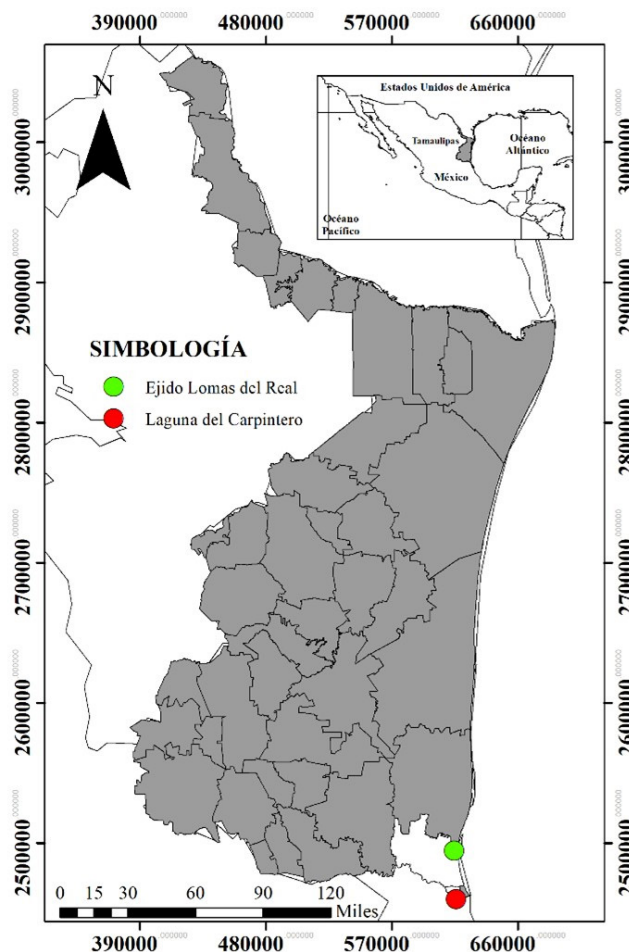
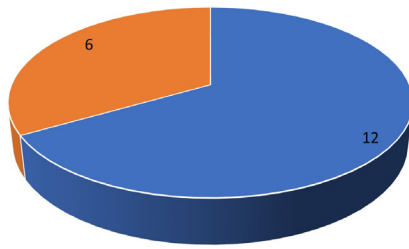
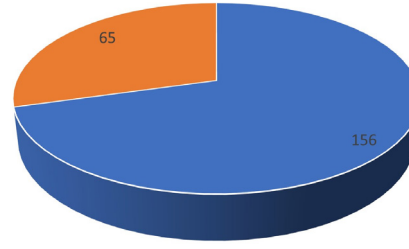


Figura 1. Localización de los sitios de muestreo de Scolytinae en Tamaulipas, México.



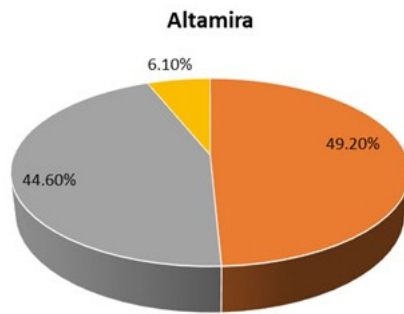
■ Tampico ■ Altamira

Figura 2. Numero total de especies de Scolytinae y Platypodinae recolectadas en el ecosistema de mangle en Altamira y Tampico, Tamaulipas, México.

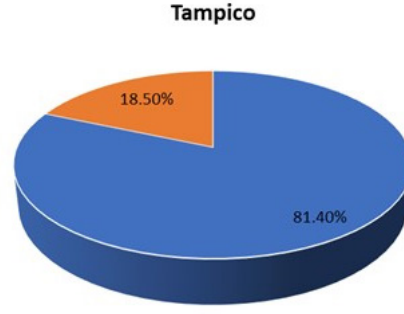


■ Tampico ■ Altamira

Figura 3. Abundancia de Scolytinae y Platypodinae recolectadas en el ecosistema de mangle en Altamira y Tampico, Tamaulipas, México.



■ Etanol ■ Metanol ■ Testigo



■ Etanol ■ Metanol

Figura 4. Numero total de especies de Scolytinae y Platypodinae recolectadas por tipo de atrayente en el ecosistema de mangle en Altamira y Tampico, Tamaulipas, México.

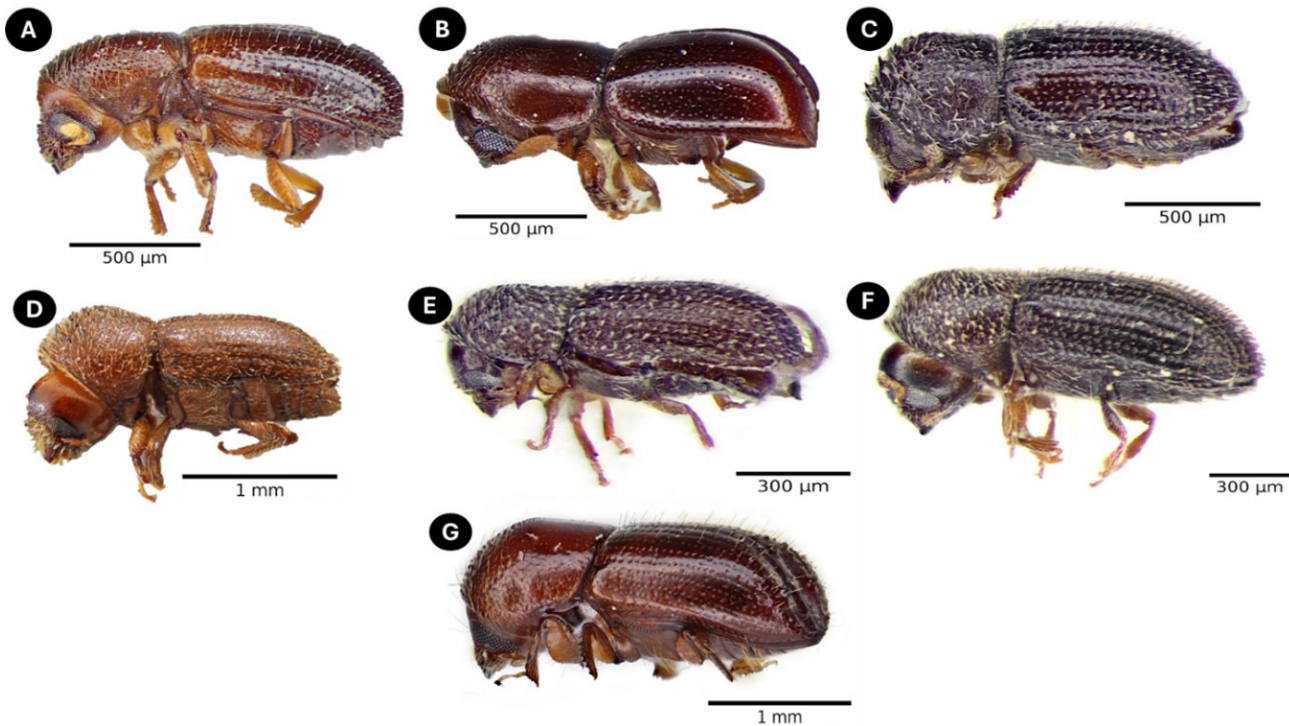


Figura 5. Especies de Scolytinae asociadas con el ecosistema de mangle en Altamira y Tampico, Tamaulipas, México. A) *Coccotrypes cyperi* Beeson, B) *Corthylus papulans* Eichhoff, C) *Hypothenemus eruditus* Westwood, D) *Hypothenemus javanus* Eggers, E) *Hypothenemus interstitialis* Hopkins, F) *Hypothenemus obscurus* F. y G) *Xyleborus bispinatus* Eichhoff.

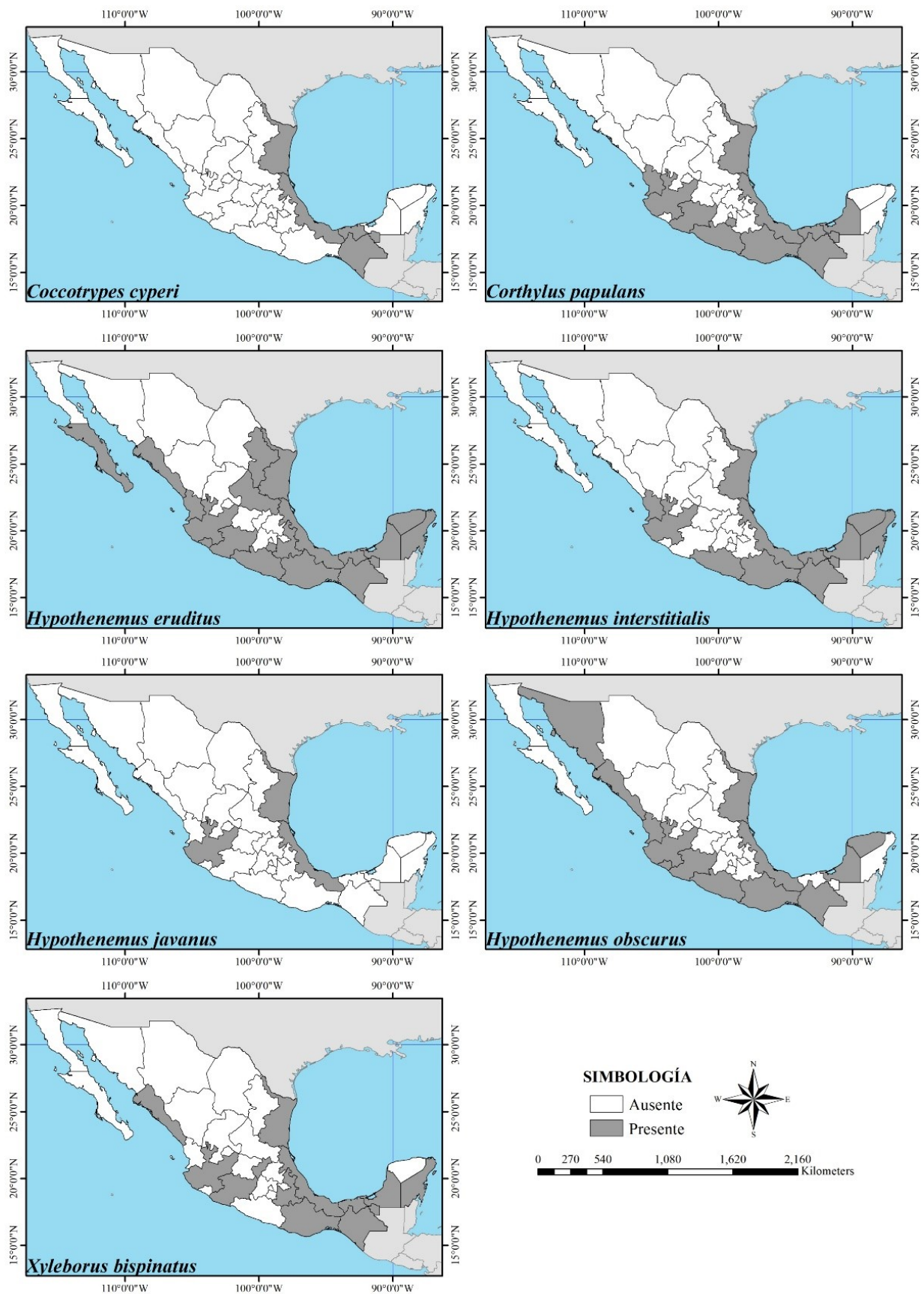


Figura 6. Distribución geográfica de las especies de Scolytinae que contempla los registros previos por el Dr. T. H. Atkinson, y los nuevos registros para Tamaulipas, México. Escala: 1 cm = 256 Km.

Description of *Femuros minimum* n. sp. (Hymenoptera: Cynipidae) collected on *Quercus glaucooides* Mart. & Gal. (Fagaceae) in Puebla, Mexico

Descripción de *Femuros minimum* n. sp. (Hymenoptera Cynipidae) colectado *Quercus glaucooides* Mart. & Gal. (Fagaceae) en Puebla (México)

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RESUMEN

Se describe una nueva especie en México de avispa agalladora de los encinos con base a su generación asexual, *Femuros minimum* Bravo-Cuautle, Cuesta-Porta & Pujade-Villar n. sp., que induce pequeñas agallas en las yemas apicales de *Quercus glaucooides* Mart. & Gal. (Sección *Quercus*). Se comenta su distribución, datos biológicos y se ilustran los caracteres morfológicos distintivos. Finalmente, se exponen los caracteres diagnósticos para diferenciarlos de las especies cercanas.

PALABRAS CLAVE: nueva especie, avispa agalladora, Cynipini, Fagaceae, taxonomía, morfología, distribución, biología.

ABSTRACT

A new species of oak gall wasp from Mexico is described based on its asexual generation: *Femuros minimum* Bravo-Cuautle, Cuesta-Porta & Pujade-Villar n. sp., which induces small galls on the apical buds of *Quercus glaucooides* (section *Quercus*). The distribution and biological data of the new species are discussed, and its distinctive morphological characters are illustrated. Finally, the characters that differentiate it from closely related species are presented.

KEY WORDS: new species, oak gallwasp, Cynipini, Fagaceae, taxonomy, morphology, distribution, biology.

Mexico is one of the main centers of oak diversification and endemism, hosting approximately 250 described species (Romero-Rangel *et al.* 2015), 86 of which are endemic (Valencia 2004). The family Cynipidae (Hymenoptera) is notable for including phytophagous wasps that induce galls, belonging to the tribe Cynipini, which primarily attack Fagaceae, especially the genus *Quercus*. Due to the close relationship between oaks and cynipids, and the high number of species of *Quercus* in Mexico, it is believed that diversity of these gall wasps in the country is also high. Until 2022, 205 cynipid species had been recorded in Mexico (Martínez-Romero *et al.* 2022).

The genus *Femuros* was described by Kinsey (1937a) to include two Mexican species that exhibited a posterior femur widened on the ventral margin near the distal area and constricted toward the tibia (*F. repandae* Kinsey, 1937, and *F. ruidum* Kinsey, 1937). Later, Kinsey (1937b) described five additional Mexican species: *F. lusum* Kinsey, 1937, *F. perfectum* Kinsey, 1937, *F. geniale* Kinsey, 1937, *F. integrum* Kinsey, 1937, and *F. ocri* Kinsey, 1937. Melika & Abrahamson (2002) synonymized *Femuros* under *Andricus*. However, Pujade-Villar & Ferrer-Suay (2015)

reinstated the genus, adding another distinguishing characteristic—the presence of an internal carina on the hind tibia. Finally, Cuesta-Porta *et al.* (2025) reviewed *Femuros* and transferred four of Kinsey's species to the genus *Andricus* (*A. lusum*, *A. perfectum*, *A. geniale*, and *A. integrum*), while including a new combination under *Femuros*, *F. bracteatus* Weld, 1944.

Since the latest revision (Cuesta-Porta *et al.* 2025), *Femuros* has been characterized by a nearly smooth or smooth lower head, no radiating carina from the clypeus (or a very short one), absence of a malar groove, a visible occipital carina behind the ocelli, a transcutal articulation, a posterior femur widened on the ventral margin near the distal end, and an internal carina on the hind tibia.

Currently, four species are known, all of them from their asexual form: three species described from Mexico (*F. repandae*, *F. ocri*, and *F. ruidum*) and one from the United States (*F. bracteatus*). The galls of this genus are round, partially covered by small bracts, with parenchymatous tissue connecting to the larval chamber, and have a lignified outer layer. There is some variation in gall size among species, making it difficult to distinguish them without

the emergence of the adult wasp. Despite these advances, the diversity of *Femuros* in Mexico remains insufficiently known, and additional undescribed species are likely to occur.

This study describes a new *Femuros* species from Mexico that induces small galls on the apical parts of the branches of an endemic Mexican oak species, *Quercus glaucoides* Mart. & Gal.

MATERIAL AND METHODS

Asexual females were obtained from galls collected on *Q. glaucoides* (section *Quercus*), located in the Botanical Garden of Puebla, Mexico. The galls were preserved in emergence vials, and the adults were stored in 70% ethanol until morphological analysis.

The terminology used to describe morphological structures follows the works of Liljeblad *et al.* (2008), Melika (2006), and Melika *et al.* (2010). Wing venation abbreviations follow Ronquist & Nordlander (1989), and cuticular surface terminology follows Harris (1979).

The measurements and abbreviations used here include: F1–F12, referring to the first and subsequent flagellomeres; POL (postocellar distance), the distance between the inner margins of the posterior ocelli; OOL (ocellar-ocular distance), the distance from the outer edge of a posterior ocellus to the inner margin of the compound eye; LOL (lateral-frontal ocelli distance), the distance between the lateral and frontal ocelli. The width of the radial cell of the forewing was measured from the wing margin to the Rs vein.

SEM images were taken by the first author using an environmental scanning electron microscope (FEI Quanta 200 ESEM) without any coating (gold or carbon). Gall images were taken by the first author using a Canon PowerShot SX510 HS digital camera directly through a binocular magnifier. Adult images were captured by the first author using a digital camera mounted on a Carl Zeiss III microscope.

The type material is deposited in the University of Barcelona (UB, J. Pujade-Villar collection).

RESULTS

Femuros minimum Bravo-Cuautle, Cuesta-Porta & Pujade-Villar, **sp. nov.**
(Figs 1–3)

<http://zoobank.org/B4B458B5-A6FE-49A3-B84B-64B34499A903>

Type material. HOLOTYPE asexual female deposited in the JP-V (UB) collection, with the following labels: MEX, Jardín Botánico de Puebla, *Q. glaucoides*, (28.x.2024) 1.i.2025 (white label); Holotype *Femuros minimum* Bravo-Cuautle, Cuesta-Porta & Pujade-Villar n. sp., desig. JP-V 2025 (red label). PARATYPES 5♂ with the same labels as holotype except for the dates of collection and they were manually extracted (ext.): (28.x.2024) ext. 1.iii.2025: 1♂; (28.i.2025) extr. 1.iii.2025: 4♂.

Additional material. MEX Jardín Botánico de Puebla, *Q. glaucoides*, (28.i.2025) extr. 1.iii.2025: 1♂ dissected for

SEM.

Etymology. The name *minimum* refers to the small size of the galls.

Diagnosis. *Femuros minimum* n. sp. differs from other species of *Femuros* by the number of flagellomeres (12 in the new species, 11 with a very inconspicuous suture in the other species) and by the length of the malar space (3.0x in the new species, around 2.1x in the others). The new species is morphologically closely related to *F. ocrici*, but it differs in the following characters: ventral spine of hypopygium 5.0x as long as broad in ventral view (3.5–4.0x in *F. ocrici*), body color dark brown (reddish-brown in *F. ocrici*); anterior parallel lines and parapsidal lines black (without black marks in *F. ocrici*); and ventral margin of clypeus straight (rounded in *F. ocrici*). The new species also differs from all *Femuros* species by the small size of its gall, around 4.0 mm in diameter (averaging 8.0–15 mm in the other species).

Description. Asexual female. Body length 2.6–3.1 mm (n=6).

Color (Fig. 3e). Head, mesosoma, and metasoma chestnut brown with some darker areas on anterior parallel lines and parapsidal lines; mesopleura and scutellar foveae black, antennae with darker flagellomeres, legs chestnut brown, coxae lighter, third femur and tibia black. Wing hyaline, veins brown.

Head (Figs. 1a–c). Transversally ovate, broadest part below toruli, broader than high and narrower than mesosoma in frontal view, with short dense white setae; 1.4x as broad as high in anterior view and head 2.1x as broad as broad as long in dorsal view. Gena alutaceous to smooth, slightly broadened behind eye, as broad as cross diameter of eye, measured in lateral view along transfacial line. Malar space weakly alutaceous to smooth, without striae radiating from clypeus, malar sulcus absent; eye 3.0x as high as length of malar space. Inner margins of eyes parallel. POL 3.0x as long as OOL, OOL shorter than the diameter of the lateral ocellus and shorter than LOL, all ocelli rounded and of same size. Transfacial distance equal to height of eye; toruli located above mid height of head, frons shorter than height of lower face, diameter of antennal torulus 1.7x as long as the distance between them, distance between torulus and eye slightly longer than the diameter of torulus. Lower face almost smooth, raised, central part below the toruli alutaceous, slightly pubescent. Clypeus impressed, smooth, straight ventrally, medially not incised, anterior tentorial pits distinct; epistomal sulcus and clypeo-pleurostomal line present. Frons uniformly alutaceous, without striae, with few setae and no piliferous points; interocellar area alutaceous. Vertex and occiput alutaceous to coriaceous.

Antenna (Fig. 1d). Longer than head + mesosoma but shorter than body, with 12 flagellomeres; pedicel around 1.3x longer than broad; pedicel slightly longer than broad, flagellomeres not broadening apically; F1 1.2x as long as F2 and 1.2x as long as scape + pedicel; F3 1.1x as long as

F4, F5 slightly longer than F6, F7=F8, F9=F10=F11, F12 1.6x as long as F11. Placodeal sensilla present on F4–F12, inconspicuous.

Mesosoma (Fig. 2 a–c) 1.25x as long as high, concave in lateral view, pubescent. Sides of pronotum alutaceous to coriaceous, with white setae and weak, without carinae on upper latero-posterior part, anterolateral rim of pronotum visible. Mesoscutum. 1.1x as broad as long in dorsal view; weakly coriaceous, without rugae, pubescent, with piliferous points more visible in the posterior half between notauli. Notaulus complete, deep, converging posteriorly, at posterior end, distance between notauli shorter than distance between notaulus and mesoscutal side; median mesoscutal line absent; parapsidal and anterior parallel lines distinct, smooth, darker than the rest of mesosoma. Mesoscutellum as long as broad, rugose centrally with more prominent rugae on lateral and posterior margins; interspaces weakly coriaceous; overhanging metanotum, with sparse long setae. Mesoscutellar foveae oval, delimited posteriorly by a weak carina, deep, with weakly alutaceous to smooth bottom, separated by a median triangular area. Circumscutellar carina absent. Mesopleuron smooth, with some piliferous points and sparse setae, speculum smooth and shining; mesopleural triangle alutaceous, with dense white setae; axillar carinae present, axillar area smooth and axillula strongly pubescent, subaxillular bar triangular to rectangular, not projected dorsally; metapleural sulcus reaching mesopleuron at mid-height. Metascutellum subrectangular, coriaceous with some weak longitudinal carina, convex ventrally; metanotal trough alutaceous, with dense setae; ventral bar of metanotal trough weakly sculptured; central propodeal area nearly smooth and glabrous; lateral propodeal carinae parallel, slightly converging near nucha, medial propodeal area smooth and shining; lateral propodeal areas uniformly alutaceous, densely pubescent. Nucha almost smooth and shining dorsally, with parallel lateral sulci.

Legs. Hind coxa broad, hind femur strongly broadened, with apical lobe; hind tibia with a weak ventral carina on the ventral part extending to half the length of the tibia; base of tarsal claws with strong tooth.

Forewing. (Fig. 3a). Hyaline 1.2x as long as body, pubescent, with marginal cilia; radial cell open, around 3.4x as long as broad; areolet present. Rs+M vein poorly defined, directed toward middle of the basal vein.

Metasoma. (Fig. 2d–e). Shorter than head + mesosoma, longer than high. 2nd metasomal tergum occupying 3/4 of metasoma, with anterolateral patches of sparse white setae, without micropunctures, remaining terga uniformly smooth and glabrous. Prominent part of ventral spine of hypopygium needle-like, about 5.0x as long as broad, with sparse lateral setae, without apical tuft.

Gall. (Fig. 3c–3d). Spherical, unilocular, sessile, located on the main branches, solitary or sometimes in pairs, up to 4.0 mm in diameter, parenchymal tissue is narrow because the larval chamber occupies most of the gall, surface

covered with short bracts, brown when mature.

Biology. The new species induces galls on *Q. glaucooides* Mart & Gal. (section *Quercus*), and only the asexual form is known. Galls were collected between October and January and the adults emerged in the second half of January. After cutting the galls in March, live adults were observed inside, suggesting that emergence likely occurs from January to March. Galls detach easily and probably fall to the ground to complete development.

Distribution. Mexico, known only from Puebla State.

DISCUSSION

According to Cuesta-Porta *et al.* (2025), all species included in the genus *Femuros* produce unilocular and spherical galls, as also occurs in the species described here. The gall of the new species is the smallest among all *Femuros* species: *F. ruidum* has the largest galls (up to 23 mm in diameter), followed by *F. repandae* (17 mm), *F. ocri* (12 mm), *F. bracteatus* (8.4 mm), and *F. minimum* (4.0 mm). The main structural difference in the gall of *F. minimum* is the reduction of the parenchymal tissue, although it is still present as in the rest of *Femuros* galls. Consequently, the adult body size of *F. minimum* falls within the range of the other *Femuros* species *F. ruidum* (3.0–4.3 mm), *F. repandae* (3.2–4.0 mm), *F. ocri* (3.0–4.5 mm), *F. bracteatus* (2.1–3.5 mm), and *F. minimum* (2.6–3.6 mm). The new species falls into couplet 8 of the identification key to Feron species published in Cuesta-Porta *et al.* (2025) along with *F. ruidum* and *F. ocri*. We propose the following update to the identification key:

8. Mesopleuron densely pubescent with visible punctures and deep piliferous points, speculum mostly pubescent except for a small central glabrous smooth area (Fig. 22a); propodeal carinae bent outwards (Fig. 22c) *F. ruidum*
- Mesopleuron sparsely pubescent, mostly smooth with scattered delicate shallow piliferous points; speculum smooth and glabrous at least on median 1/2 length of speculum (Fig. 16c); propodeal carinae parallel (Fig. 17c) 9
9. Antenna with 11 flagellomeres, F11 sometimes with a faint incomplete suture; ventral spine of hypopygium 3.5–4.0x, reddish-brown body coloration; without black marks along anterior parallel lines and parapsidal lines; and ventral margin of clypeus rounded *F. ocri*
- Antenna with 12 flagellomeres, F12 not partially divided; ventral spine of hypopygium 5.0x as long as broad in ventral view; dark brown body color; anterior parallel lines and parapsidal lines black; and ventral margin of clypeus straight *F. minimum* sp. nov.

Following this study, the genus *Femuros* is now represented by four species in Mexico, while *F. bracteradus* is the only species known in the United States (Arizona). The adults of the new species differ from other *Femuros* species by the number of flagellomeres (see Diagnosis) and is morphologically most similar to *F. ocri* (see Diagnosis).

Both species mentioned have smaller galls compared with other *Femuros*, but the galls of the new species are clearly smaller than those of *F. ocri*.

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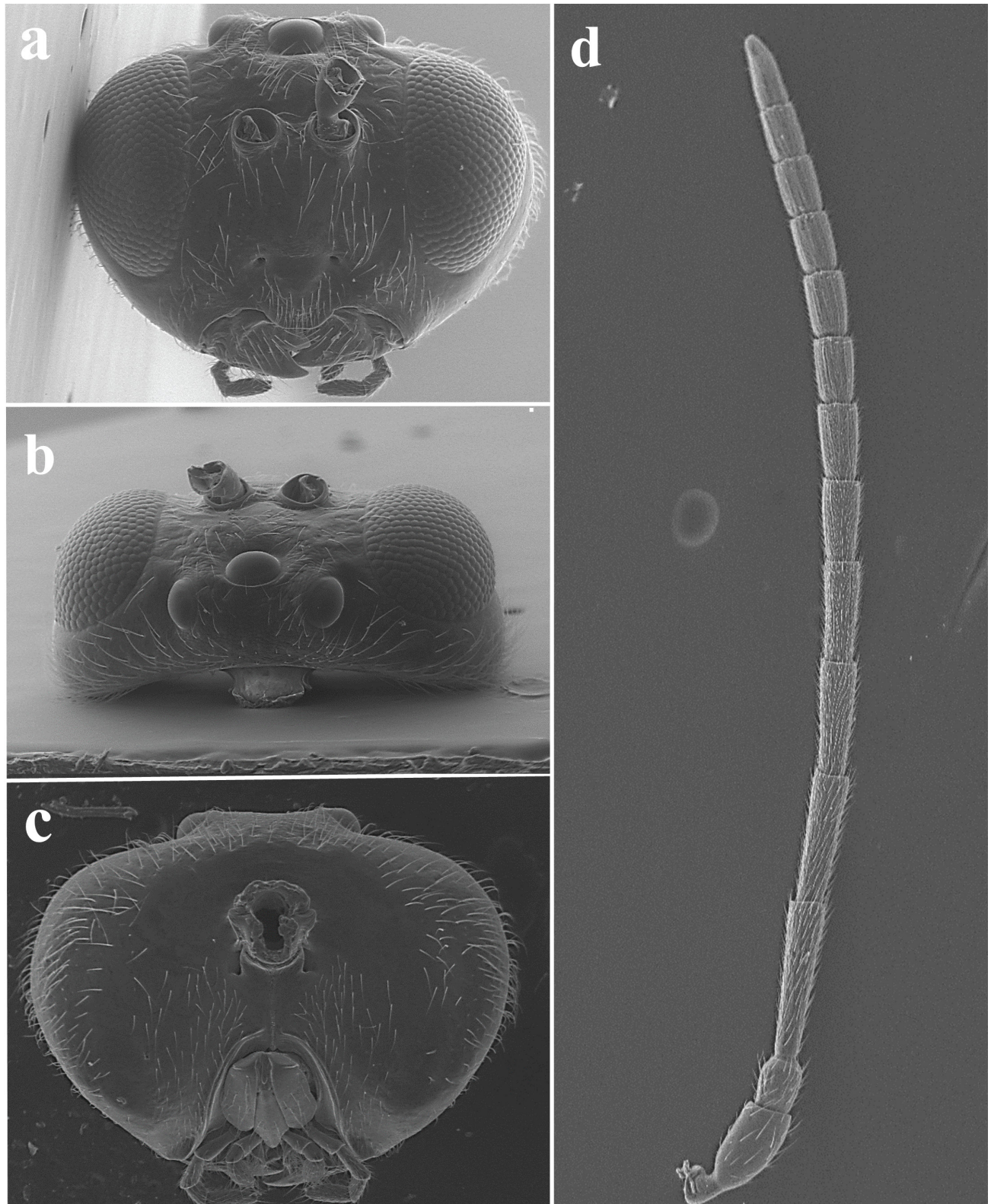


Figure 1. *Femuros minimum* sp. nov.: (a) head in frontal view, (b) head in dorsal view, (c) head in posterior view, (d) antenna.

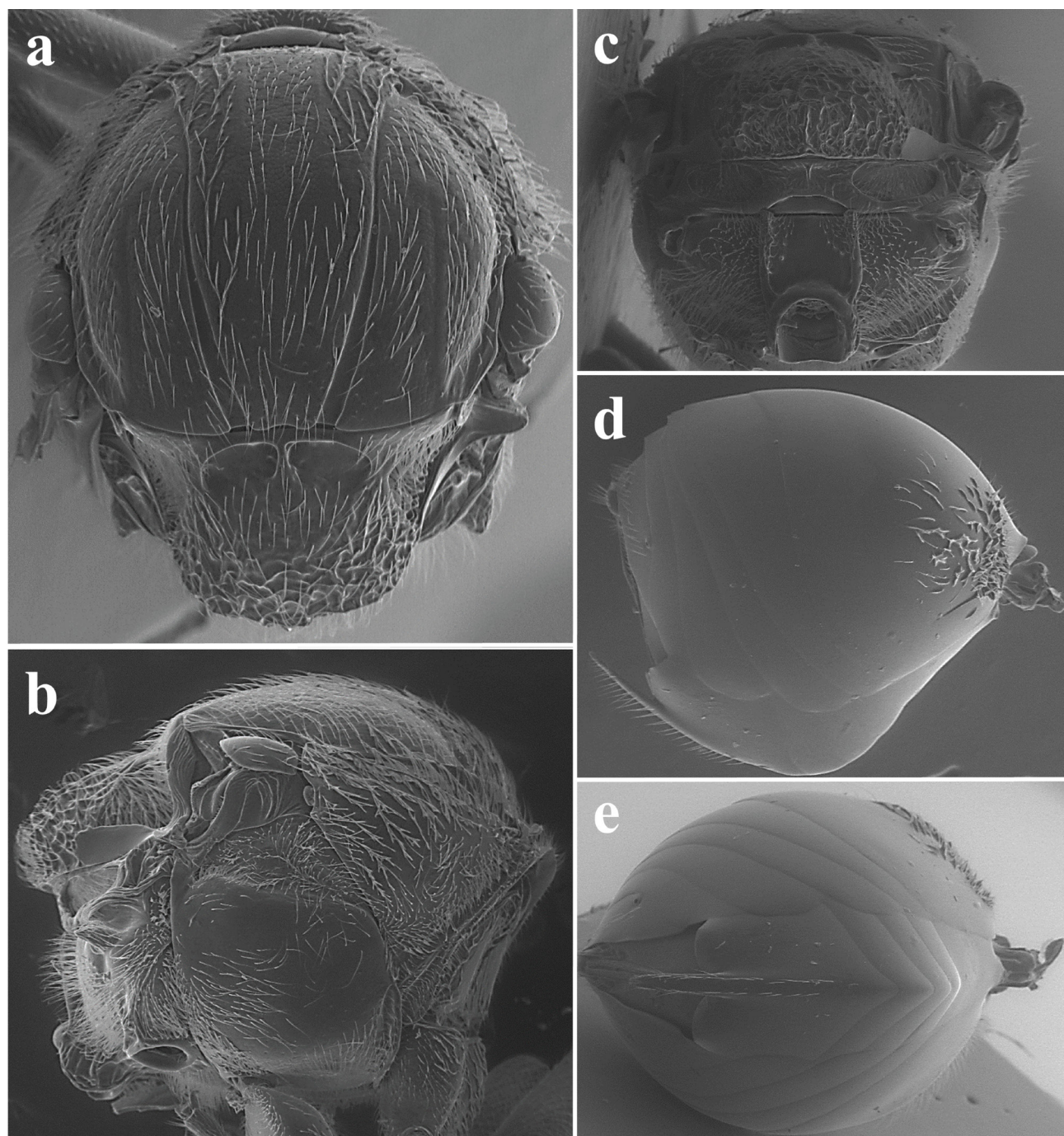


Figure 2. *Femuros minimum* sp. nov.: (a) mesosoma in dorsal view, (b) mesosoma in lateral view, (c) propodeum, (d) metasoma in lateral view, e) metasoma in ventral view.

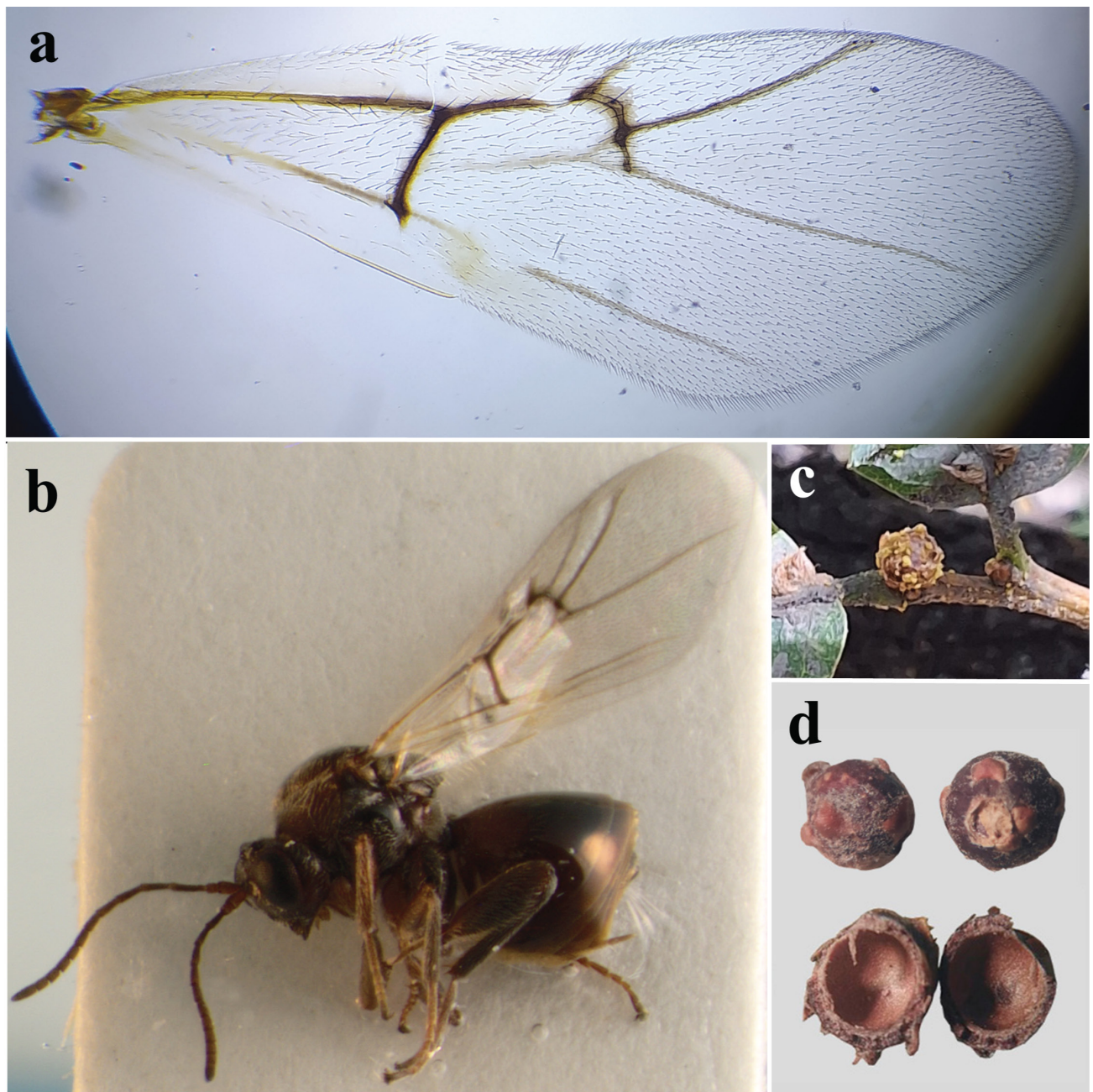


Figure 3. *Femuros minimum* sp. nov.: (a) fore wing, (b) habitus, (c) gall, (d) isolated galls and transverse cut of gall.

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- Los nombres científicos siempre deberán escribirse en cursivas o itálicas; nunca subrayados. La primera vez que se mencionen deben ir seguidos por el autor (es) que describió (eron) la especie y el año de descripción, separados por una coma; por ejemplo: *Oxyporus lawrencei* Campbell, 1974. Este formato debe incluirse también en el Resumen y Abstract. En caso de citar nombres regulados por un código de nomenclatura diferente al de Zoología, deberá colocarse entre paréntesis al menos el nombre de la familia a la que pertenece el taxón. El apellido del descriptor debe estar escrito completo.
- Al final del artículo y en página aparte, se incluirá la sección para los pies de figura y los cuadros, con sus respectivos encabezados. Para facilitar la edición de los mismos, nunca utilice imágenes de los cuadros, siempre elaborarlos con texto con las herramientas del procesador Word.
- Las figuras, mapas o fotografías serán presentadas por separado del texto, en formato TIFF con una resolución de 300 dpi o mayor. Todas deben numerarse de manera continua como figuras, de acuerdo con su señalamiento en el cuerpo del manuscrito. En caso de existir varias, es indispensable organizarlas en láminas. Se pueden enviar imágenes a color, siempre y cuando sean de alta resolución (600 dpi), con muy buena definición y que su publicación a color se considere indispensable. Las imágenes que no reúnan estos requisitos no se publicarán a color. Todas deben incorporarse a la plataforma de la revista. Para su evaluación, es necesario enviar un archivo pdf con todas las imágenes.
- En las imágenes y cuadros, en caso de haber sido tomados o modificados de otra fuente, mencionar el autor de los mismos o aclarar que son de autoría propia cuando fueron realizados por el o los autores.

- Agradecimientos: Además de los nombres de personas e instituciones que apoyaron aspectos del trabajo, pueden incluirse créditos a proyectos, programas, becas u otros datos pertinentes al trabajo o al (los) autor (es). Para ello, mencione el nombre de la institución u organización que dio el apoyo, agregue nombre y/o número del proyecto o contrato.
- Literatura citada: la cita en texto se incluye sin comas entre autor y año ejemplo: (López 1980), (López y Hernández 1980). Es oportuno recordar que de esta forma, se diferenciará entre las citas bibliográficas y la referencia al descriptor y año de descripción de un taxón. No utilizar el operador “&” (ampersand); no incluir sangrías, justificación o numeración. En la lista, todas las referencias se deben organizar en orden alfabético. Cuando se mencionen varios artículos del mismo autor, éstos se presentarán en orden alfabético, cronológico y por número de autores. Si hay dos artículos con el (los) mismo (s) autor (es) y año, deberán diferenciarse por las letras ‘a’, ‘b’, ... En cursivas deberán escribirse los títulos de libro, nombre de revista y título de tesis (en lo posible evitar citas de tesis y resúmenes de eventos académicos); no se permiten citas de páginas de internet, pero pueden incluirse referencias a publicaciones electrónicas o digitales, bases de datos o software. Las conjunciones de los autores deben escribirse en el idioma original de la obra citada. Ejemplo: y, and, et, und.

Ejemplos:

Libros [sin páginas totales]:

Morón, M.A., B.C. Rattclife y C. Deloya (Eds). 1997. *Atlas de escarabajos de México: Coleoptera: Lamellicornia, Vol. I Familia Melolonthidae*. CONABIO-SME, México, D.F.

Coloque la ciudad correspondiente en función de la fecha de la obra. Algunas ciudades de edición pueden cambiar con el tiempo. Es el caso de la Ciudad de México.

Capítulos de libro [los nombres de los editores ordenados de manera similar que los nombres de los autores del capítulo]:

Edmunds, G.F. and D. Waltz. 1995. Ephemeroptera. (pp. 126-163). In: Merritt, R.W. and K.W. Cummins (Eds.). *An Introduction to the Aquatic Insects of North America*. Kendall-Hunt, Dubuque.

Artículos [El nombre de la revista debe escribirse completo. Es indispensable incluir el número de la revista cuando éste existe. Para el caso particular de Folia Entomológica Mexicana, Acta Zoológica Mexicana, Zootaxa y, en general, para aquellas publicaciones que durante un tiempo utilizaron o siguen utilizando sólo el

número (excluyendo el volumen), coloque el número de la revista entre paréntesis]:

Fitzgerald, T.D., A. Pescador-Rubio, M.T. Turna and J.T. Costa. 2004. Trail marking and processionary behavior of the larvae of the weevil *Phelypera distigma* (Coleoptera: Curculionidae). *Journal of Insect Behavior*, 17(5): 627-646.

Huerta, C. y G. Halfpiter. 2000. Factores involucrados en el comportamiento subsocial de *Copris* (Coleoptera: Scarabaeidae: Scarabaeinae). *Folia Entomológica Mexicana*, (108): 95-120.

Kohlmann, B. and A. Solís. 2006. New species of dung beetles (Coleoptera: Scarabaeidae: Scarabaeinae) from Mexico and Costa Rica. *Zootaxa*, (1302): 61-68.

Tesis [Evitar en lo posible este tipo de referencias]:

Contreras-Ramos, A. 1990. *The immature stages of Platyneuromus (Corydalidae) with a key to the genera of larval Megaloptera of Mexico*. M. Sc. Thesis, University of Alabama, Tuscaloosa.

Memorias de congresos, simposios y otras reuniones [Evitar en lo posible este tipo de referencias. En caso de ser necesario contactar al editor].

Publicaciones y sitios web, bases de datos y Software (debe incluirse la dirección electrónica y la fecha de consulta):

Oksanen, J., F. Guillaume Blanchet, R. Kindt, P. Legendre, R. B. O'Hara, G. L. Simpson, P. Solymos, M. H. H. Stevens and H. Wagner 2011. vegan: Community Ecology Package. R package version 1.17-8. <http://www.rproject.org/>. Fecha de consulta: 12 de diciembre de 2016.

Linbos. 2014. Los insectos del bosque seco. <http://1.linbos.net/>. Fecha de consulta: 12 de diciembre de 2016.

Sistema Meteorológico Nacional. 2016. Información climatológica. <http://smn.cna.gob.mx/es/climatologia/informacion-climatologica>. Fecha de consulta: 12 de diciembre de 2016.

Steinkraus, D. 2004. Strange facts about soldier beetles infected with the poorly known fungal pathogen, *Erynnopsis lampyridarum*. Papers of the 2004 Entomological Society of America Annual Meeting and Exhibition. Disponible en: https://esa.confex.com/esa/2004/techprogram/paper_17245.htm. Fecha de consulta: 12 de diciembre de 2016.

No existen sobretiros impresos, solo digitales, los cuales pueden descargarse desde la página de la revista: <http://148.202.248.171/dugesiana/index.php/DUG/issue/archive>.

Descripción de taxones

Se debe adaptar a las características de un artículo. La descripción del taxón debe contener los siguientes elementos: nombre, diagnóstico, descripción, material tipo, etimología, biología, distribución y comentarios taxonómicos. Para los trabajos de Taxonomía deben tomarse en cuenta las consideraciones del Código Internacional de Nomenclatura Zoológica cuarta edición (1999).

Taxonomía y técnicas de estudio

Los encabezados del manuscrito deberán incluir: Resumen, Abstract (inglés), Introducción, Tratamiento taxonómico, Discusión, Agradecimientos y Literatura citada. En caso de considerar necesaria la inclusión de otras secciones, éstas se pueden incluir respetando: encabezados escribir con mayúsculas y centrados en el texto; subtítulos con mayúsculas y minúsculas y con sangría. Este tipo de contribuciones debe ser un aporte detallado al estudio de un taxón particular. Los encabezados quedan a juicio del autor (es), pero es recomendable que estén acompañadas de ilustraciones. En estas secciones se incluyen aquellos trabajos de tipo catálogo, inventarios, descripción o redescritión de especies, claves, etc. Los manuscritos sobre grupos particulares (ejemplo: Odonata, Coleoptera, entre otros) deben mencionar aspectos sobre la biología del grupo, técnicas de estudio (en campo y gabinete), así como claves dicotómicas, mínimo para nivel de familia. Se recomienda la inclusión de figuras en las claves las cuales deben organizarse en láminas. Los trabajos deben ser originales y enfocarse principalmente a México o la región Neotropical.

Ensayo

Los encabezados del texto de un ensayo deberán incluir: Resumen, Abstract (inglés), Introducción, Discusión, Conclusiones y/o sugerencias, Agradecimientos y Literatura citada. En caso de considerar necesaria la inclusión de otras secciones, éstas se pueden incluir respetando: encabezados escribir con mayúsculas y centrados en el texto; subtítulos con mayúsculas y minúsculas y con sangría. Los ensayos deben ser trabajos analíticos y con propuestas o posiciones claras de parte del autor (es).

Nota Científica

No se aceptan notas científicas.

Reseña bibliográfica

No se aceptan reseñas bibliográficas.

Para mayores detalles, contactar a:

Editor

Dr. José Luis Navarrete-Heredia, Centro de Estudios en Zoología, CUCBA, Universidad de Guadalajara, Apdo. Postal 134, 45100, Zapopan, Jalisco, México,

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Asistente editorial

M. en C. Ana Laura González-Hernández, Centro de Estudios en Zoología, CUCBA, Universidad de Guadalajara, Apdo. Postal 134, 45100, Zapopan, Jalisco, México, alaura.gonzalez@academicos.udg.mx

The received manuscripts for their assessment and possible publishing in *Dugesiana* are revised at least by two anonymous specialists who are familiar with the area of study and usually not part of the editorial committee. It is essential that the reference of three specialists is sent by the author as well as their e-mail addresses since they are considered when assessing. Add a file with the list of the names.

All manuscripts will be evaluated in the Turnitin System to verify their originality.

In order to have reviewed the work to be published in *Dugesiana*, the manuscript has to follow the next publishing rules:

- The manuscript is to be written in Word, double space, with a margin of 2.5 cm and indentation of 0.5 cm.
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- Written with Times New Roman format, size 12.
- Numbered pages. Add page number below and centered.
- Manuscripts in Spanish, English and Portuguese are accepted.

Text should contain the following aspects: original language title, title in Spanish, abstract, key words, abstract in English, introduction, material, as well as methods. Results, a discussion, acknowledgments and references to quoted literature (as literature cited). Headline should be written in capitals, bold and centered. Manuscripts in Spanish, English and Portuguese are preferred. Should the manuscript be in Spanish or Portuguese, then it should contain a title in English.

- Short title as headline, no more than 16 words. Regarding scientific names, last name and year of description should be separated by a comma. It is important to add name of order and family in parenthesis. Every title should be written in two languages, where mainly English and Spanish will be used.
- Complete name of authors in capital and small letters, followed by the corresponding addresses related with superscript numbers as well as the corresponding author marked with an asterisk.
- Addresses or affiliations of authors should be indicated initially with the superscript number that corresponds to them. Mention the dependencies in hierarchical order from lower to higher degree. For example: Departamento de Zoología, Instituto de Biología, Universidad Nacional Autónoma de México. Do not abbreviate the names of institutions,

and add the city, state and country (in this order).

- Add ORCID number for each author.
- Abstract: paragraph that should not exceed 300 words, not include references.
- Keywords: up to eight words, many of which are included in the title of the paper.
- Resumen: Spanish version of abstract (must be a translation, never something different). Not exceed 300 words.
- Palabras clave: Spanish version of keywords, with the same indications.
- Subtitle headings in "Introduction, Material and Methods, Results, Discussion and Acknowledgments" sections will be written in initial capital letters and bold.
- Scientific names should be always be written in italics, but never underlined. The first time mentioned, they should be followed by the author/authors who described the specie and year of description, separated by a comma. e.g. "Oxyporus lawrencei Campbell, 1974." This format should also be included in the abstract. In case of quoting names regulated by a nomenclature code being different to the one used in zoology, the family name belonging to taxon should be written in parenthesis. The descriptor full name should be written as well.
- At the end of the article and in a separated page, the footnotes and tables, with their respective headings, will be included. For editing purposes, the tables should never be drawn as images, do it using the Word tools.
- Figures, maps, or photographs will be presented separately from the text in TIFF format with a resolution of 300 ppp or larger. All of them should be numbered continuously as figures, according to the manuscript body. In case of having several, it is essential to have them organized as plates. Colored pictures are accepted, provided that their resolution is high (600 ppp), as well as their definition; as long as their colored publishing is really essential. Pictures not accomplishing these requirements will not be published in color. All of them should be uploaded in the platform of the magazine. As for their assessment, it is necessary to send a pdf file including all the pictures.
- Regarding pictures and charts taken from another source, either the author should be mentioned or their authorship should be mentioned.

- Acknowledgments: besides the names of people and institutions that supported the work, credit can be given to projects, programs, scholarships as well as other data concerning the work as well as authors. In that case, you should include not only the name of the institution, that is, organization, but also the name and/or number of project, that is, contract.
- Literature cited: literature is to be written without commas between the author and year, e.g.: (López 1980), (López y Hernández 1980). It is worth remembering that in this way, quote and reference to the descriptor as well as a taxon's description can be differentiated. Do not use the operator "&" (ampersand); as well as indentation, justification or numeration. All references should be organized alphabetically. If several articles from the same author are to be mentioned; these will be presented in alphabetical, chronological order as well as by number of authors. In case there are two articles with the same author and year, they are to be differentiated by using the letters 'a', 'b'... Titles of book, magazine as well as the title of a thesis should be written in italics (quotes from thesis and congress or symposium reports should be avoided as much as possible). Website references are not allowed; nevertheless, references from electronic publishing, data base as well as software may be included. Conjunctions from the authors should be written in the original language, e.g.: y, and, et, und.

Examples:

Books [no total page number]:

Morón, M.A., B.C. Rattclife y C. Deloya (Eds). 1997. *Atlas de escarabajos de México: Coleoptera: Lamellicornia, Vol. I Familia Melolonthidae*. CONABIO-SME, México, D.F.

Write the corresponding city according to the date. Some cities might change through time, such as Mexico City.

Book chapters [the name of the editors organized in a similar way as the name of the chapter's authors]:

Edmunds, G.F. and D. Waltz. 1995. Ephemeroptera. (pp. 126-163). In: Merritt, R.W. and K.W. Cummins (Eds.). *An Introduction to the Aquatic Insects of North America*. Kendall-Hunt, Dubuque.

Articles [the journal's name should be written completely; not to forget that the number of journal should be written when it exists. As for Folia Entomologica Mexicana, Acta Zoologica Mexicana, Zootaxa as well as publishing only using the number (leaving out the volume), parenthesis should be for the number of the magazine.

Fitzgerald, T.D., A. Pescador-Rubio, M.T. Turna and J.T. Costa. 2004. Trail marking and processionary behavior of the larvae of the weevil *Phelypera distigma* (Coleoptera: Curculionidae). *Journal of Insect Behavior*, 17(5): 627- 646.

Huerta, C. y G. Halfpter. 2000. Factores involucrados en el comportamiento subsocial de *Copris* (Coleoptera: Scarabaeidae: Scarabaeinae). *Folia Entomologica Mexicana*, (108): 95-120.

Kohlmann, B. and A. Solís. 2006. New species of dung beetles (Coleoptera: Scarabaeidae: Scarabaeinae) from Mexico and Costa Rica. *Zootaxa*, (1302): 61-68.

Thesis [is recommended to avoid this type of references]:

Contreras-Ramos, A. 1990. *The immature stages of Platyneuromus (Corydalidae) with a key to the genera of larval Megaloptera of Mexico*. M. Sc. Thesis, University of Alabama, Tuscaloosa.

Congresses, symposia and other meetings reports [Avoid in this type of references. If is necessary to include this type of references please contact the editor].

Online publications and websites, database and software (e-mail address and search date should be included)

Oksanen, J., F. Guillaume Blanchet, R. Kindt, P. Legendre, R. B. O'Hara, G. L. Simpson, P. Solymos, M. H. H. Stevens and H. Wagner 2011. vegan: Community Ecology Package. R package version 1.17-8. <http://www.rproject.org/>. Search date: 12 de diciembre de 2016.

Linbos. 2014. Los insectos del bosque seco. <http://1.linbos.net/>. Search date: December 12th, 2016.

Sistema Meteorológico Nacional. 2016. Información climatológica. [http:// smn.cna.gob.mx/es/climatologia/informacion-climatologica](http://smn.cna.gob.mx/es/climatologia/informacion-climatologica). Search date: December 12th, 2016.

Steinkraus, D. 2004. Strange facts about soldier beetles infected with the poorly known fungal pathogen, *Erynnopsis lampyridarum*. Papers of the 2004 Entomological Society of America Annual Meeting and Exhibition. Available on: [https://esa.confex.com/esa/2004/techprogram/paper_17245. htm](https://esa.confex.com/esa/2004/techprogram/paper_17245.htm). Search date: December 12th, 2016.

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Taxa description

Adapted to the features of the article. It should include the following elements: name, diagnosis, description, type of material, etymology, biology, distribution, as well as taxonomic remarks. For these papers recommendations from “The International Code of Zoological Nomenclature”, fourth edition (1999) should be kept in mind.

Taxonomy and study methodology

Headlines should include: resumen, abstract (in English), introduction, taxonomical work, discussion, thank-you note, as well as references. In case of considering other sections, those can be included provided that the headlines are written in capital letters and centered; subtitles in capital and small letters as well as an indentation. This type of contributions will be a detailed study of a particular taxon. Regarding contributions for taxonomy and study methodology areas, headlines are up to the author's criteria; however, it is highly recommended to have pictures. Catalogs, inventories, description of species, among others should be included in this section. Manuscripts about some particular groups, such as Odonata, Coleoptera, among others, should mention aspects about the taxa biology, study methodologies (in field and others), as well as dichotomous keys, at least at a family level. It is highly recommended to include figures that should be organized as prints. The work should be original and focused mainly either on Mexico or the Neotropical region.

Essay

Headings should include: Resumen, Abstract, Introduction, Discussion, Conclusions and / or suggestions, Acknowledgments and Literature quoted. In case it is considered necessary to include other sections, these can be included respecting: headings written in capital letters and centered in the text; uppercase and lowercase captions and indented. The essays must be analytical works and with clear propositions or positions on the part of the author(s).

Scientific note

Not accepted.

Book reviews

Not accepted.

It is considered that the author(s) agree with publishing the results of the research in the journal “Dugesiana” and to testify that there is no conflict of interests, as well as to claim that it is an original version and it has not been sent to another magazine to be assessed.

For any other situation not related to the ones mentioned, please get in touch with the editor.

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